



## Correlation of Cooking duration and Phytonutrient release in Vegetables

Barkataki Manash P.<sup>1</sup>, Bhattacharya Mahua<sup>1</sup>, Bhoir Roshni<sup>1</sup>, Pillai Nisha<sup>1</sup>, J.R. Parvathi<sup>1</sup> and Lilwani Simran R.\*<sup>1</sup>

School of Biotechnology and Bioinformatics, D. Y. Patil University, Navi Mumbai, Maharashtra-400 614, India  
simravi123@yahoo.com

Available online at: [www.isca.in](http://www.isca.in), [www.isca.me](http://www.isca.me)

Received 10<sup>th</sup> March 2015, revised 31<sup>st</sup> March 2016, accepted 9<sup>th</sup> April 2016

### Abstract

The culinary arts across continents have provided a whole range of variant practices that now concentrate to contain the essential components of the nutriment used. Among the disparate healthy cooking practices, boiling is considered as a quick and effortless way for both preserving and extracting the water soluble vitamins. In this study, an attempt was carried out to correlate the extent of phytochemical release and antioxidant activity in raw and cooked food items. Common vegetables that form any part of appetizer, entrée or main course of food were targeted for this swot. Extracts obtained after 20 min of cooking showed increased phytochemical contents with corresponding free radical scavenging activity. Based on the targeted food samples an ideal cooking duration for each is recommended in accordance to their scavenging property.

**Keywords:** Polyphenols, Flavonoids, Antioxidant activity, *Lactuca sativa*, *Solanum lycopersicum*, *Brassica oleracea*.

### Introduction

A side track of modernization and advancements is an overgrowing need to satiate hunger contemplating every nutritious nuance of the given food. The very realization that “the groundwork of all happiness is good health” has led to the egregiousness of culinary arts. Apart from being the basic necessity, food and healthy food habits plays a significant role in curbing disease that range from cardiovascular diseases to cancer<sup>1-3</sup>. The “good effects” of this subsistence are majorly contributed by fruits and vegetables which are an excellent source of vitamins, minerals, nutrients and Phytochemicals<sup>4,5</sup>. The cooking methods are majorly categorized into water media based (steaming, boiling, poaching, stewing, braising); fat media based (roasting, frying); heat media based (grilling/ boiling, baking) and smoke media based (braising, roasting, grilling and smoking) of which the most preferred mode used for vegetables is boiling<sup>6</sup>. Phytonutrients or phytochemicals are mostly water soluble<sup>7</sup> and tend to leach out when vegetables are submerged in water for prolonged period<sup>8,9</sup>; their thermo labile nature also lead to the loss during exposure to heat<sup>10-12</sup>. Apart from these detriments, bactericidal activity enabling food consumption without harmful microbes, improved digestibility and increased availability of nutrients are beneficial factors credited to cooking<sup>13,14</sup>. Conversely, cooking vegetables increase the availability of antioxidants by enabling their release from trapped fibrous part of the vegetables<sup>15</sup>.

The present work aims at analyzing the effect of methods and cooking duration on the release of phytonutrients and antioxidants from 8 different vegetables belonging to *Lactuca*, *Brassica* and *Solanum* genus. The *modus operandi* was to compare pressure cooked approach and boiling approach of cooking against the raw usage of the same. The results of the

phytonutrient release were used to decide an ideal and optimum approach for consuming food.

### Materials and Methods

**Plant material:** Fresh high quality packaged samples of Lettuce Leaf (*Lactuca sativa* L. *crispa*), Lettuce Iceberg (*Lactuca sativa* L. *capitata*), Tomato (*Solanum lycopersicum*), Cherry Tomato (*Solanum lycopersicum* var. *cerasiforme*), Cabbage (*Brassica oleracea* var. *capitata* L. *alba*), Red Cabbage (*Brassica oleracea* var. *capitata* L. *rubra*), Cauliflower (*Brassica oleracea* var. *botrytis*) and Broccoli (*Brassica oleracea* var. *italica*) were purchased from Godrej Nature's Basket Gourmet retail store, Thane, Maharashtra, India in the month of March, 2012. These set of vegetables were selected owing to their availability throughout the year, usage in multi-cuisines, widespread consumption in raw as well as cooked forms. Another important criterion that was decisive in targeting these vegetable were their antioxidant potential<sup>16-25</sup>.

**Preparation of extracts:** Based on the framed methodology (Figure-1), two major types of extracts were used for this experiment; one being the raw extract and the other being the cooked extracts. Each of the collected vegetables were properly washed to remove dirt particles and dried for few minutes to remove traces of water. For raw extracts, 20 gm of edible portion of each vegetable was evenly chopped and dispensed into 200 mL of distilled water; after 6 hrs of soaking, the vegetable were crushed using a mortar and pestle, squeezed and filtered with muslin cloth (extract A). Three decoction were considered for cooked extract of which the first two were boiled and the last being pressure cooked. 20gm of edible portion of each vegetable was boiled in 200 mL of distilled water in an open vessel for 10mins (extract B) and 20mins (extract C);

whereas for the latter, the same quantity of vegetable was pressure cooker at high flame for 10 mins (extract D) in 200 mL distilled water. After cooling, the extracts were filtered using muslin cloth; the final concentration of the sample was made upto 200 mL with distilled water for further assay.

**Estimation of Water Soluble Polyphenolic and Flavanoid Contents:** Folin – Ciocalteu Method was employed for evaluating the polyphenol content<sup>26</sup> and aluminum chloride method was applied for flavanoid content<sup>27</sup>. For polyphenolic assay, to each of 20µL of 1% aqueous extracts (A, B, C and D), 1.58 mL of distilled water was dispensed followed by addition of 100µL Folin-Ciocalteu reagent. After thorough mixing, the solution was left undisturbed for few minutes; 300µL sodium carbonate solution was added and left for incubation at room temperature for 2 hrs. Sample absorbance was measured at 765 nm with gallic acid was used as a standard to obtain standard graph with the concentration range of 200-1000µg/mL. The results obtained were expressed in µg/mL of Gallic Acid Equivalent (GAE).

In the flavanoid estimation assay, 0.5 mL each of the 1% aqueous extracts (A, B, C and D) was mixed with 1.5 mL of 95% ethanol, 0.1 mL of 10% aluminum chloride, 0.1 mL of 1M potassium acetate and 2.8 mL of distilled water. After incubating at room temperature for 30 min, the absorbance of

the reaction mixture was measured at 415 nm. Quercetin in 80% ethanol was used as the standard to obtain a standard graph with the concentration range of 20-100µg/ml and the results were expressed in µg/ml of Quercetin Equivalent (QE).

**Antioxidant assay:** 1,1-Diphenyl 2-picrylhydrazyl-Radical Scavenging Activity (DPPH-RSA) method<sup>28</sup> was followed to estimate the radical scavenging activity of extracts (A, B, C and D) and the results were compared against the RSA activity of standard L-Ascorbic Acid (Vitamin C), a control sample. DPPH being free radical shows a diminution of its optimum absorption at 517nm by getting reduced with an antioxidant compound. 3mL of DPPH solution was added to 1mL of each extracts respectively and onto the standard solution of Vit C (10-100 µg/ml). Absorbance was taken after a time span 30 min at 517 nm; percentage inhibition activity was calculated using the following formula:

$$\frac{A_0 - A_1}{A_0} * 100$$

where:  $A_0$  is the absorbance of the control and  $A_1$  is the absorbance of extract/standard taken as Ascorbic acid. The results were expressed in percentage inhibition.

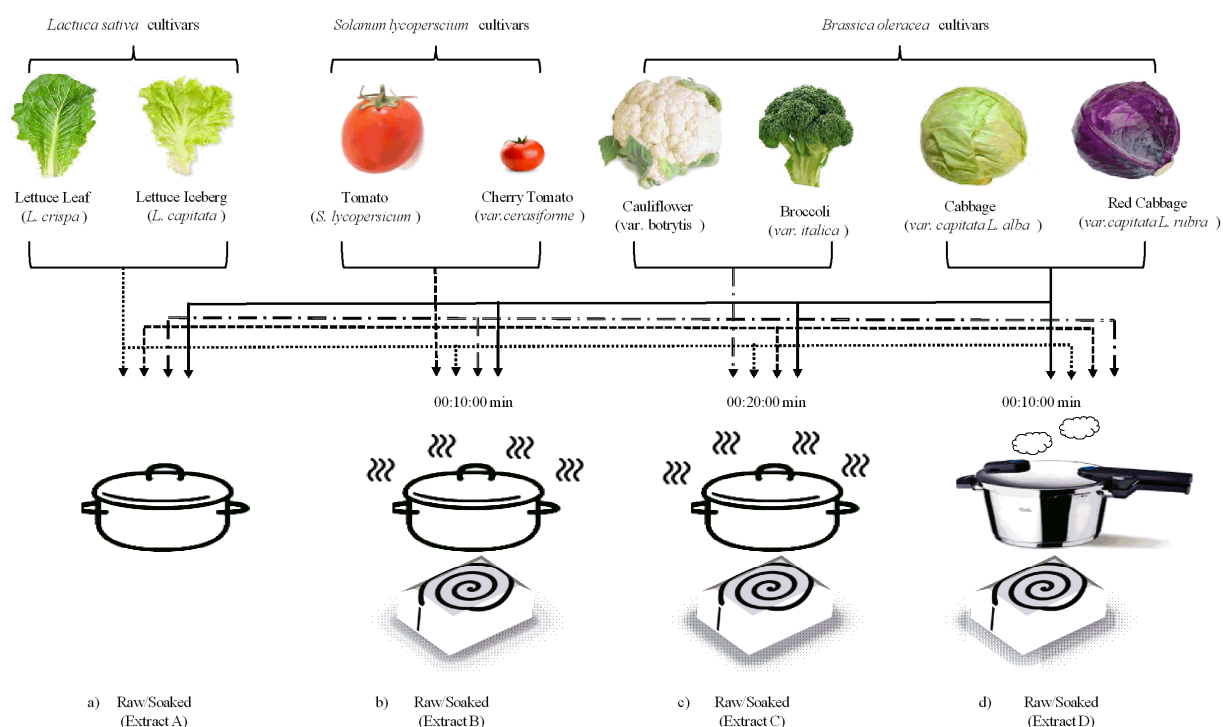


Figure-1

Cooking Trials a) Extract A: Raw; b) Extract B: 10 mins open vessel boiling; c) Extract C: 20 mins open vessel boiling; d) Extract D: Pressure Cooking for 10 mins

**Statistical Analysis:** One-way ANOVA method followed by Dunnett's multiple comparison test using Graph Pad Prism software (Version 6.03) was employed for statistical analysis; results were denoted as mean $\pm$ SEM. Values of  $p \leq 0.05$  were considered to be statistically significant.

## Results and Discussion

Polyphenols found ubiquitously in fruits and vegetables<sup>29</sup> have various health benefits providing anti-ageing, anti-inflammatory, anti-microbial and anti-cancer properties<sup>30-33</sup>. Among these, the most common being the flavonoids (especially catechins) are relatively less toxic in comparison to other active plant compounds (for instance alkaloids)<sup>34</sup>. This is suggestive of former being suitable for consumption among both animals and humans. Studies implicate flavonoids as biological "response modifiers" with anti-allergic<sup>42</sup>, anti-inflammatory<sup>43</sup> and antimicrobial capabilities<sup>44-46</sup>. Many of these phytochemicals also possess the antioxidant properties or "scavenger" effect that circumvents the negative impact of free radicals<sup>47-50</sup>.

*Lactuca sativa* L. *crispa* (lettuce leaf) displayed significant

release of polyphenols and flavonoids after 20 min of boiling; pressure cooked approach showed one third of this (Figure-2A and 2B). *Lactuca sativa* L. *capitata* (lettuce iceberg) showed similar amount of polyphenol release for both boiling and pressure cooked extracts whereas flavonoids were seen more in pressure cooked approach. The raw extracts of the same in comparison to *L. crispa* showed slightly more release of both the phytochemicals. These results could be complemented with highest antioxidant activity in extract C for both the vegetables. An unswerving correlation between the phytochemical release and antioxidant activity could not be deduced due to the values obtained (Figure-2C).

In *Solanum* cultivars, *S. lycopersicum* (tomato) showed maximum release of polyphenols after 10 min of boiling (Figure-3A) and that of flavonoids (Figure-3B) in pressure cooked method. For *S. lycopersicum* var. *cerasiforme* (cherry tomato), both polyphenol and flavanoid compounds were observed to be maximum at pressure cooked approach with 20 min boiling extracts succeeding it. The upper limit of antioxidant activities for both the cultivars were seen with extract B followed by extracts C and D (Figure-3C).

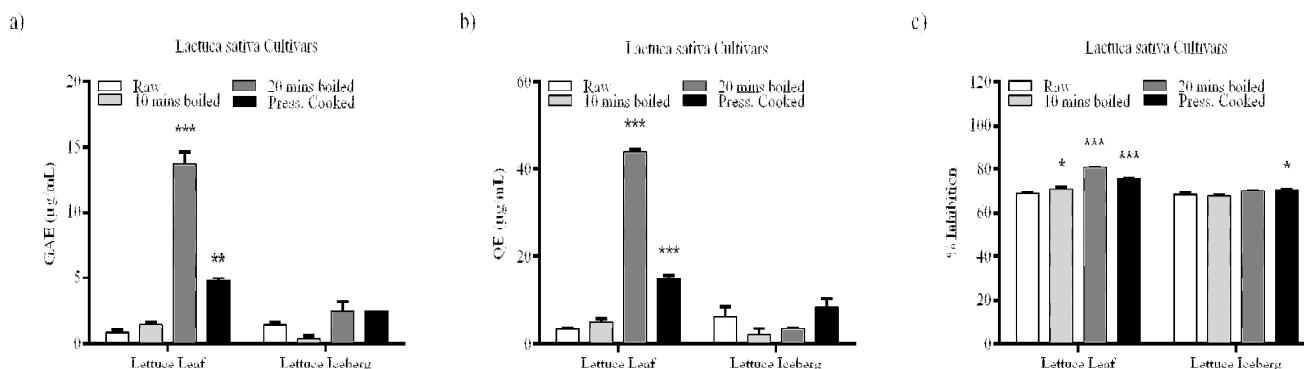


Figure-2

*Lactuca sativa* cultivars' phytonutrient release and antioxidant activity. a) Water Soluble Polyphenol Release; b) Water Soluble Flavonoid Release; c) Antioxidant Potential. Each bar represents Mean  $\pm$  SEM. \* $P \leq 0.05$ , \*\* $P < 0.01$  and \*\*\* $P < 0.001$ ; was considered significantly different compared to the raw group

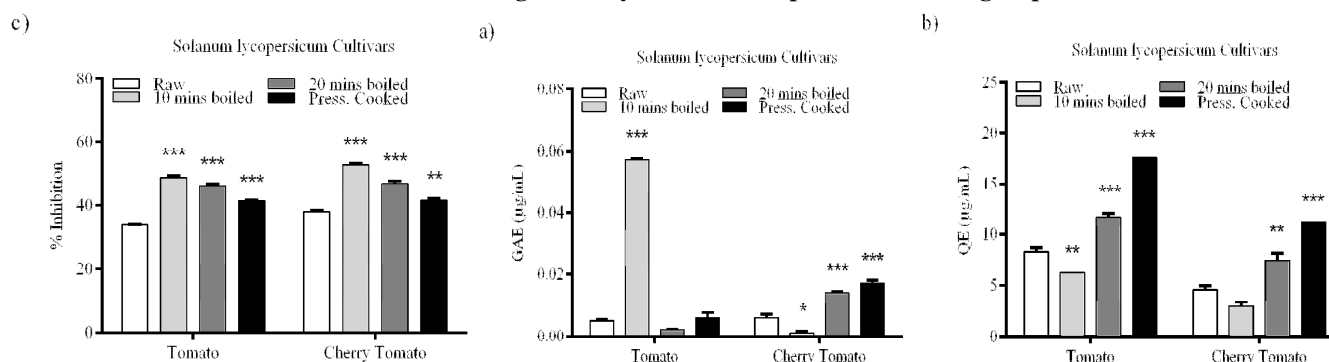


Figure-3

*Solanum lycopersicum* cultivars' phytonutrient release and antioxidant activity. a) Water Soluble Polyphenol Release; b) Water Soluble Flavonoid Release; c) Antioxidant Potential. Each bar represents Mean  $\pm$  SEM. \* $P \leq 0.05$ , \*\* $P < 0.01$  and \*\*\* $P < 0.001$ ; was considered significantly different compared to the raw group

In *B. oleraceavar. capitata* cultivars, both *L. alba* (cabbage) and *L. rubra* (red cabbage) gave maximum polyphenol release in extracts C and that of flavonoids in extracts D (Figure-4A and B). Releases of flavonoids were accounted more than polyphenols in both the cases; pressure cooked approach showed twice the flavonoids discharge than in 20 min of boiling. Red cabbage exhibited an increased release of polyphenols and flavonoids from raw to boiling (Figure-4B); whereas in cabbage the release of flavonoids were found to be more in 10 min of cooking than 20 min. All the cooking processes showed similar antioxidant values with slight increase in the extract D of red cabbage. Among the *Brassica oleracea* varieties, var. *botrytis* gave maximum release of polyphenols and flavonoids in extracts A (Figure-5A) with var. *italica* giving the similar results in extracts D (Figure-5B). Antioxidant values were similar in both the varieties showing 75% activity (Figure-5C).

## Conclusion

From the above results, considerable variation (Table-1) in the release of polyphenols and flavonoids could be correlated to the

cooking duration and nature and initial content of the phytochemicals<sup>16-25</sup>. Most of vegetables under study showed release of phytochemicals on cooking with exception to cauliflower. In this study flavonoids were shown to be less thermo liable than polyphenols with the former exhibiting higher values even after cooking. Common varieties of the selected vegetables (lettuce leaf, tomato, cabbage and cauliflower) showed a direct correlation between increase in antioxidant potential and cooking duration in contrast to their exotic counterparts (lettuce iceberg, cherry tomato, red cabbage and broccoli). Increased antioxidant activities in *Lactuca* and *Brassica* cultivars were linked to flavanoids whereas for *solanum* the same was linked to polyphenolic content. In case of the exotic varieties, phytonutrients like sulforaphane in broccoli<sup>51</sup>, high levels of lycopene in cherry tomato<sup>52</sup> and anthocyanin like isomers of cyanidin 3-O-(acyl) diglucoside-5-O-glucoside, cyanidin 3-O-(acyl1) (acyl2) diglucoside-5-O-glucoside, and cyanidin 3-O-(acyl1) (acyl2) diglucoside-5-O-(malonyl) glucoside in red cabbage<sup>23,53</sup> can be attributed to antioxidant potential of these vegetables apart from the targeted phytochemicals.

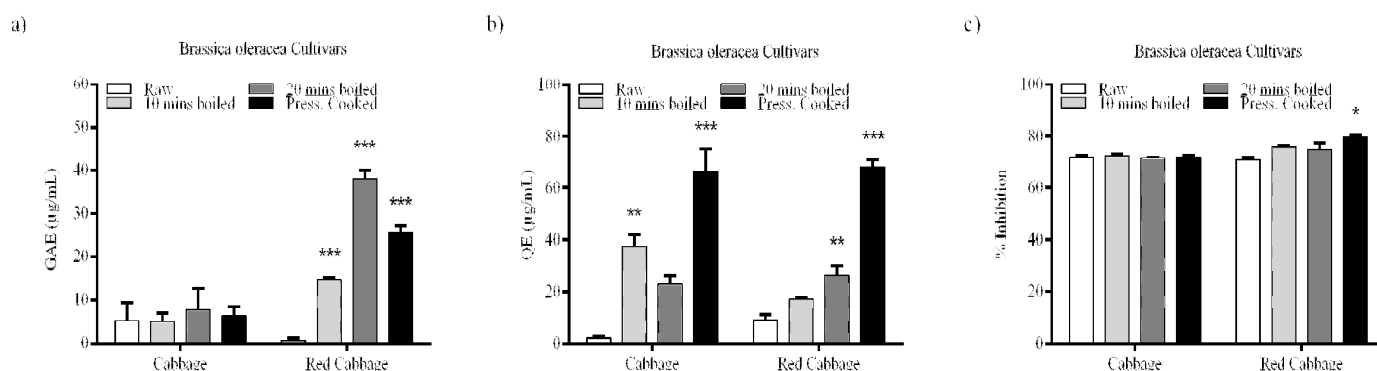


Figure-4

**Brassica oleracea cultivars' phytonutrient release and antioxidant activity. a) Water Soluble Polyphenol Release; b) Water Soluble Flavonoid Release; c) Antioxidant Potential. Each bar represents Mean ± SEM. \*P ≤ 0.05, \*\*P < 0.01 and \*\*\*P < 0.001; was considered significantly different compared to the raw group**

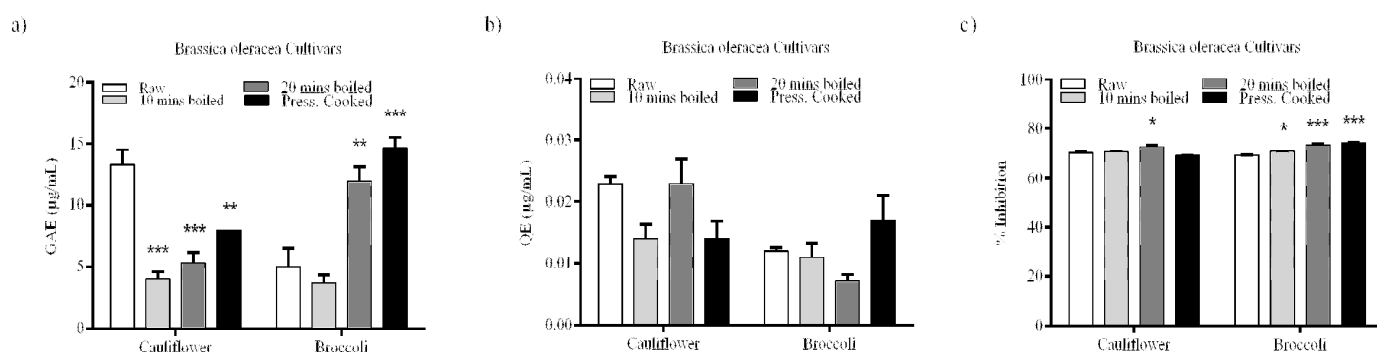


Figure-5

**Brassica oleracea cultivars' phytonutrient release and antioxidant activity. a) Water Soluble Polyphenol Release; b) Water Soluble Flavonoid Release; c) Antioxidant Potential. Each bar represents Mean ± SEM. \*P ≤ 0.05, \*\*P < 0.01 and \*\*\*P < 0.001; was considered significantly different compared to the raw group**

**Table-1**  
**Polyphenol, Flavanoid release and antioxidant values across the extracts of the selected vegetables**

Vegetables and its Cultivars Phyto-chemicals	<i>Lactuca sativa</i>		<i>Solanum lycopersicum</i>		<i>Brassica oleracea</i>			
	<i>L. crispa</i>	<i>L. capitata</i>	-	<i>cereas-iforme</i>	<i>L. alba</i>	<i>L. rubra</i>	<i>botrytis</i>	<i>italica</i>
	Lettuce Leaf	Lettuce Iceberg	Tomato	Cherry Tomato	Cabbage	Red Cabbage	Cauliflower	Broccoli
Polyphenols	C (~14µg/ml)	C/D (>5µg/ml)	B (0.06µg/ml)	D>C (~0.02µg/ml)	C (>10µg/ml)	C (~45µg/ml)	A (~14µg/ml)	D (~15µg/ml)
Flavanoids	C (~45µg/ml)	D (>5µg/ml)	D (~20µg/ml)	D (~15µg/ml)	D (>70µg/ml)	D (>70µg/ml)	A/C (>0.03µg/ml)	D (~0.02µg/ml)
Antioxidant activity	C (80%) >D>B>A (60-70%)	Same (~70%)	B(50%) C>D (40-50%)	B (55%) C>D (40-50%)	Same (~75%)	D (~80%) Rest (~75%)	C/B/A>D (~75%)	D/C>B/A (~75%)
Extract A (raw), Extract B (10 min boiling), C (20 min boiling), D (pressure cooked for 10 min). Polyphenol concentration is expressed in Gallic Acid Equivalent (GAE) and flavanoids are expressed in µg/ml of Quercetin Equivalent (QE).								

Boiling of *Solanum* cultivars for 10 min and *Brassica* and *Lactuca* vegetables for 20 min to is recommended as a healthier cooking practice to avail maximum benefit of their antioxidant potential.

## Acknowledgment

The authors thank Dr. Debjani Dasgupta, Prof. and Head, School of Biotechnology and Bioinformatics, D. Y. Patil University, Navi Mumbai for providing the necessary access to the infrastructure for successful completion of this project.

## References

- Liu R.H. (2003). Health benefits of fruit and vegetables are from additive and synergistic combinations of phytochemicals. *Am. J. Clin. Nutr.*, 78(3 Suppl), 517S–520S.
- Block G., Patterson, B. and Subar A. (1992). Fruit, vegetables, and cancer prevention: a review of the epidemiological evidence. *Nutr. Cancer*, 18(1), 1–29.
- Kaur C. and Kapoor H.C. (2008). Antioxidants in fruits and vegetables - the millennium's health. *Int. J. Food Sci. Technol.*, 36(7), 703–725.
- Hanif R., Iqbal Z. and Iqbal M. (2006). Use of vegetables as nutritional food: Role in human health. *J. Agric. Biol. Sci.*, 1(1), 18–22.
- Lintas C. (1992). Nutritional aspects of fruits and vegetable consumption, in *Options Mediterraennes*. 19.
- Andrews S. (2009). Food and Beverage Service- A Training Manual.
- Konings E.J.M. (2006). Water-soluble vitamins. *J. AOAC Int.*, 89(1), 285–8.
- Prodanov M., Sierra I. and Vidal-Valverde C. (2004). Influence of soaking and cooking on the thiamin, riboflavin and niacin contents of legumes. *Food Chem.*, 84(2), 271–277.
- Krehl W.A. and Winters R.W. (1950). Effect of cooking methods on retention of vitamins and minerals in vegetables. *J. Am. Diet. Assoc.*, 26(12), 966–72.
- Ford J.E., Hurrell R.F. and Finot P.A. (1983). Storage of milk powders under adverse conditions. *Br. J. Nutr.*, 49 355–364.
- Sieber R., Eberhard P., Fuchs D., Gallmann, P.U. and Strahm W. (2009). Effect of microwave heating on vitamins A, E, B1, B2 and B6 in milk. *J. Dairy Res.*, 63(01), 169
- Watanabe F. *et al.* (1998). Effects of microwave heating on the loss of vitamin B(12) in foods. *J. Agric. Food Chem.*, 46(1), 206–210.
- Kataria A. and Chauhan B.M. (1988). Contents and digestibility of carbohydrates of mung beans (*Vigna radiata* L.) as affected by domestic processing and cooking. *Plant Foods Hum. Nutr.*, 38(1), 51–59.
- Bishnoi S. and Khetarpaul N. (1993). Effect of domestic processing and cooking methods on in-vitro starch

- digestibility of different pea cultivars (*Pisum sativum*). *Food Chem.*, 47(2), 177–182.
15. Choi Y., Lee S.M., Chun J., Lee H.B. and Lee J. (2006). Influence of heat treatment on the antioxidant activities and polyphenolic compounds of Shiitake (*Lentinus edodes*) mushroom. *Food Chem.*, 99(2), 381–387.
16. Degl' Innoocenti E., Pardossi A., Tattini M. and Guidi L. (2008). Phenolic compounds and antioxidant power in minimally processed salad. *J. Food Biochem.*, 32(5), 642–653.
17. Souri E., Amin G., Farsam H. and Andaji S. (2004). The antioxidant activity of some commonly used vegetables in Iranian diet. *Fitoterapia*, 75(6), 585–8.
18. Xin Z. and Song K. (2004). Antioxidant activity of salad vegetables grown in Korea. *J. Food Sci. Nutr.*, 9(4), 289–294.
19. Jahan I.A. *et al.* (2010). Chemical and antioxidant properties of broccoli growing in Bangladesh. *Dhaka Univ. J. Pharm. Sci.*, 9(1), 31–37.
20. Riso P., Visioli F., Erba D., Testolin G. and Porrini M. (2004). Lycopene and vitamin C concentrations increase in plasma and lymphocytes after tomato intake. Effects on cellular antioxidant protection. *Eur. J. Clin. Nutr.*, 58(10), 1350–8.
21. Chandra H.M. and Ramalingam S. (2011). Antioxidant potentials of skin, pulp, and seed fractions of commercially important tomato cultivars. *Food Sci. Biotechnol.*, 20(1), 15–21.
22. Kaur C. and Kapoor H.C. (2002). Anti-oxidant activity and total phenolic content of some Asian vegetables. *Int. J. Food Sci. Technol.*, 37(2), 153–161.
23. Podsedek A., Sosnowska D., Redzynia M. and Anders B. (2006). Antioxidant capacity and content of Brassica oleracea dietary antioxidants. *Int. J. Food Sci. Technol.*, 41(Supplement 1), 49–58.
24. Halvorsen B.L. *et al.* (2002). Nutrient Requirements A Systematic Screening of Total Antioxidants in Dietary Plants 1. *J. Nutr.*, 132(September 2001), 461–471.
25. Lenucci M.S., Cadinu D., Taurino M., Piro G. and Dalessandro G. (2006). Antioxidant composition in cherry and high-pigment tomato cultivars. *J. Agric. Food Chem.*, 54(7), 2606–2613.
26. Ronald E. Wrolstad, Terry E. Acree, Haejung An, Eric A. Decker, Michael H. Penner, David S. Reid, Steven J. Schwartz, Charles F. Shoemaker, Denise M. Smith, Peter Sporns (2001). *Current Protocols in Food Analytical Chemistry Curr. Protoc.*, 1199, John Wiley and Sons, Inc., . doi:10.1002/0471142913
27. Chang C., Yang M.H., Wen H.M. and Chern J.C. (2002). Estimation of total flavonoid content in propolis by two complementary colorimetric methods. *J. food drug Anal.*, 10(3), 178–182.
28. Shanmugam S., T.S.K. and Selvam K.P. (2010). *Laboratory Handbook on Biochemistry* 141 (PHI Learning Pvt. Ltd., at <[http://books.google.co.in/books?id=gJveIjmEWc0Candprintsec=frontcoverandsource=gbs\\_ge\\_summary\\_randcad=0#v=onepageandqandf=false](http://books.google.co.in/books?id=gJveIjmEWc0Candprintsec=frontcoverandsource=gbs_ge_summary_randcad=0#v=onepageandqandf=false)>
29. Khanbabaee K. and Ree T. Van. (2001). Tannins: Classification and Definition. *R. Soc. Chem.*, 18 641–649.
30. Duthie G.G., Gardner P.T. and Kyle J. a M. (2003). Plant polyphenols: are they the new magic bullet?. *Proc. Nutr. Soc.*, 62(3), 599–603.
31. De la Lastra C.A. and Villegas I. (2005). Resveratrol as an anti-inflammatory and anti-aging agent: mechanisms and clinical implications. *Mol. Nutr. Food Res.*, 49(5), 405–30.
32. Taguri T., Tanaka T. and Kouno I. (2004). Antimicrobial Activity of 10 Different Plant Polyphenols against Bacteria Causing Food-Borne Disease. *Biol. Pharm. Bull.*, 27(12), 1965–1969.
33. Daglia M. (2012). Polyphenols as antimicrobial agents. *Curr. Opin. Biotechnol.*, 23(2), 174–81.
34. Cheeke P.R. (1989). *Toxicants of Plant Origin: Alkaloids 352*, Taylor and Francis. at <[International Science Community Association](http://books.google.co.in/books?hl=enandlr=andid=eASgQyXq8xMCandoi=fndandpg=PR1anddq=alkaloid+toxicityandots=pffkw-rMEJandsig=n6G0041y--sSzGBigXvd149cVM0></a></li>
<li>35. Chen X., Nishida H. and Konishi T. (2003). Baicalin promoted the repair of DNA single strand breakage caused by H<sub>2</sub>O<sub>2</sub> in cultured NIH3T3 fibroblasts. <i>Biol. Pharm. Bull.</i>, 26(2), 282–4.</li>
<li>36. Cummings J. and Smyth J.F. (1989). Flavone 8-acetic acid: our current understanding of its mechanism of action in solid tumours. <i>Cancer Chemother. Pharmacol.</i>, 24(5), 269–272.</li>
<li>37. Chauhan P.S., Satti N.K., Suri K.A., Amina M. and Bani S. (2010). Stimulatory effects of Cuminum cyminum and flavonoid glycoside on Cyclosporine-A and restraint stress induced immune-suppression in Swiss albino mice. <i>Chem. Biol. Interact.</i>, 185(1), 66–72</li>
<li>38. Schümann J. <i>et al.</i> (2003). Silibinin protects mice from T cell-dependent liver injury. <i>J. Hepatol.</i>, 39(3), 333–340.</li>
<li>39. Cushnie T.P.T. and Lamb A.J. (2005). Antimicrobial activity of flavonoids. <i>Int. J. Antimicrob. Agents</i>, 26(5), 343–56.</li>
<li>40. Yamamoto Y. and Gaynor R.B. (2001). Therapeutic potential of inhibition of the NF-kappaB pathway in the treatment of inflammation and cancer. <i>J. Clin. Invest.</i>,</li>
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- 107(2), 135–42.
41. Ayoola G. *et al.* (2009). Phytochemical screening and free radical scavenging activities of the fruits and leaves of *Allanblackia floribunda* Oliv (Guttiferae). *Int. J. Heal. Res.*, 1(2).
42. Kawai M. *et al.* (2007). Flavonoids and related compounds as anti-allergic substances. *Allergol. Int.*, 56(2), 113–23
43. García-Lafuente A., Guillamón E., Villares A., Rostagno M.A. and Martínez J.A. (2009). Flavonoids as anti-inflammatory agents: implications in cancer and cardiovascular disease. *Inflamm. Res.*, 58(9), 537–52.
44. Hodek P., Trefil P. and Stiborová M. (2002). Flavonoids-potent and versatile biologically active compounds interacting with cytochromes P450. *Chem. Biol. Interact.*, 139(1), 1–21.
45. Laks P.E. and Pruner M.S. (1989). Flavonoid biocides: Structure/activity relations of flavonoid phytoalexin analogues. *Phytochemistry*, 28(1), 87–91.
46. Harborne J.B. and Williams C.A. (2000). Advances in flavonoid research since 1992. *Phytochemistry*, 55(6), 481–504.
47. Halliwell B. (2009). Free radicals and antioxidants: A personal view. *Nutr. Rev.*, 52(8), 253–265.
48. Buettner G.R. (1993). The pecking order of free radicals and antioxidants: lipid peroxidation, alpha-tocopherol, and ascorbate. *Arch. Biochem. Biophys.*, 300(2), 535–43.
49. Craig W.J. (1997). Phytochemicals: Guardians of our Health. *J. Am. Diet. Assoc.*, 97(10), S199–S204.
50. Rao B.N. (2003). Bioactive phytochemicals in Indian foods and their potential in health promotion and disease prevention. *Asia Pac. J. Clin. Nutr.*, 12(1), 9–22.
51. Zhang Y., Talalay P., Cho C.G. and Posner G.H. (1992). A major inducer of anticarcinogenic protective enzymes from broccoli: isolation and elucidation of structure. *Proc. Natl. Acad. Sci. U. S. A.*, 89(6), 2399–403.
52. Raffo A. *et al.* (2002). Nutritional Value of Cherry Tomatoes (*Lycopersicon esculentum* Cv. Naomi F1) Harvested at Different Ripening Stages. *J. Agric. Food Chem.*, 50(22), 6550–6556.
53. Moreno D.A., Pérez-Balibrea S., Ferreres F., Gil-Izquierdo Á. and García-Viguera C. (2010). Acylated anthocyanins in broccoli sprouts. *Food Chem.*, 123(2), 358–363.