

Analysis of Glutathione S-Transferase Pi (GSTP1) Expression in Non Melanoma Skin Cancer using Bioinformatic Tools

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Abstract

A bioinformatics study was carried out to study the expression of GSTP1 in non melanoma skin cancer (actinic keratosis and squamous cell carcinoma) by selecting non melanoma skin cancer research (record number GDS 2200) from the NCBI site. The experiment was analyzed using GEO data set and expression of GSTP1 in both control and actinic keratotic (AK) lesion, and squamous cell carcinoma (SCC) tumor biopsies data from 5 patients with non-melanoma skin cancer (NMSC) were analyzed. Known interactions for GSTP1 and coexpressed genes were also carried out using STRING database. The result shows that GSTP1 was over-expressed in the non melanoma samples (actinic keratosis and squamous cell carcinoma) compare with the control. Although one sample in the actinic keratosis shows higher expression, all the samples in squamous cell carcinoma show high expression of GSTP1. Result from STRING data base show that GSTP1 interact and co-expressed with a number of genes which are important in understanding of cancer and progression of the disease. From the data generated, we concluded that GSTP1 was over-expressed in non melanoma skin cancer.

Keywords: Analysis, glutathione, expression, skin cancer, Bioinformatic tools.

Introduction

Human GSTP1 has drawn considerable interest due to its over expression in cancer cells and likely involvement in acquired resistance of chemotherapeutic agents. The multifunctional enzyme glutathione S-transferase pi (GSTP1) is an important enzyme belonging to the glutathione S-transferase (GST) family of enzymes that carries out detoxification thereby inactivating electrophilic carcinogens by conjugation with reduced glutathione^{1,2}. The enzyme participates in detoxification reactions by catalyzing the conjugation of many electrophilic and hydrophobic compounds with reduced glutathione. The soluble GSTs are categorized into 4 main classes based on their biochemical, immunologic, and structural properties. These classes include alpha, mu, pi, and theta. Hypermethylation has been found to have an effect on the regulatory sequence closed to the GST gene during the early stages of carcinogenesis³⁻⁶. Several classes of GST were earlier been reported in tissues of human beings and are being classified as alpha, mu, pi, and theta. An increased expression of GST pi has been reported in cancers of the breast, bladder, pancreas, colon, stomach, lung, neck and head, cervix, ovary, and, as well as soft tissue sarcoma, testicular embryonal carcinoma, glioma and meningioma⁷⁻¹⁶. GSTP1 functions in metabolism of xenobiotics and is engaged in susceptibility to cancer, and other diseases. Reports link increased expression of glutathione S-transferase (GST) family of enzymes to acquired resistance to antineoplastic drugs¹⁷. Particular GST subclasses expression protects cancer cells from the cytotoxicities of cancer drugs, and antineoplastic drug resistance has been connected to over-expression of GST¹⁸. Substrate specificity of GST enzymes is

of extensive among which are known to possess mutagenic properties. Serum GST pi elevation has been developed and exploited as a marker for serum tumor gastrointestinal cancer⁸ and non-Hodgkin's lymphoma¹⁹ and also as a method of predicting chemotherapy sensitivity. GST plays distinct roles in resistance to chemotherapy drugs via direct detoxification as well as inhibition of the MAP kinase pathway. GSTs localized mainly in the cytosol and are dimeric in nature. In addition to their catalytic role in detoxification, the enzymes also have broad ligand binding properties²⁰. The expression of GST-pi has been compared in several malignant and normal tissues²¹ and the research suggest that expression of GST-pi increases in many tumors compared to normal tissues. GST3 has been demonstrated to be abundantly expressed in human skin²². The aim of this paper is investigate whether there is over-expression of GSTP1 in non melanoma skin cancer specifically actinic keratosis lesion and squamous cell carcinoma.

Methodology

Selecting the experiment: Suitable experiment was selected from NCBI GEO data set by pointing Internet browser at <http://www.ncbi.nlm.nih.gov/gds>. Skin cancer was entered into search option and experiment with the title "non melanoma skin cancer" was selected. The detail of the experiment appears. The record number of the experiment GDS 2200, was noted for further analysis. The normal, actinic keratosis and squamous cell carcinoma samples number was also noted (normal: GSM47616, GSM47617, GSM47618, GSM47619, GSM47620, and GSM47621; actinic keratosis: GSM47612, GSM47613, GSM47614, GSM47615 and squamous cell carcinoma:

GSM47622, GSM47623, GSM47624, GSM47625 and GSM47626).

Analysis of the experiment using NCBI Geo data set data analysis tools: The record GDS 2200 was clicked and the main page having experimental details and analysis tools appears. The gene of interest (GSTP1) was entered into “find gene name or symbol” and Go button was clicked. A new window having expression pattern image of GSTp1 in the experiment appears

along with gene annotation details. The image was click to visualize the differential expression of GSTP1 in both normal, actinic keratosis and squamous cell carcinoma samples (figure 3). Genes that are expressed up/down in the disease state were also analyzed by clicking Go button on the data analysis tools. A new window having list of gene showing differential expression in this experiment appears along with expression pattern images and gene annotation details. The image below the cluster was clicked to find co-expressed genes.

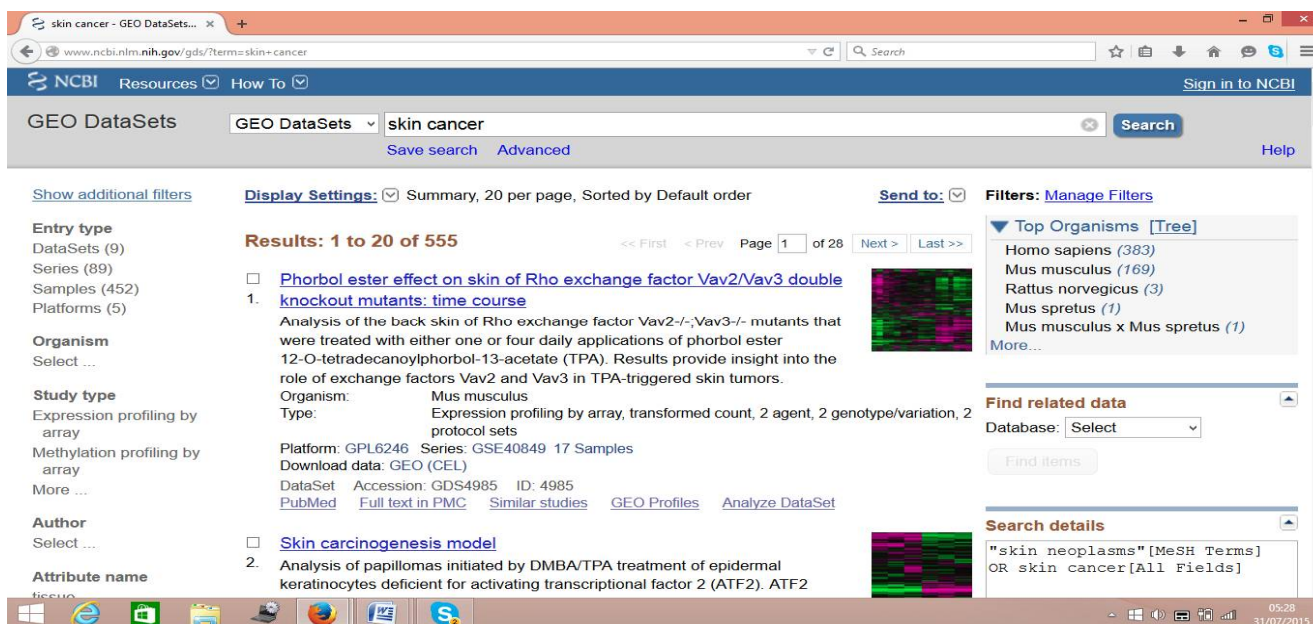


Figure-1
Selecting experiment from NCBI site

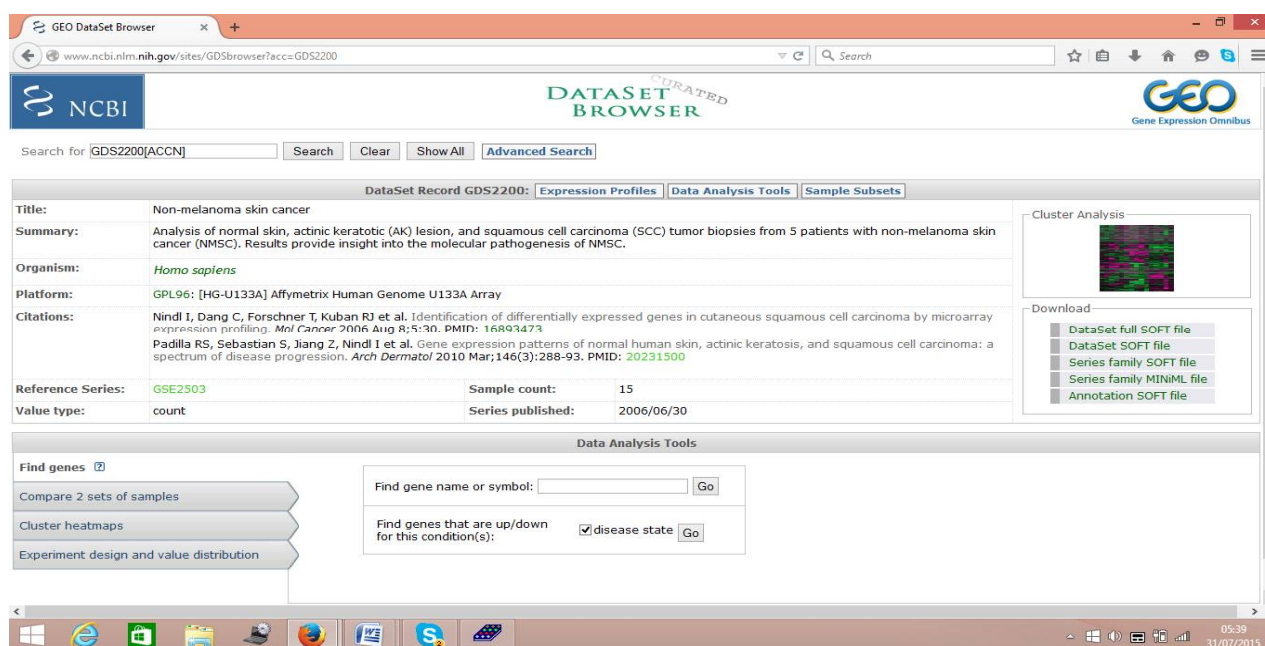


Figure-2
New window showing details of the selected experiment and data analysis tools

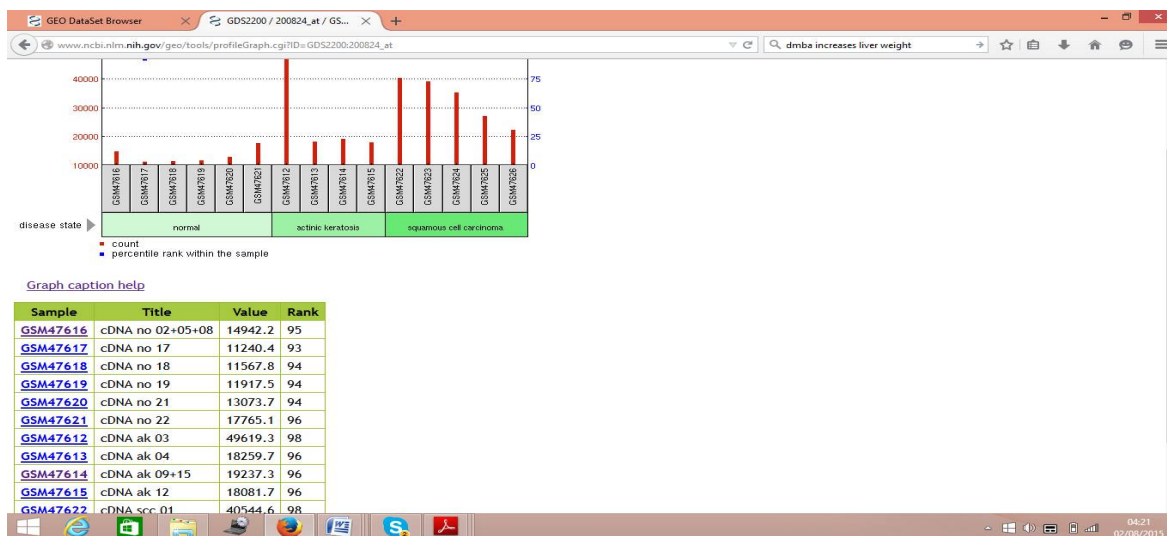


Figure 3
Visualizing the differential expression of GSTP1

Finding known interactions for GSTP1 and co-expressed genes using STRING database: The knowledge about interactions of GSTP1 and co-expressed genes will be helpful in understanding their role in cancer. The browser was pointed to STRING database at <http://string-db.org/> (figure 4) GSP1 was searched into the protein name. Homo sapiens were selected as the organism's name. The interactions were finally analyzed.

Results and Discussion

The result of the analysis of glutathione s-transferase pi expression in non melanoma skin cancer using bioinformatics tools were display in figure-4. The original samples were collected from 5 patients and were Analysis for normal skin, actinic keratotic (AK) lesion, and squamous cell carcinoma (SCC) tumor biopsies from 5 patients with non-melanoma skin cancer (NMSC)²³.

The result shows that GSTP1 was over-expressed in the non melanoma samples (actinic keratosis and squamous cell carcinoma) compare with the control. Although one sample in the actinic keratosis shows higher expression, all the samples in squamous cell carcinoma show high expression of GSTP1. GSTP1 has become a subject of particular interest with regard to cancer, because many cancer cell lines and tumors are characterized by high GSTP1 expression²⁴. it gene is encoded by a single gene has been mapped to chromosome 11q13. The mechanism by which GSH carried out detoxification involves nucleophilic attack on the sulphur atom of GSH onto the electrophilic group of many foreign substances and detoxification of anticancer drugs occurs not only by acting directly on the molecules but rather on a metabolite²⁵. This action of GSTP1 significantly decreases the reactivity of these xenobiotics and increases their solubility favoring their elimination. In view of these, GST enzymatic activity may play a key role in the detoxification of a variety of anticancer

drugs^{25,26}. Over-expression of GSTP1 was observed in nearly all of human tumor cell lines, including those that are chemotherapeutic resistance^{27,28}. A varieties of human cancers such as kidney; colon, breast, ovarian and lung usually express high levels of GSTP1 in comparison to the surrounding tissues. Accordingly, GSTP1-1 expression has been explored to be a marker for cancer development^{29,30}. High expression levels have been correlated not only with drug resistance in patients undergoing chemotherapy but also with disease progression.

Analysis of the known and predicted interactions of GSTP1 have indicated that GSTP1 interact with a number of genes CPY1B1, CPY1A1, MAPK8, GSTB2B, EPHX1, GSST1, PRDX6, GSTO1, GSTT1, CYP2E1 and CYP1A2 (figure-6). The knowledge about interactions of GSTP1 and co-expressed genes will be helpful in understanding their role in cancer as well as progression of the disease. Association between several isoenzymes from GST classes with mitogen-activated protein kinase (MAPK) has been well established. Mitogen-activated protein kinase (MAPK) is involved in cell death signaling and cell survival and GSTP1 has been implicated in inhibition of this pathway thereby favoring survival of cancer cells. GSTs function non-enzymatically by sequestering the kinase in a complex, therefore preventing it from acting on downstream targets, consequently regulating pathways that control apoptotic cell death and cell proliferation. GST P1 was found to inhibit c-Jun N-terminal kinase (JNK) through direct protein-protein inter-action³¹. JNK is a MAP kinase involved in cellular differentiation, stress response, inflammation, apoptosis and proliferation. Variety of stress such as ultraviolet (UV) radiation a protein synthesis inhibitors, may result in activation of JNK which subsequently phosphorylates c-Jun, a part of the activator protein-1 (AP-1) transcription factor. This activation results in induction of AP-1-dependent target genes involved in cell proliferation and cell death³².

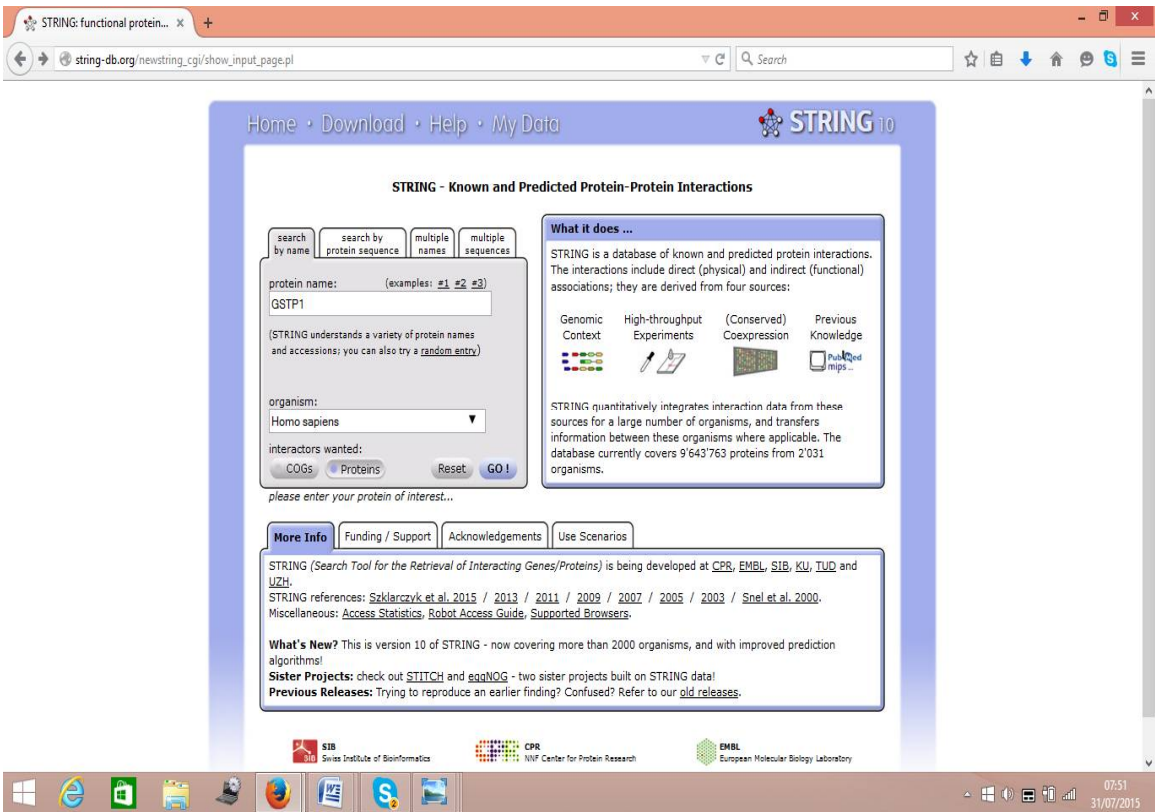


Figure-4
String database interface for finding known interactions and co-expressed genes

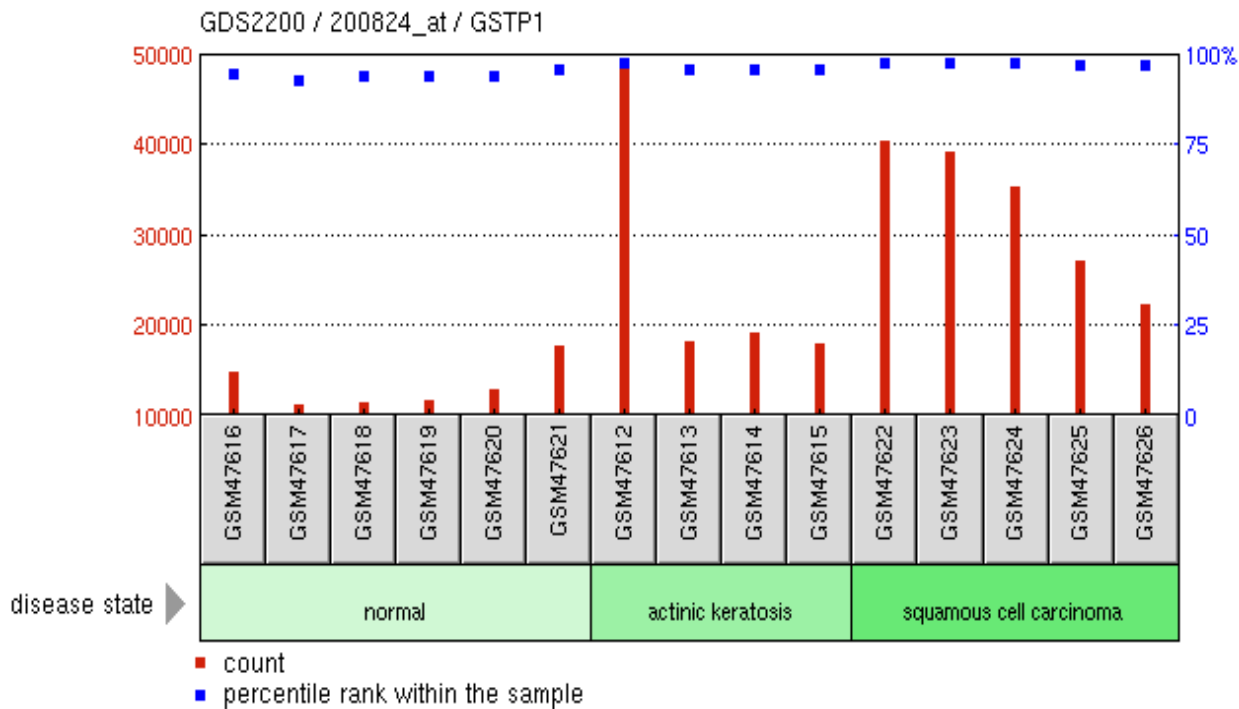


Figure-5
Expression profile of GSTP1 from normal (non-sun-exposed skin), normal (sun-exposed skin), AKs, and SCCs

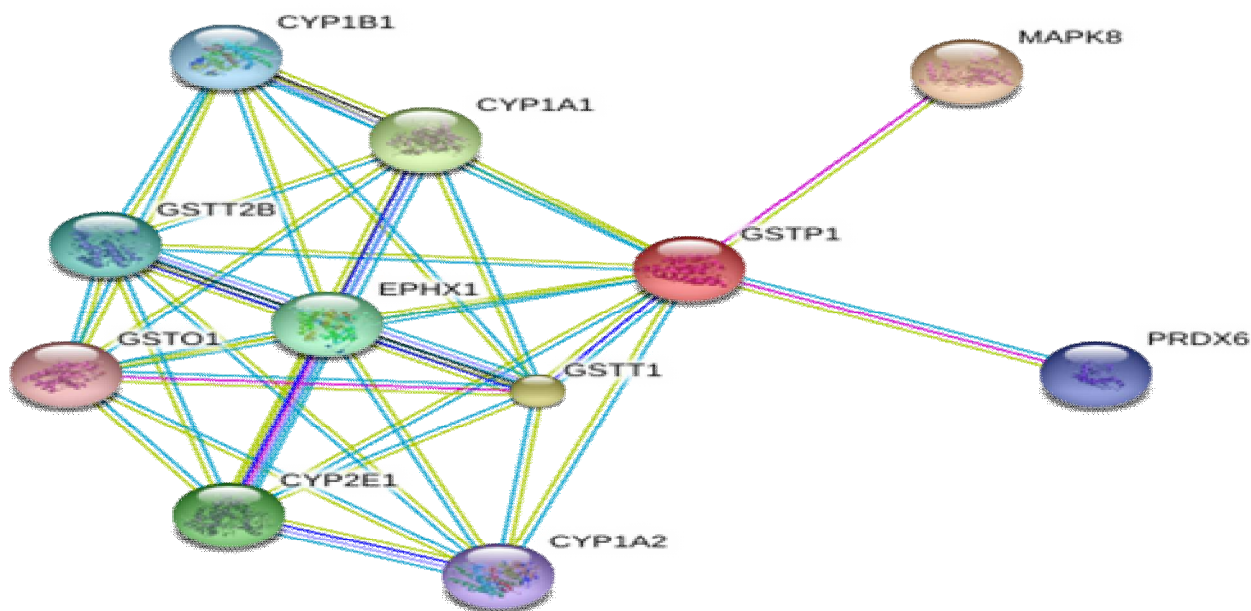


Figure-6
Evidence view of association of GSTP1 with other genes, Different line colors represent the types of evidence for the association

In this research, we have predicted that GSTP1 interact with about 10 different enzymes or proteins (table-1). Each of these enzymes may play significant role in detoxification. Understanding these interactions between GSTP1 and the predicted partners will help in understanding synergistic mechanism of the drug resistance to chemotherapeutics agent by the cancer cells. The score of these interactions were given in table 1 and it indicates the degree of the association. Mitogen-activated protein kinase 8 (MAPK8) was found to strongly interact with GSTP1 and it has the highest score of 0.997 with amino acid 427. The least of the interaction was found between Glutathione S-transferase omega 1 (GSTO1) and GSTP1 and the score 0.956 with about 241 amino acids.

A more complex association have shown that GSTP1 interact indirectly with a number of genes (MAP2K4, MAP3K1, CDC42, FASLG, MAPK8IP3, NFATC2, HRAS, RHOA, SP1, APP, FOXO3, MAP2K7, JUN, FOS, STAT3, FOXO1, MAPT, MAPKB1P2, CRK, MAPK8, BCL2, BCL2L1, BCL2L1, DUSP16, DUSP4, PXN, IRS1, JUNB, IRS2, DUSP2, DUSP8, ATF2, DUSP1, JUND, DUSP10, and BAD (figure 7). Clear understanding of interactions of these genes with GSTP1 will help in understanding cell signaling involved in expression of GST pi that were earlier reported in cancers of the breast, bladder, pancreas, colon, stomach, lung, neck and head, cervix, ovary, and, as well as soft tissue sarcoma, testicular embryonal carcinoma, glioma and meningioma⁷⁻¹⁶. Accordingly inhibition of GSTP1 would increase the sensitivity of cancer cells to chemotherapeutic agents.

Table-1
Predicted partners of GSTP1

Enzyme/Protein	No. of amino acids	Score
Mitogen-activated protein kinase 8 (MAPK8)	427	0.997
Glutathione S-transferase theta 1(GSTT1)	240	0.993
Cytochrome P450, family 1, subfamily A, polypeptide 1(CYP1A1)	512	0.980
Cytochrome P450, family 2, subfamily E, polypeptide 1(CYP2E1)	455	0.971
Epoxide hydrolase 1, microsomal (EPHX1)	244	0.979
Glutathione S-transferase theta 2B (GSTT2B)	244	0.970
Cytochrome P450, family 1, subfamily B, polypeptide 1 (CYP1B1)	543	0.968
Peroxisredoxin 6 (PRXD6)	224	0.967
Cytochrome P450, family 1, subfamily A, polypeptide 2 (CPY1A2)	516	0.962
Glutathione S-transferase omega 1 (GSTO1)	241	0.956

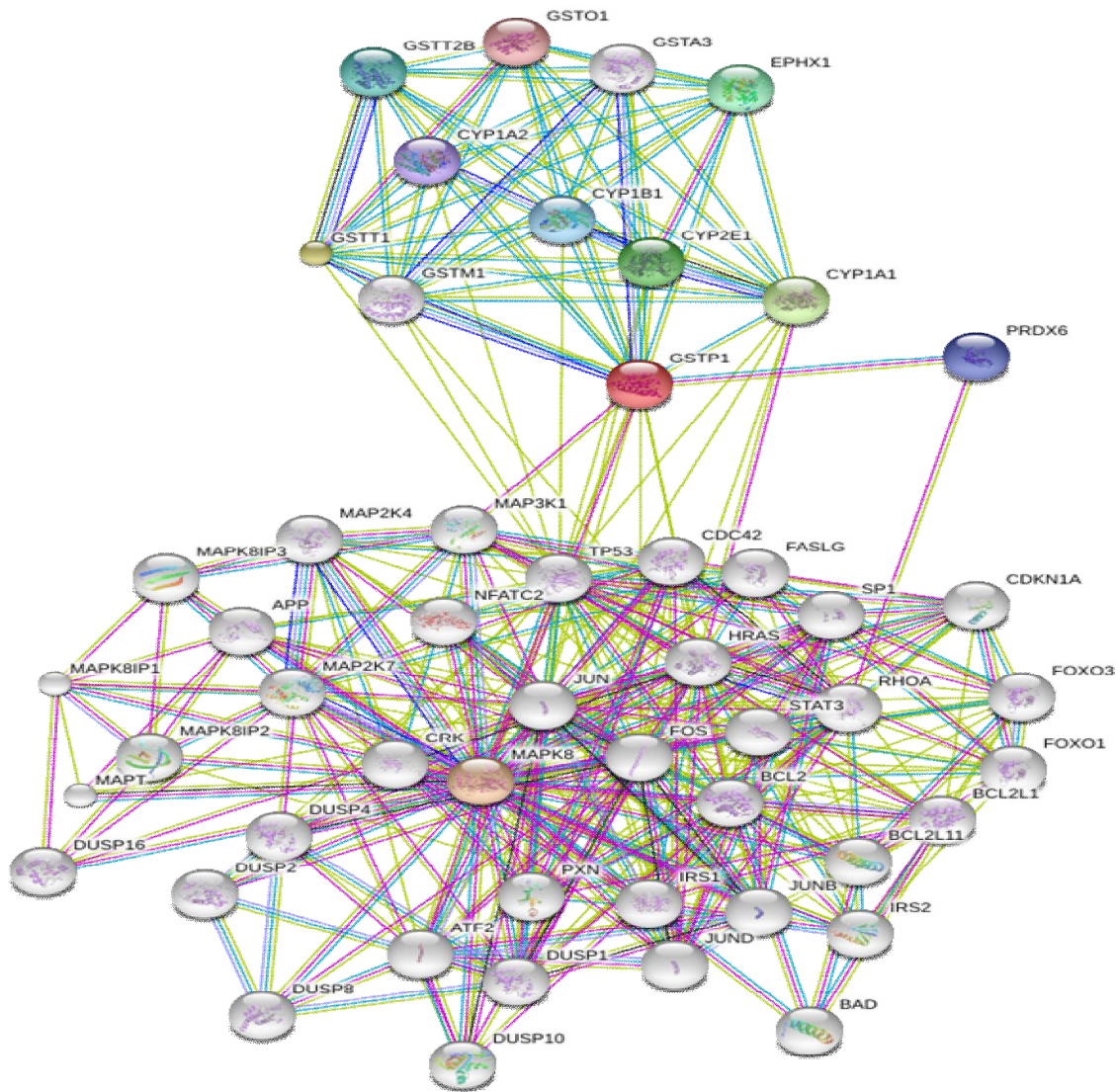


Figure-7

A Complex Evidence view of association of GSTP1 with other genes, Different line colors represent the types of evidence for the association

Conclusion

From the result of the analysis of glutathione s-transferase expression in non melanoma skin cancer using bioinformatics tools, we have arrived at the conclusion that GSTP1 was over-expressed in non melanoma skin cancer. Accordingly inhibition of GSTP1 would increase the sensitivity of cancer cells to chemotherapeutic agents.

References

1. Toffoli G., Frustaci S., Tumiotto L., Talamini R., Gherlinzoni F., Picci P. and Boiocchi M., Expression of MDR1 and GST-pi in human soft tissue sarcomas: relation

to drug resistance and biological aggressiveness, *Ann Oncol*, **3**, 63-9 (1992)

2. Jeronimo C, Usadel H, Henrique R, Oliveira J, Lopes C, Nelson WG and Sidransky D., Quantitation of GSTP1 methylation in non-neoplastic prostatic tissue and organ-confined prostate adenocarcinoma, *J Natl Cancer Inst.*, **93**, 1747-52 (**2001**)
3. Lee WH, Morton RA, Epstein JI, Brooks JD, Campbell PA, Bova GS, Hsieh WS, Isaacs WB and Nelson WG., Cytidine methylation of regulatory sequences near the p1-class glutathione S transferase gene accompanies human prostatic carcinogenesis, *Proc Natl Acad Sci U S A*, **91**, 11733-7 (**1994**)

4. Brooks JD, Weinstein M, Lin X, Sun Y, Pin SS, Bova GS, Epstein JI and Isaacs WB, Nelson WG. CG island methylation changes near the GSTP1 gene in prostatic intraepithelial neoplasia, *Cancer Epidemiol Biomarkers Prev*, **7**, 531-6 (1998)
5. Cairns P, Esteller M, Herman JG, Schoenberg M, Jeronimo C, Sanchez-Cespedes M, Chow NH, Grasso M, Wu L and Westra WB, Sidransky D. Molecular detection of prostate cancer in urine by GSTP1 hypermethylation, *Clin Cancer Res.*, **7**, 2727-30 (2001)
6. Jeronimo C, Usadel H, Henrique R, Silva C, Oliveira J, Lopes C and Sidransky D., Quantitative GSTP1 hypermethylation in bodily fluids of patients with prostate cancer. *Urology*, **60**, 1131-5 (2002)
7. Niitsu Y, Takahashi Y, Saito T, Hirata Y, Arisato N, Maruyama H, Kohgo Y and Listowsky I., Serum glutathione-S-transferase-pi as a tumor marker for gastrointestinal malignancies, *Cancer*, **63**, 317-23 (1989)
8. Randall BJ, Angus B, Akiba R, Hall A, Cattan AR, Proctor SJ, Jones RA and Horne CH., Glutathione S-transferase (placental) as a marker of transformation in the human cervix uteri: an immunohistochemical study, *Br J Cancer*, **62**, 614-8 (1990)
9. Kantor RR, Giardina SL, Bartolazzi A, Townsend AJ, Myers CE, Cowan KH, Longo DL, Natali PG. Monoclonal antibodies to glutathione S-transferase pi-immunohistochemical analysis of human tissues and cancers, *Int J Cancer*, **47**, 193-201 (1991)
10. Satta T, Isobe K, Yamauchi M, Nakashima I, Takagi H., Expression of MDR1 and glutathione S transferase-pi genes and chemosensitivities in human gastrointestinal cancer, *Cancer*, **69**, 941-6 (1992)
11. Green JA, Robertson LJ and Clark AH., Glutathione S-transferase expression in benign and malignant ovarian tumors. *Br J Cancer*, **68**, 235-9 (1993)
12. Inoue T, Ishida T, Sugio K, Maehara Y, Sugimachi K. Glutathione S transferase Pi is a powerful indicator in chemotherapy of human lung squamous-cell carcinoma, *Respiration*, **62**, 223-7(1995)
13. Bentz BG, Haines GK 3rd, Radosevich JA, Glutathione S-transferase pi in squamous cell carcinoma of the head and neck, *Laryngoscope*, **110**, 1642-7 (2000)
14. Trachte AL, Suthers SE, Lerner MR, Hanas JS, Jupe ER, Sienko AE, Adesina AM, Lightfoot SA, Brackett DJ and Postier RG., Increased expression of alpha-1-antitrypsin, glutathione S transferase pi and vascular endothelial growth factor in human pancreatic adenocarcinoma. *Am J Surg.*, **184**, 647-8 (2002)
15. Simic T, Mimic-Oka J, Savic-Radojevic A, Opacic M, Pljesa M, Dragicevic D, Djokic M and Radosavljevic R., Glutathione S-transferase T1-1 activity upregulated in transitional cell carcinoma of urinary bladder. *Urology*, **65**, 1035-40 (2005)
16. Arai T, Miyoshi Y, Kim SJ, Taguchi T, Tamaki Y and Noguchi S., Association of GSTP1 CpG islands hypermethylation with poor prognosis in human breast cancers, *Breast Cancer Res Treat*, **100**, 169-76 (2006)
17. Wang W, Xia C Q, Liu N, Gan L and Zheng J., Mechanistic study of potentiation of chemotherapy by a haloenol lactone derivative in vitro. *Cancer Chemotherapy and Pharmacology*, **62**, 117-122 (2008)
18. Morrow CS, Smitherman PK, Diah SK, Schneider E and Townsend AJ., Coordinated action of glutathione S-transferases (GSTs) and multidrug resistance protein 1 (MRP1) in antineoplastic drug detoxification. Mechanism of GST A1-1- and MRP1-associated resistance to chlorambucil in MCF7 breast carcinoma cells, *J Biol Chem.*, **7**, 273, 20114-20 (1998)
19. Katahira T, Takayama T, Miyanishi K, Hayashi T, Ikeda T, Takahashi Y, Takimoto R, Matsunaga T, Kato J and Niitsu Y., Plasma glutathione S-Transferase P1-1 as a prognostic factor in patients with advanced non-Hodgkin's lymphoma (stages III and IV), *Clin Cancer Res*, **10**, 7934-40 (2004)
20. Sheehan D, Meade G, Foley V M. and Dowd C A., Structure, function and evolution of glutathione transferases: implications for classification of non-mammalian members of an ancient enzyme superfamily, *Biochem J.*, **360**, 1-16 (2001)
21. Moscow J A, Fairchild C R, Madden M J, Ransom D T, Wieand H , O'Brien E E, Poplack DG and Cossman J, Myers C E and Cowan, K. H. Expression of anionic glutathione-S-transferase and P-glycoprotein genes in human tissues and tumors, *Cancer Res*, **49**, 1422-1428, (1989)
22. Konohana A, Konohana I, Schroeder WT, O'Brien WR, Amagai M, Greer J, Shimizu N, Gammon WR, Siciliano M J and Duvic M., Placental glutathione-S-transferase-pi mRNA is abundantly expressed in human skin, *J Invest Derm*, **95**, 119-126, (1990)
23. Nindl I, Dang C, Forschner T, Kuban RJ Meyer T, Sterry W and Stockfleth E., Identification of differentially expressed genes in cutaneous squamous cell carcinoma by microarray expression profiling, *Mol Cancer*, **5**, 1476-4598 (2006)
24. Lo H.-W, Stephenson L, Cao X, Milas M, Pollock R and Ali-Osman F., Identification and functional characterization of the human Glutathione S-transferase P1 gene as a novel transcriptional target of the p53 tumor suppressor gene, *Mol Cancer Res.*, **6**, 843-850 (2008)
25. Sau A, Tregno F P, Valentino F, Federici G and Caccuri AM, Glutathione transferases and development of new

- principles to overcome drug resistance, *Arch Bioch Biophys*, **500**, 116-122 (2010)
26. Akhdar H, Legendre C, Aninat C and More F., Anticancer Drug Metabolism: Chemotherapy Resistance and New Therapeutic Approaches, InTech, 6-170 (2012)
27. Mannervik B, Castro VM, Danielson UH, Tahir MK, Hansson J and Ringborg U., Expression of class Pi glutathione transferase in human malignant melanoma cells, *Carcinogenesis*, **8**, 1929–1932 (1987)
28. Shea TC, Kelley SL and Henner WD., Identification of an anionic form of glutathione transferase present in many human tumors and human tumor cell lines, *Cancer Res.*, **48**, 527–533 (1988)
29. Tidefelt U, Elmhorn-Rosenborg A, Paul C, Hao XY, Mannervik B and Eriksson LC, Expression of glutathione transferase pi as a predictor for treatment results at different stages of acute nonlymphoblastic leukemia, *Cancer Res.*, **52**, 3281–3285 (1992)
30. Howells RE, Dhar KK, Hoban PR, Jones PW, Fryer AA and Redman CW et al., Association between glutathione-S-transferase GSTP1 genotypes, GSTP1 over-expression, and outcome in epithelial ovarian cancer, *Int J Gynecol Cancer*, **14**, 242–250 (2004)
31. Adler V, Yin Z, Fuchs SY, Benezra M, Rosario L and Tew KD et al., Regulation of JNK signaling by GSTp, *EMBO J.*, **18**, 1321–1334 (1999)
32. Karin M and Gallagher E., From JNK to pay dirt: JUN kinases, their biochemistry, physiology and clinical importance, *IUBMB Life*, **57**, 283–295(2005)