# Performances of Green Seaweed *Enteromorpha intestinalis*, Salt-marsh grass *Porteresia coarctata* and Mangrove litter as Prawn feed Ingredients

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#### **Abstract**

Three experimental feeds were formulated by reducing fishmeal at a level of 5%, incorporated with selected ingredients like green seaweed, Enteromorpha intestinalis (referred as ENT feed); salt- marsh grass, Porteresia coarctata (referred as POT feed) and mangrove litter (referred as ML feed). Juvenile prawn, Macrobrachium rosenbergii were exposed to the feeds of floral origin during the experimental trial and performances were evaluated in terms of growth characteristics and muscle protein content. Incorporation of E. intestinalis in the formulated feed resulted in significantly higher (p<0.01) average body weight (80.48 gm), condition factor (1.30), specific growth rate (1.45% day<sup>-1</sup>), feed conversion ratio (1.76) and survival (68.19%) as confirmed through ANOVA. Length-weight relationship varied significantly (p<0.01) thus exhibiting isometric growth (p= 3.008) in prawns treated with ENT feed as compared to others where allometric growth pattern was observed. Muscle protein content also reflected similar pattern with significantly greater (p<0.01) in the group of prawns treated with ENT feed. The present study thus confirms the suitability of the particular seaweed species for its incorporation in the formulated prawn feed.

Keywords: Floral ingredients, macrobrachium rosenbergii, growth characteristics, muscle protein content.

## Introduction

Feeds supplemented with ingredients of floral origin is currently gaining momentum in the area of organic feed formulation research perhaps to replace ingredients of animal origin<sup>1,2</sup> in order to increase disease resistance<sup>3</sup> and egg production<sup>4</sup>. However in aquatic animal nutrition, fishmeal happens to be the primary source of dietary protein and lipid because of its biological value. Interrupted availability and high price is gradually making this component very limited for incorporation in fish feeds by the manufacturers. Researchers are on the track screening alternative protein sources of plant origin as partial or total fishmeal replacement with a major focus on different terrestrial plants as protein sources in feed. As far as marine plant resources are concerned, incorporation of seaweeds as feed supplement have been used for shrimps and prawns<sup>5-10</sup>, sea bass<sup>11</sup>, snake head<sup>12</sup> and rainbow trout<sup>13</sup>. Apart from seaweed inclusion, feeds incorporated with salt-marsh grass, lucerne grass and mangrove litter have been reported for Penaeus Macrobrachium rosenbergii, monodon. Metapenaeus monoceros and Cirrhinus mrigala<sup>14-17</sup>. Research findings reveal that feed incorporated with seaweed meal has resulted in improved performance, better feed efficiency, better pellet stability and improved animal product quality which may be attributed to the enormous genetic potential of these lower group of plants with a genome that is more than twice the size of yeast which enables them in serving many industries such as food, animal feed, cosmetics, pharmaceuticals and biofuels<sup>18</sup>. Literature review enlightened some interesting facts which may serve as a baseline information for the present work. Such as green seaweed is found to be a good source of dimethyl sulfonyl propionate (DMSP) which has been proven for its attractant property thereby providing improved growth performance and increased feed efficiency in shrimps<sup>19, 20</sup>, salt-marsh grass traditionally used as animal food and fodder<sup>21</sup> is found to contain chemical compounds like fatty acids, hydrocarbons, steroids, sterol ester, triacylglycerol, triterpenes, waxes<sup>22</sup> and finally mangrove litter (or foliage) where the presence of constituents like protein, polyphenol, tannin, potassium, phosphate and nitrogen<sup>23-26</sup> are reported. Thus the present experimental work is an attempt to evaluate the performances of these floral ingredients incorporated in *M. rosenbergii* feed, in terms of growth characteristics and muscle protein concentration.

# Methodology

**Experimental floral ingredients and feeds:** Live and healthy plant materials (*E. intestinalis*, *P. coarctata* and mangrove litter) were collected from the estuarine zone of Uttar gobindapur village, Kakdwip block of Sundarbans (21°52'35.7" N latitude and 88°11' 55.0" E longitude) during low tide condition. All the collected plant materials were washed in ambient water and then with freshwater to remove epiphytes and other extraneous matter, dried in hot air oven under 55°C to preserve the biochemical constituents and finally processed. The experimental feeds were formulated to meet the nutritional requirements of prawn<sup>27</sup> as given in table 1. The proximate composition of the floral ingredients as well as the formulated feeds were estimated by the following methods: Lowry for

Protein<sup>28</sup>, Dubois for Carbohydrate<sup>29</sup>, Soxhlet for Lipid<sup>30</sup> and AOAC<sup>31</sup> for ash and moisture contents.

**Feeding trial:** A feeding trial was run at Uttar gobindapur village, Kakdwip block of Sundarbans ( $21^{\circ}52'35.7''$  N latitude and  $88^{\circ}11'55.0''$  E longitude) in grow-out ponds for 240 days of experimental duration as shown in figure 1. The experimental facility consisted of triplicate ponds for each dietary treatment. Each pond was well connected to adjacent estuary so that possible hydrological variations affect all the ponds simultaneously. The hydrological condition ranged within the optimum requirements recommended for freshwater prawn culture as given in table 2. Prawn juveniles were procured from a local hatchery and acclimated to the pond conditions prior to feeding trial. Juveniles were stocked at a density of  $2 \text{ No/m}^2$  with average initial weight of  $2.5 \pm 0.14$  gm and fed twice daily at 0630 and 1800 hrs. The uneaten feed was checked at regular intervals.

**Growth characteristics of prawn:** Individual weights were taken at every 30 days interval throughout the trial. The following response variables were determined from the experimental ponds:

Fulton's condition equation was used to find out the condition factor<sup>32</sup> as follows:

$$K = \frac{\overline{w}}{(\overline{TL})^3} \times 10^3$$

Where K is the condition factor,  $\overline{w}$  is the average weight (g) and  $\overline{TL}$  is the average total length (cm)

Specific growth rate (SGR) was calculated using the conventional equation:

$$SGR = \frac{\ln w_f - \ln w_i}{t} \times 100$$

Where  $W_f$  and  $W_i$  are the average final and initial weight in time t.

Final average body weight (ABW), feed conversion ratio (FCR) and survival were estimated after harvest by drag netting and dewatering the pond finally as:

FCR= total feed intake/ total biomass gain

Survival= (number of fish harvested/number of fish stocked)  $\times 100$ 

Length-weight relationship (*LWR*) was calculated by using the conventional formula<sup>33</sup> as given below:

$$W = a.TL^b$$

Where W is fish body weight (g), TL is total length (cm), a is the proportionality constant and b is the isometric exponent. The parameters a and b were estimated by non-linear regression analysis.

Table- 1
Formula and proximate composition of the experimental feeds

Feed Ingredients	ML feed	ENT feed	POT feed
Fish meal	30	30	30
Soybean oil cake	11	11	11
Mustard oil cake	11	11	11
Rice polish	23	23	23
Wheat flour	16	16	16
Oyster shell dust	2	2	2
Shark oil	2	2	2
Mangrove litter	5	-	-
E. intestinalis	-	5	-
P. coarctata	=	-	5
Proximate composition (% dry matter)			
Protein	$34.73 \pm 1.60$	$34.64 \pm 1.20$	34.09 ± 1.30
Carbohydrate	$35.04 \pm 0.27$	$33.19 \pm 0.76$	$36.28 \pm 0.90$
Lipid	$5.3 \pm 0.46$	$6.12 \pm 0.49$	$6.6 \pm 0.32$
Ash	$10.4 \pm 0.31$	$10.6 \pm 0.12$	$12.4 \pm 0.23$
Moisture	$10.2 \pm 0.36$	$9.82 \pm 0.15$	$10.7 \pm 0.69$

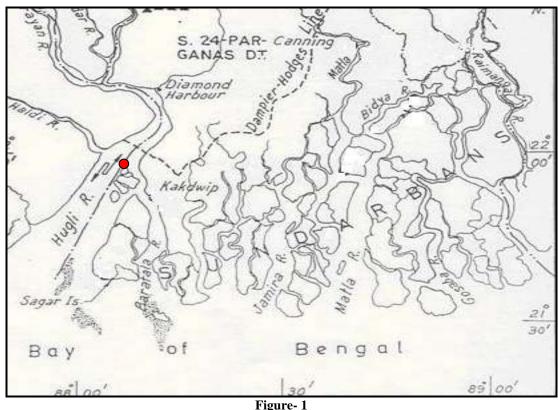
<sup>\*</sup>Results are mean ± SD

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Table- 2
Hydrological parameters of the experimental ponds treated with feeds of floral origin

Parameters	Ponds treated with ML feed	Ponds treated with ENT feed	Ponds treated with POT feed
Surface water temperature (°C)	$30.8 \pm 1.57$	$30.81 \pm 1.50$	$30.88 \pm 1.45$
Surface water salinity (psu)	$1.52 \pm 1.33$	$1.86 \pm 1.09$	$2.15 \pm 1.39$
pH	$7.47 \pm 0.24$	$7.76 \pm 0.11$	$7.70 \pm 0.15$
Dissolved oxygen (mg l <sup>-1</sup> )	$5.38 \pm 0.34$	$5.37 \pm 0.44$	$5.50 \pm 0.38$
Nitrate concentration (µg-at 1 <sup>-1</sup> )	$9.56 \pm 2.03$	$10.85 \pm 1.60$	$9.14 \pm 1.53$
Phosphate concentration (µg-at l <sup>-1</sup> )	$0.98 \pm 0.25$	$1.15 \pm 0.17$	$0.98 \pm 0.19$

<sup>\*</sup>Results are mean ± SD of three replicates



Map showing the *M. rosenbergii* culture site at Uttar gobindapur village, Kakdwip block of Sundarbans

Muscle protein concentration of prawn: Protein content was analyzed as per the standard Lowry's method. Protein present in the sample reacts with folin-ciocalteau reagent to give a coloured complex which is produced by the reduction of phospho-molybdate by tyrosine and tryptophan liberated from the protein by the action of alkaline copper. The intensity of the colour depends on the amount of protein present in the sample and is measured spectrophotometrically against standards and blank.

**Statistical analysis:** The collected data were finally subjected to one-way analysis of variance (ANOVA). All statistical calculations were performed with SPSS 9.0 for windows.

## **Results and Discussion**

**Proximate composition of the selected floral ingredients:** The proximate compositions of the floral ingredients like *E. intestinalis*, *P. coarctata* and mangrove litter as observed from the study region are provided in table 3. The protein content (% DW) in the selected samples varied as per the order, *E. intestinalis* > mangrove litter > *P. coarctata*. In case of carbohydrate content (% DW), the order is *P. coarctata* > mangrove litter > *E. intestinalis*. The lipid content (% DW) varied as per the sequence, mangrove litter > *E. intestinalis* > *P. coarctata*. The order of ash content (% DW) is *E. intestinalis* > *P. coarctata* > mangrove litter. The sequence of moisture content (% DW) is *P. coarctata* > mangrove litter > *E.* 

*intestinalis*. Earlier studies conducted shows that crude protein, crude lipid, total carbohydrate, ash and moisture contents in *E. intestinalis* ranged from 4-16%, 21-38%, 2-7%, 16-39% and 21-45% respectively<sup>34-40</sup>, while for *P. coarctata* these values varied from 5-12%, 35-50%, 0.9-2%, 13-23% and 42-68% <sup>41-44</sup> and for mangrove litter they ranged from 6-14%, 5-45%, 7-16%, 2-18% and 59-75% respectively<sup>45-49</sup>.

Table-3
Proximal composition of the experimental floral ingredients
(% dry matter)

Proximate composition	Mangrove litter	E. intestinalis	P. coarctata
Protein	$15.47 \pm 1.67$	$17.23 \pm 1.74$	$10.23 \pm 1.51$
Carbohydrate	$33.12 \pm 1.27$	$32.46 \pm 1.16$	$46.7 \pm 1.36$
Lipid	$9.02 \pm 0.62$	$4.67 \pm 1.10$	$1.26 \pm 0.35$
Ash	$9.36 \pm 1.03$	$25.43 \pm 1.28$	$17.53 \pm 1.02$
Moisture	$54.83 \pm 1.36$	$30.77 \pm 0.46$	$55.3 \pm 0.57$

<sup>\*</sup>Results are mean ± SD

**Proximate composition of the formulated feeds:** The feed composition is presented in table 1. The protein content ranged between  $34.09 \pm 1.30\%$  (POT feed) to  $34.73 \pm 1.60\%$  (ML feed), lipid content ranged between  $5.30 \pm 0.46\%$  (ML feed) to  $6.60 \pm 0.32\%$  (POT feed), carbohydrate content ranged between  $33.19 \pm 0.76\%$  (ENT feed) to  $36.28 \pm 0.90\%$  (POT feed), ash content ranged between  $10.4 \pm 0.31\%$  (ML feed) to  $12.40 \pm 0.23\%$  (POT feed) and moisture content ranged between  $9.82 \pm 0.15\%$  (ENT feed) to  $10.70 \pm 0.69\%$  (POT feed) respectively. The results obtained in the present study are in agreement with the values of several research findings<sup>50-53</sup>.

**Growth characteristics of prawn:** Prawns treated with ENT feed represented higher average body weight (80.48 gm), better condition factor (1.3±0.07), low FCR (1.76) and high SGR (1.45±0.13% day<sup>-1</sup>), the differences being statistically significant at 1% level as provided in table 4. At harvest, survival was 56.28%, 68.19% and 61.55% as recorded from respective treatments which also varied significantly (p<0.01).

Feeds of floral origin and its application in M. rosenbergii farming is the pioneer work from the present study region. A similar approach was undertaken by researchers through inclusion of floral ingredients in different regions of Indian subcontinent, but they were mostly related to replacement of fishmeal with terrestrial plant proteins. In this context, better weight gain, SGR and survival was observed in M. rosenbergii treated with feeds containing 7% Murraya koenigi extract<sup>54</sup>. Similar findings was obtained with significantly better growth performance and body pigmentation of P. monodon fed with formulated feed containing 5% of red seaweed, Catenella repens extract from Sundarbans. Inclusion of salt-marsh grass, P. coarctata in feeds of P. monodon at 100 g kg<sup>-1</sup> is also reported by workers $^{55}$ . Even better growth was obtained in M. rosenbergii through inclusion of E. intestinalis and P. coarctata in the feed. The inclusion of decomposed mangrove leaves or foliage in formulated feeds of Metapenaeus monoceros have been reported previously with better growth and conversion efficiency. However the present work reflected comparatively inferior growth performance in prawns applied with mangrove litter incorporated feed which may be attributed to the presence of certain organic or inorganic constituents in litter (such as polyphenols, tannins etc.) that might have contaminated the feed to certain extent resulting in reduced digestibility and inferior growth performance<sup>56-58</sup>. Supplementation of feeds using Enteromorpha sp. is also reported for Penaeus monodon, P. indicus, P. stylirostris, Litopenaeus vannamei and M. rosenbergii with better growth performance and FCR<sup>59-62</sup>. Superior growth performance of cultured species may be attributed to the formulated feed incorporated with E. intestinalis, although inclusion rate was low. Another probable reason could be the presence of compound dimethyl sulfonyl propionate (DMSP) in this particular order of seaweed (Ulvales) which has been proven as an attractant, thereby accelerating faster feed consumption and subsequent digestion by the culture species. The growth characteristics of prawn thus exhibited in the present study may be a direct consequence of inclusion of E. intestinalis in the feed which is under order Ulvales.

Table- 4
Variations in growth characteristics of prawn related to incorporation of floral ingredients in feed

Parameters	Prawns treated with ML feed	Prawns treated with ENT feed	Prawns treated with POT feed
Condition factor (K)	$1.14 \pm 0.11^{c}$	$1.30 \pm 0.14^{a}$	$1.19 \pm 0.14^{b}$
Survival (%)	56.28°	68.19 <sup>a</sup>	61.55 <sup>b</sup>
SGR (% day <sup>-1</sup> )	$1.18 \pm 0.99^{c}$	$1.28 \pm 1.16^{a}$	$1.21 \pm 1.08^{b}$
FCR	2.0ª	1.76°	1.88 <sup>b</sup>
ABW (gm)	61.76 <sup>c</sup>	80.47 <sup>a</sup>	66.16 <sup>b</sup>
LWR (b)	2.890°	3.008 <sup>a</sup>	2.931 <sup>b</sup>

<sup>\*</sup>Means with different superscripts in a row differ significantly (p<0.01); values are means of three replicates.

Growth of prawns depends on sex, stage and environmental factors such as nature of food, water temperature and salinity  $^{63}$ . LWR is an important indicator to measure the growth variations in a particular organism  $^{64}$  as well as for stock assessment  $^{65}$ . Easier techniques of calculating average weight at given length group is usually done by the method of LWR  $^{66,67}$ . The health conditions of a fish population  $^{68}$ ,  $^{69}$  is also assessed by this procedure. Variation in LWR obtained between the culture species may be attributed to the different feed composition, population density, environmental conditions and genetic make up of the species which has been well documented in earlier studies  $^{70.74}$ . A curvilinear growth pattern was observed irrespective of treatments with a gradual increase over the

feeding trial as shown in figures 2a, 2b and 2c. The isometric exponent (b) values showed significant variation (p<0.01) with the highest being obtained from the culture species fed with E. intestinalis incorporated feed as given in table 4. This value exactly coincides with the ideal condition of isometric growth pattern, when  $b=3.0^{75}$ . Difference in ingredient selection of feed clearly affected LWR of individual organisms, which coincides with the earlier observations that b value close or equal to 3.0 reflects better feed acceptance/ or environmental condition in a culture system. Hence the greater preference for ENT feed by the culture species would have affected this parameter.

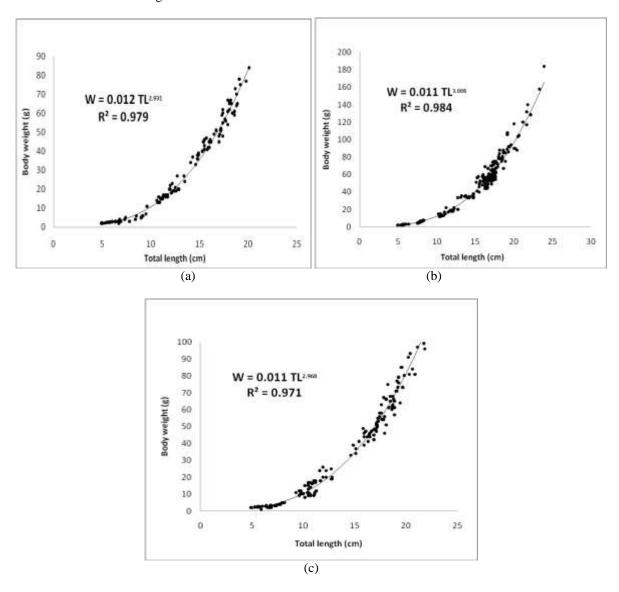


Figure- 2
Length-Weight Relationship curves of *M. rosenbergii* fed with different experimental feeds (a= ML feed, b= ENT feed, c= POT feed)

Muscle protein concentration of prawn: Protein is one of the major component which indicates shrimp/prawn meat edibility in terms of its amino acid profile. The present study highlights the highest protein content in the cultured species treated with E. intestinalis incorporated feed followed by the groups treated with P. coarctata and mangrove litter incorporated feeds as given in table 5. ANOVA results confirm significant variation (p<0.01) in muscle protein content of prawns which may be attributed to the influence of algal protein in the formulated feed that are consumed by the culture species and due to the nature of enzymes present in the digestive tract which are indicative that M. rosenbergii can digest protein of both plant and animal origin<sup>76-81</sup>. Higher protein concentration is directly linked with the translation mechanism occurring within the cells, which is the prime source of protein synthesis<sup>82</sup>. The values obtained were similar to the results as reported earlier where isonitrogenous feeds were prepared from fishmeal, acid silage and fermented fish silage in M. rosenbergii culture<sup>83</sup>. Similar observations were reported in the case of protein value while experimenting with some alternative practical feeds for M. rosenbergii<sup>84</sup>. Higher protein values were recorded in cultured prawn (M. rosenbergii)<sup>85</sup>, whereas, similar result was found in M. idella idella<sup>86</sup> where the protein ranged from 58.60 to 64.28% in males and 58.36 to 61.86% in females. The protein content varied between 58.17% to 64.15% in M. malcolmsonii<sup>87</sup>. The protein content was maximum in small sized males of M. scabriculum 59.15% and M. idae 61.44% 9. Juveniles of M. rosenbergii when exposed to formulated feeds containing different levels of cuttlefish liver lipid was also found to have muscle protein content between 64 - 66% which are in agreement to those reported in the present study where the feeds are incorporated with floral ingredients. Feed supplemented with 50% methanolic extract of Ricinus communis as feed additive when applied to groups of P. monodon exhibited muscle protein content of around 62%<sup>91</sup>. Thus the results are indicative that the feed protein is efficiently utilized for the increase in biomass by boosting up the growth of prawns.

> Table- 5 Effect of experimental feeds on muscle protein concentration of *M. rosenbergii*

,	Prawns treated with ML feed	Prawns treated with ENT feed	Prawns treated with POT feed
Protein (% dry weight)	64.17 ± 1.76°	68.78 ± 5.05 <sup>a</sup>	$65.92 \pm 2.75^{\text{b}}$

<sup>\*</sup>Means with different superscripts in a row differ significantly (p<0.01); values are means of three replicates.

## Conclusion

In the present study, *E. intestinalis* incorporated feed exhibited the best results in terms of growth characteristics and muscle protein content. The availability of this particular seaweed from the mangrove dominated Indian Sundarbans creates a viable opportunity for utilizing them in fish feed, which could be a step

towards organic feed formulation in future. The species can also be cultured through "rope culture technology", which is cost-effective. The present programme can thus open the avenue of alternative livelihood for the island dwellers of Indian Sundarbans.

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### References

- 1. Hansen H.R., Hector B.L. and Feldmann J., A qualitative and quantitative evaluation of the seaweed feed of North Ronaldsay sheep, *Animal Feed Science and Technology*, 105, 21–28 (2003)
- 2. Leupp J.L., Caton J.S., Soto-Navarro S.A. and Lardy G.P., Effects of cooked molasses blocks and fermentation extract or brown seaweed meal inclusion on intake, digestion, and microbial efficiency in steers fed low-quality hay, *Journal of Animal Science*, **83**, 2938–2945 (2005)
- **3.** Turner J.L., Dritz S.S., Higgins J.J. and Minton J.E., Effects of *Ascophyllum nodosum* extract on growth performance and immune function of young pigs challenged with *Salmonella typhimurium*, *Journal of Animal Science*, **80**, 1947–1953 (**2002**)
- **4.** Bratova K. and Ganovski K.H., Effect of Black Sea algae on chicken egg production and on chick embryo development, *Veterinarno- Meditsinski Nauki*, **19**, 99–105 (**1982**)
- **5.** Banerjee K., Mitra A. and Mondal K., Cost-effective and ecofriendly shrimp feed from red seaweed *Catenella repens* (Gigartinales: Rhodophyta), *Current Biotica*, **4(1)**, 23-43 (**2010**)
- 6. Cruz-Sua´ rez L.E., Ricque-Marie D., Tapia-Salazar M. and Guajardo-Barbosa C., Uso de harina de kelp (Macrocystis pyrifera) en alimentos para camaro´ n. In: Avances en Nutricio´n Acui´cola V Memorias del Quinto Simposium Internacional de Nutricio´n Acui´cola, Me´rida, Me´xico, 19–22 Noviembre 2000 (Cruz-Sua´ rez, L.E., Ricque-Marie, D., Tapia-Salazar, M., Olvera-Novoa, M.A. & Cerecedo-Olvera, R. eds), vol. 5, pp. 227–266, Universidad Auto´noma de Nuevo Leo´ n, Monterrey, Me´xico; ISBN 970-694-52-9 (2000)
- 7. Da Silva R. L. and Barbosa J. M., Seaweed meal as a protein source for the white shrimp *Litopenaeus vannamei*, *Journal of Applied Phycology*, 21, 193–197 (2009)

- **8.** Mondal K., Ghosh R., Bhattacharyya S. B., Zaman S., Mallik A., Das M. and Mitra A., Partial replacement of fish meal with mangrove based plant ingredients and its effect on water quality, growth performance and length-weight relationship of freshwater prawn *Macrobrachium rosenbergii*, *Species*, **3(8)**, 15-21 **(2013)**
- Moss S.M., Growth rates, nucleic acid concentrations, and RNA/DNA ratios of juvenile white shrimp, *Penaeus* vannamei Boone, fed different algal feeds, *Journal of* Experimental Marine Biology and Ecology, 182, 193– 204 (1994)
- 10. Pen aflorida V.D. and Golez N.V., Use of seaweed meals from *Kappaphycus alvarezii* and *Gracilaria heteroclada* as binders in feeds of juvenile shrimp *Penaeus monodon*, Aquaculture, **143**, 393–401 (**1996**)
- 11. Valente L.M.P., Gouveia A., Rema P., Matos J., Gomes E.F. and Pinto I.S., Evaluation of three seaweeds *Gracilaria bursapastoris*, *Ulva rigida* and *Gracilaria cornea* as dietary ingredients in European seabass (*Dicentrarchus labrax*) juveniles, *Aquaculture*, 252, 85–91 (2006)
- **12.** Hashim R. and Mat-Saat N.A., The utilization of seaweed meals as binding agents in pellet feeds for snakehead (*Channa striatus*) fry and their effects on growth, *Aquaculture*, **108**, 299–308 (**1992**)
- 13. Vila A.S., Coughlan S., Guiry M.D. and Kraan S., The red alga *Porphyra dioica* as a fish-feed ingredient for rainbow trout (*Oncorhynchus mykiss*): effects on growth, feed efficiency and carcass composition, *Journal of Applied Phycology*, 21, 617-624 (2009)
- 14. Mitra A., Mondal K. and Banerjee K., Effect of saltmarsh grass (*Porteresia coarctata*) feed on growth performance of black tiger shrimp, *Penaeus monodon*, In: Sinha A., Datta S. and Mahapatra B.K. (Eds.) *Diversification of Aquaculture*, Narendra Publishing House, New Delhi, 219-232 pp (2011)
- **15.** Vhanalakar S.A. and Muley D.V., The potential of Alfalfa leaf meal as a fish feed, *All About Feed*, **20** (7), 12-13 (**2012**)
- **16.** Vijayaraghavan S. and Ramadhas V., Conversion efficiency in the shrimp, *Metapenaeus monoceros* (Fabricius), fed decomposed mangrove leaves, *Indian Journal of Marine Sciences*, **9**, 123-125 (**1980**)
- 17. Vijayaraghavan S. and Wafar S., Further studies in using mangrove foliage as a prawn feed, *Mahasagar- Bulletin of the National Institute of Oceanography*, **16** (3), 309-316 (1983)
- **18.** Tsappis A., The algae opportunity, *All About Feed*, **21** (5), 21-23 (2013)

- **19.** Men-Qing L., Qing C.H. and Aksnes A., Identification of feeding stimulants for shrimp, *Marine Fisheries Research*, **22**, 71–74 (**2001**)
- **20.** Van Alstyne K.L., Wolfe G.V., Freidenburg T.L., Neill A. and Hicken C., Activated defense systems in marine macroalgae: evidence for an ecological role for DMSP cleavage, *Marine Ecology Progress Series*, **213**, 53–65 (**2001**)
- **21.** Bandaranayake W.M., Bioactivities, bioactive compounds and chemical constituents of mangrove plants, *Wetlands Ecology and Management*, **10**, 421-452 (**2002**)
- **22.** Rollet B., Bibliography on mangrove research, 1600-1975, *UNESCO Paris*, Pub Information Retreival Ltd., London, 479 pp (**1981**)
- 23. Basak U.C., Das A.B. and Das P., Chlorophylls, carotenoids, proteins and secondary metabolites in leaves of 14 species of mangrove, *Marine Science Bulletin*, 58, 654–659 (1996)
- **24.** Basak U.C., Das A.B. and Das P., Seasonal changes in organic constituents in leaves of nine mangrove species, *Marine and Freshwater Research*, **49**, 369–372 (**1998**)
- **25.** Gody S.A.P., Mayworm M.A.S., Lo V.K. and Salatino A., Contents of lignin, nitrogen and tannins in leaves of typical species of mangroves, *Revista Brasileira de Botanica*, **20**, 35–40 (**1997**)
- **26.** Tam N.F.Y., Wong Y.S., Lan C.Y. and Wang L.N., Litter production and decomposition in a subtropical mangrove swamp receiving wastewater, *Journal of Experimental Marine Biology and Ecology*, **226**, 1–18 (**1998**)
- 27. Mukhopadhyay P. K., Rangacharyulu P. V., Mitra G. and Jana B. B., Applied Nutrition in Freshwater Prawn, *Macrobrachium rosenbergii* Culture, *Journal of Applied Aquaculture*, 13 (3-4), 317-340 (2003)
- **28.** Lowry O.H., Rosebrough N.J., Farr A. L. and Randall R. J., Protein measurement with the Folin-Phenol reagents, *Journal of Biological Chemistry*, **193**, 265-275 (**1951**)
- **29.** Dubois M., Gilles K.A., Hamilton J. K., Rebers P. A. and Smith F., Colorimetric method for determination of sugars and related substances, *Analytical Chemistry*, **28**, 350-356 (**1956**)
- 30. Tecator., Fat extraction on feeds with the Soxtec System HT the influence of sample preparation and extraction media- Application note AN 67/83 (1983.06.13), Soxtec System HT Manual, Tecator AB, Sweden (1983)
- **31.** AOAC., Methods 942.05, 962.09, 920.36. In: Horritz, E. (Ed.) Official Methods of Analysis, 12th Edition, 684 pp, AOAC, Washington DC, USA (**1990**)
- **32.** Chow S. and Sandifer P.A., Differences in growth, morphometric traits and male sexual maturity among

- pacific white shrimp, *Penaeus vannamei*, from different commercial hatcheries, *Aquaculture*, **92**, 165-178 (**1991**)
- **33.** Ricker W.E., Linear regression in fishery research, *Journal of Fisheries Research Board of Canada*, **30**, 409-434 (**1973**)
- 34. Arasaki S. and Arasaki T., Vegetables from the Sea, Japan Publishing Inc., Tokyo, 196 pp (1983)
- 35. Banerjee K., Ghosh R., Homechaudhury S. and Mitra A., Biochemical composition of marine macroalgae from Gangetic Delta at the Apex of Bay of Bengal, *African Journal of Basic and Applied Science*, **1(56)**, 96–104 (2009)
- **36.** Chakraborty S. and Santra S.C., Biochemical composition of eight benthic algae collected from Sunderban, *Indian Journal of Marine Science*, **37(3)**, 329–332 (**2008**)
- **37.** Kaliaperumal N., Chennubhotla V. S. K., Kalimuthu S., Ramalingam J. R., Selvaraj M. and Najmuddin M., Chemical composition of seaweeds, *CMFRI Bulletin*, **41**, 31-51 (**1987**)
- **38.** Manivannan K., Thirumaran G., Devi G.K., Hemalatha A. and Anantharaman P., Biochemical Composition of Seaweeds from Mandapam Coastal Regions along Southeast Coast of India, *American-Eurasian Journal of Botany*, **1** (2), 32-37 (2008)
- **39.** Manivannan K., Karthikai Devi G., Thirumaran G. and Anantharaman P., Mineral composition of marine macroalgae from Mandapam coastal regions; Southern coast of India, *American-Eurasian Journal of Botany*, **1**, 42–51 (**2009**)
- **40.** Naidu K. A., Tewari A., Joshi H. V., Viswanath S., Ramesh H. P. and Rao S. V., Evaluation of nutritional quality and food safety of seaweeds of India, *Journal of Food Safety*, **13** (2), 17-90 (1993)
- **41.** Jagtap T. G., Bhosale S. and Singh C., Characterization of *Porteresia coarctata* beds along the Goa coast, India, *Aquatic Botany*, **84**, 37–44 (**2006**)
- **42.** Jones A. L. and Harwood J. L., Lipids and lipid metabolism in the marine alga *Enteromorpha intestinalis*, *Phytochemistry*, **34(4)**, 969–972 (**1993**)
- **43.** Marcum K. B. and Murdoch C. L., Salt tolerance of the coastal salt marsh grass, *Sporobolus virginicus* (L.) kunth, *New Phytologist*, **120**, 281-288 (**1991**)
- **44.** Phlenger C. F., Effect of salinity on growth of a salt marsh grass, *Ecology*, **52** (**5**), 908-911 (**1971**)
- **45.** Hoq M. E., Islam M. L., Paul H. K. and Ahmed S. U., Decomposition and seasonal changes in nutrient constituents in mangrove litter of Sundarbans mangrove, Bangladesh, *Indian Journal of Marine Sciences*, **31** (2), 130-135 (2002)

- **46.** Odum E.P. and Heald R.J., Detritus based food web of an estuarine mangrove community. In: Cronin L. E. (Ed.) *Estuarine Research*, Academic Press, New York-London, **1**, 256-286 (**1975**)
- **47.** Untawale A.G., Bhosle N.B., Dhargalkar V.K., Matondkar S.G.P. and Bukhari S.S., Seasonal variation in major metabolites of mangrove foliage, *Mahasagar-Bulletin of the National Institute of Oceanography*, **11**, 105-110 (**1978**)
- 48. Untawale A.G., Wafar S. and Jagtap T. G., Application of remote sensing techniques to study the distribution of mangroves along the estuaries of Goa. In: Gopal B., Turner R.E., Wetzel R.G. and Whigham D. F. (Eds.) Wetlands- Ecology and Management, National Institute of Ecology and International Scientific Publications, India, 51-67 pp (1980)
- **49.** Vijayaraghavan S., Ramadhas V., Kumari L. K. and Royan J. P., Biochemical changes and energy content of the mangrove, *Rhizophora mucronata* leaves during decomposition, *Indian Journal of Marine Sciences*, **9**, 120-123 (**1980**)
- **50.** Davis D.A. and Arnold C.R., Replacement of fish meal in practical feeds for the Pacific white shrimp, *Litopenaeus vannamei*, *Aquaculture*, **185**, 291–298 (**2000**)
- **51.** Samocha T., Davis D.A., Saoud I.P. and DeBault K., Substitution of fish meal by coextruded soybean poultry by-product meal in practical feeds for the Pacific white shrimp, *Litopenaeus vannamei*, *Aquaculture*, **231**, 197-203 (**2004**)
- **52.** Jintasataporn O., Tabthipwon P. and Yenmark S., Feedary protein for growth is the goal of nutritionists for the development of cost effective feeds, *Kasetsart Jounal of Natural Science*, **38**, 1-7 (**2004**)
- **53.** Hossain M.A. and Paul L., Low-cost feed for monoculture of giant freshwater (*Macrobrachium rosenbergii* de Man) in Bangladesh, *Aquaculture Research*, **38**, 232-238 (**2007**)
- 54. Md Hasanuzzaman A. F., Md Siddiqui N. and Md Chisty A. H., Optimum Replacement of fishmeal with Soybean Meal in Feed for *Macrobrachium rosenbergii* (De Man, 1879) Cultured in Low Saline Water, *Turkish Journal of Fisheries and Aquatic Sciences*, 9, 17-22 (2009)
- **55.** Mitra A., Basu S., Banerjee A. and Banerjee K., Culture Of Freshwater Prawn using feed incorporated with saltmarsh grass, *Porteresia coarctata* in Indian Sundarbans, *Seshaiyana*, **15** (1), 16-17 (2007)
- **56.** Becker K. and Makkar H.P.S., Effects of feedary tannic acid and quebracho tannin on growth performance and metabolic rates of common carp (*Cyprinus carpio L.*), *Aquaculture*, **175**, 327–335 (**1999**)

- 57. Maitra S. and Ray A.K., Inhibition of digestive enzymes in rohu, *Labeo rohita* (Hamilton), fingerlings by tannin: an in vitro study, *Aquaculture Research*, **34**, 93–95 (2003)
- **58.** Pinto L.G.Q., Pezzato L.E., Miranda E.C., Barros M.M. and Furuya W.M., Effect of tannin on digestibility of Nile tilapia (*Oreochromis niloticus*) feeds, *Acta Scientiarum*, **22**, 677–681 (**2000**)
- 59. Bray W. A. and Lawrence A. L., Influence of dietary fatty acids on reproduction of P. stylirostris (Abstract), Journal of the World Aquaculture Society, 19, 18A (1988)
- 60. Bray W. A., Lawrence A. L. and Lester L. J., Reproduction of eyestalk- ablated *Penaeus stylirostris* fed various levels of total dietary, *Journal of the World Aquaculture Society*, 21, 41-52 (1990a)
- 61. Emerson W. D., Induced maturation of prawn *Penaeus indicus*, *Marine Ecology Progress Series*, 2, 121-131 (1980)
- **62.** Emerson W.D., Hayes D. P. and Ngonyame M., Growth and maturation of *Penaeus indicus* under blue and green light, *South Africa Journal of Zoology*, **18**, 71-75 (**1983**)
- 63. Dall W., Hill B. J., Rothlisberg P. C. and Staples D. J., Biology of the Penaeidae, In: Blaxter J. H. S. and Southward A. J. (Eds.) *Advances in Marine Biology*, Academic Press, London, UK, 27, 489pp (1990)
- 64. Jayachandran K.V. and Joseph N.I., Length-weight relationship of two palaemonid prawns *Macrobrachium idella* and *M. scabriculm* A comparative study, *Fishery Technology*, 25, 189-195 (1988)
- **65.** Abohweyere P.O. and Williams A. B., Length-weight relationship and condition factor of *Macrobrachium macrobrachion* in the Lagos-Lekki lagoon system, Nigeria, *Research Journal of Biological Sciences*, **3(11)**, 1333-1336 (**2008**)
- Beyer J. E., On length weight relationship computing the mean weight of the fish of a given length class, *Fish Byte*,5 (1), 11-13 (1987)
- 67. Hart A. I. and Abowei J. F. N., A study of the length-weight, condition factor and age of ten fish species from the lower Nun river, Niger Delta, *African Journal of Applied Zoology and Environmental Biology*, 9, 13-19 (2007)
- **68.** Abowei J.F.N., Tawari C.C., Deekae S.N. and Amakiri N.E., A study of the length-weight relationship and condition factor of *Pseudotolithus elangatus* (Browdich, 1825) from Bonny estuary, Niger Delta, Nigeria, *International Journal of Tropical Agriculture Food Systems*, **2(3-4)**, 249-254 (**2008**)

- **69.** Bolger T. and Connoly P.L., The selection indices for the measurement and analysis of fish condition, *Journal of Fish Biology*, **30**, 171-182 (**1989**)
- **70.** Abrahamson S., Dynamics of an isolated population of the crayfish, *Astacus astacus* (Linne) and *Pasifastacus leniusculus* (Dana), *Svensk Naturvetenskap*, 306-316 (**1966**)
- 71. Aliakbar M. and Ali M. L., Length-weight relationship of *Macrobrachium rosenbergii* (De Man) of Dakatia River, *Bangladesh Journal of Aquaculture*, 1(1), 74-78 (1978)
- **72.** Hossain M. A., Ali M. A., Islam M. S., Sahabuddin M. and Dewan S., Length-weight relationship and condition factor of *Macrobrachium rosenbergii* (De Man) of Tetulia River, *Bangladesh Journal of Fisheries*, **10**(1), 89-95 (**1987**)
- 73. Kunda M., Dewan S., Uddin M. J., Karim M., Kabir S. and Uddin M. S., Length-Weight Relationship, Condition Factor and Relative Condition Factor of *Macrobrachium rosenbergii* in Rice Fields, *Asian Fisheries Science*, 21, 451-456 (2008)
- 74. Kuris A.M., Ra'anan Z., Sagi A. and Cohen D., Morphometric differentiation of male Malaysian giant prawns *Macrobrachium rosenbergii*. *Journal of Crustacean Biology*, 7, 219-237 (1987)
- **75.** Enin U. I., Length-weight parameters and condition factor of two West African prawns, *Revue d' Hydrobiologie Tropicale*, **27(2)**, 121-127 **(1994)**
- **76.** Bhavan P.S., Devi V.G., Shanthi R., Radhakrishnan S. and Poongodi R., Basic biochemical constituents and profiles of amino acids in the post larvae of *Macrobrachium rosenbergii* fed with *Spirulina* and Yeast enriched *Artemia*, *Journal of Scientific Research*, **2(3)**, 539-547 (**2010**)
- 77. Cruz-Sua'rez L.E., Tapia-Salazar M., Nieto-Lo'pez M.G., Guajardo-Barbosa C. and Ricque-Marie D., Comparison of *Ulva clathrata* and the kelps *Macrocystis pyrifera* and *Ascophyllum nodosum* as ingredients in shrimp feed, *Aquaculture Nutrition*, 15, 421-430 (2009)
- **78.** Fair P.H., Frontner A.R., Millikin M.R. and Sick L.V., Effect of feedary fibre on growth, assimilation and cellulase activity of the prawn *M. rosenbergii*, *Proceedings of the World Mariculture Society*, **11**, 369-370 (**1980**)
- **79.** Lee P.G., Blake N.J. and Rodrick G.E., A quantitative analysis of digestive enzymes for the freshwater prawn *Macrobrachium rosenbergii*, *Proceedings of the World Mariculture Society*, **11**, 392-402 (**1980**)
- **80.** Murthy R.C., Study of proteases and esterases in the digestive system of *Macrobrachium lamarri* Edwards (Crustacea: Decapoda), *Journal of Animal Morphology and Physiology*, **24**, 211-216 (**1977**)

- 81. Tyagi A.P. and Prakash A., A study on the physiology of digestion in freshwater prawn *Macrobrachium dayenum*, *Journal of the Zoological Society of India*, 19, 77-83 (1967)
- **82.** Thomas G., Relationship between growth rate and RNA/DNA protein ratio in *Penaeus indicus* (H. Milne Edwards), *CMFRI Special Publication*, 58-60 (**1993**)
- **83.** Ali S. and Sahu N.P., Response of *Macrobrachium rosenbergii* (de Man) juveniles to fish silage as substitute for fishmeal in dry feeds, *Asian Fisheries Science*, **15(1)**, 59-69 (**2002**)
- **84.** Gupta A., Formulation and evaluation of some alternative practical feeds for giant freshwater prawn, *Macrobrachium rosenbergii* (De Man), Ph.D. Thesis submitted to PAU, Ludhiana, India (**2005**)
- **85.** Ferdose A. and Hossain M. B., Nutritional value of wild, cultured and frozen prawn *Macrobrachium rosenbergii* (De Man, 1879), *International Journal of Natural Sciences*, **1(2)**, 52-55 (**2011**)
- **86.** Dinakaran G.K. and Soundarapandian P., Biochemical Status of Edible Palaemonid Prawn *Macrobrachium idella idella* (Hilgendorf, 1898), *Advance Journal of Food Science and Technology*, **1(1)**, 19-26 (**2009**)

- 87. Soundarapandian P. and Ananthan G., Effect of unilateral eyestalk ablation on the biochemical composition of commercially important juveniles of *Macrobrachium malcolmsonii*, *International Journal of Zoological Research*, 4(2), 106-112 (2008)
- **88.** Tiwary A. K., Nutritional status of edible palaemonid prawn *Macrobrachium scabriculum* (Heller, 1862), M.Sc. Thesis, Annamalai University, India (**2009**)
- **89.** Chandra S.K., Proximate composition of edible palaemonid prawn *Macrobrachium idae* (Heller, 1862), M.Sc. Thesis, Annamalai University, India (**2009**)
- 90. Shyla G., Nair C. M., Shalin K. R., Sherief P. M. and Mukundan M. K., Liver oil of pharaoh cuttlefish *Sepia pharaonis* Ehrenberg, 1831 as a lipid source in the feed of giant freshwater prawn, *Macrobrachium rosenbergii* (De Man 1879), *Aquaculture Nutrition*, 15, 273-281 (2009)
- **91.** Sakkaravarthi K., Sankar G., Elavarasi A. and Ramamoorthy K., Biochemical Changes and Growth Performance of Black Tiger Shrimp Larvae after using *Ricinus communis* extract as feed additive, *International Journal of PharmaTech Research*, **3(1)**, 201-208 **(2011)**