

Review Paper

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A Study on Characteristics of fish mucus and their Antifungal activity

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Abstract

In their aquatic habitat, the fishes are simultaneously exposed to environments comprising various pathogenic microorganisms, so that they need a constant mechanism to combat these pathogens and eliminate potential infections. The integumentary layer in fishes is exchange between them and the external environment protects the fish from pathogenic attacks. Recently, this mucus of the fishes has gained importance in the field of biomedical research, because of its ability to tackle infections caused by bacteria, viruses, and fungi, by providing innate immunity to the fishes. It is being studied for its potential applications in human medicine. This review is an outline of the active potency of fish integumentary mucus and its and effective role against several human pathogens and the treatment of their resulting clinical infections.

Keywords: Mucin, Antimicrobial peptide, Immunological defense, Epithelial cells, Viscous colloid.

Introduction

Nature has been and still is a continuous source of medicinal products. From the ancient time plant is always used as medicine and people don't think towards animals. The main region of this is that the animals are yet poorly explored source for medicine. The animals are well known ingredients for many popular medicines some of them are used as medicinal purposes various insects, Spanish flies, leeches and fishes are listed as medicine. The therapy from animals is called Zoo therapy. Till Date, 109 animals and their medicinal uses are reported in different parts of India.

It an medicines alternative among many other therapies. When food products are used for treating disease owing to its medical and health benefits is called nutraceuticals they include the nutrients like vitamins, minerals, and dietary supplements specific diets.

The fishes are traditionally used to cure infections, reduce pain and given to patients recuperating from operation, injuries and wounds. Both freshwater and marine fish and shell fish contain nutrients essential for the proper growth and development of humans. It is required in adequate amount in all stages of human life from infancy to old age. Consumption of fish along with balanced diet ensures a healthy life with minimal susceptibility to disease. The medicinal quality of fish is harnessed to prevent and cure many harmful diseases like as arthritis, asthma, heart related disease, and various other ailments and they are also an important source of vital nutrients like the proteins, fats vitamins and minerals. But every person does not have knowledge related to the animal also used as medicine for curing different types of diseases.

Role of Fish Mucus

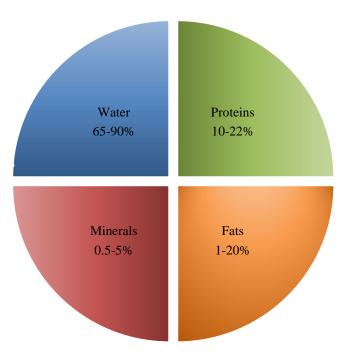
The skin mucus is mechanically protective in nature and a physical barrier between fish with their environment. It is an important component of the innate immune mechanism, provides a first physical and chemical barrier against harmful pathogens³. The Integumentary mucus of fish makes smooth the surfaces and slippery which are mechanical protective nature⁴, prevents pathogen's attacks. It contains many important elements such as proteins, lysozyme, immunoglobulin, and lectins. Nature of the mucus is depending on the species of fish, mucus varies considerably in viscosity, thickness, and glycoprotein (mucin)⁵. Fish skin contains a variety of immune cells such as macrophages, lymphocytes, eosinophilic granulocytes, dendritic cells, and cytokine ⁶. The antimicrobial property against infectious pathogens was demonstrated in rainbow trout⁷.

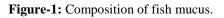
Production of Mucus

The epidermal mucus is produced by the cell present in epithelial layer of fish (goblet, club, and sacciform cells)^{8,9}. Goblet cells are special types of cell present on epidermal layer of skin and gills surfaces of the Fish. The chemical composition of mucus is depending upon the secretory cell. The Goblet cells contain dialkylated, sulphated, and glycoproteins¹⁰. Left the Goblet cells some other cells are also identified whose secretion blend with the goblet cell and give rise mucus. These include sacciform cells and acidophilic granular cells or serious goblet cells, whereas the acidophilic granular cells produce basic proteins rather than glycoproteins. One another mucus secretory cell is club cell whose secretion have larger protein and lesser carbohydrate components¹¹.

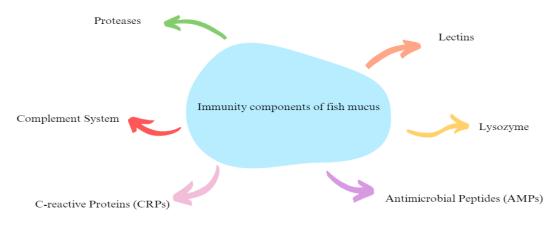
Table-1: List of medicinally important fishes¹.

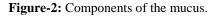
Common name	Species name	Medicinal property	
Snakehead murrel	Channa stritas	The analgesic and antitumor medicinal drug, wound healing properties and reducing postoperative pain, and antimicrobial properties.	
Walking catfish	Clarias batrachus	Reduces inflammation, Diarrhea control, and antinociceptive and antitumor properties.	
Climbing perch	Anabas testudineus	Reduces inflammation, analgesic, antimicrobial and antitumor properties.	
Catla	Catla catla	Asthma, heart diseases, and mineral deficiency.	
Stinging catfish	Heteropneustes fossilis	Antimicrobial, anticancer	
Common wood cat	Trachelypterus galeatus	Umbilical hernia and Asthma.	
Indian glassy fish	Chanda raga	Inflammatory and antimicrobial	
Tunny	Tuna	Anti-inflammatory effect on the cardiovascular system, lowering blood pressure. Control the risk of stroke and heart attacks.	
Tengra	Mystus tengra	Anticancer.	
Common carp	Cyprinus carpio	Antimicrobial.	
Cichild fish	Tilapia	Protect various diseases like Alzheimer's, Parkinson's and epilepsy. Anti- inflammatory effects and novel treatment for Cancer.	
Marbled swamp eel	Synbranchus marmoratus	Bronchitis.	





Local name	Scientific name	Parts used	Application	Disease
Nujung	Anguilla bengalesis	Fats	Fat is applied and massage to relieve pain	Rheumatoid arthritis
Manthu fermented	Puntinus sp.	Whole fish	Whole fish are cooked with bamboo shoots and taken to purify the blood	Blood purifier
Ok-langso	Channa gachua	Whole fish, Bile	Boiled fish is prescribed to eat. The bile of the fish is applied to remove thorns	Abdominal pain when picked by a thorn
Ok-meklot	Channa puntatus	Eyes	Fish eyes are used to remove the corn	Corn or Clavus
Singki	Heteropneustes fossilis	Whole fish	Boiled fish is eaten up as a tonic	Anemia
Kunchirui	Monopterus cuchia	Whole fish, Raw blood	Raw blood is used to remove a leech from the anus	Entry of leech into the anus
Kunchirui	Amphipnous cuchia	Meat blood	Raw blood is consumed and boiled fish is prescribed to eat	Premenstrual abdominal pain, Anemia
Notun	Labeo pangusia	Flesh	Used as a tonic	Weakness
Seketa	Wallago attu	Head	The boiled fish head is taken routinely to improve liver activity	Liver tonic
Kokil mas	Xenenthodon cancila	Fish body	Spine and bone of fish are used to pick out the clotted blood	Joint pain, swelling
Mangur	Claria batrachus	Whole fish	Boiled fish is for regain strength. Cooked fish is consumed to cure smallpox	Smallpox, weakness after delivery
Moa	Amblypharyngodon mola	Whole fish	Boiled fish is prescribed to eat	Premenstrual pain
Tengana	Mystus tengana	Fish body	Used to cure small pox	Small pox
Kanduli mas	Notopterus notopterus	Whole fish	Burned and cooked fish are used	Delivery pain. Abdominal pain
Kurkuri mas	Chaca chaca	Fish body	Whole dried fish is used as food	Polio
Climbing perch	Anabas testudineus	Whole fish	The fish head part is boiled with spices and taken to eat	Dysmenorrhoea





Proteases: Proteases are groups of proteolytic enzymes present of epidermal layer, hydrolysed the peptide bonds present in proteins, converting them to shorter polypeptides and amino acids. Proteases of skin mucus are natural resistance of fish against different types of pathogens¹³, they directly or indirectly act on pathogen or preventing invasion of pathogen by changing mucus consistency and destroy pathogens from the body surface¹⁴. The protease activity of integumentary mucus of L. rohita was comparatively higher than C. mrigala and C. catla, The protease enzyme play active role against the pathogenic organisms on the skin of climbing perch¹⁶. Proteases are also believed to activate and enhance the production of innate immune components like complement, and immunoglobulins in the piscine system. The presence of aminopeptidase, cathepsin B, and L-like proteases in the epidermal cell layer of the Japanese eel (Anguilla japonica) and dorsal surface of the European eel (A. anguilla) was also reported¹⁷.

Lectins: Lectins are special type of protein also present in skin mucus of fish, containing a non-catalytic carbohydraterecognition domain (CRD)¹⁸. Varieties of lectins have been reported in the fish integumentary mucus¹⁹. In catfish, *Silurusasotus* Calcium-dependent mannose-binding activity reported the occurrence of Fructose binding lectin in sea bass skin mucus. lectin (pentraxin) of with molecular mass 25 kDa lectin (pentraxin) was obtained from the integumentary mucus of a cartilaginous fish, *Raja kenojei*by^{20,21}.

Host defense peptides (AMPs): Antimicrobial peptides (AMPs) are a group of peptides, present in nature and part of the innate immune system of a variety of organisms²². It was observed that fishes are a great source of these Peptide and all major classes of peptides like defensins, cathelicidins, hepcidins, histone-derived peptides are present in fish-and called piscidins²³. It was found that the antimicrobial peptides are appear and attached to the membrane surface and displace the outer bilayer²⁴, form the channel and depolarizing the target cells²⁵. Piscidin-1 and piscidin-2 attach with cell membrane surface of the fungus and destroy it²⁶. Fish peptides show antimicrobial activity by preventing both fish and human pathogens²⁷.

A new linear antimicrobial peptide Pelteobagris purified from mucus was found to show antibacterial activity against grampositive and gram-negative bacteria and fungi¹⁸. It has been recently revealed in many studies, that there are several AMPs in the mucus fish that can be potentially used as an antibiotic for humans to cure diseases, especially skin diseases. For example, the hybrid striped was investigated to exhibit strong antifungal activity against human pathogenic fungi. Monocidins are another AMP showed wide spectrum activity against the Fungus, yeast, Gram +ve and Gram -ve bacteria, fungi. A 19 residue AMP parasin was discovered in the mucus of the injured fish and found to be quite similar to Buforin I which are an H2A-derived peptide present in toads²⁸.

Lysozyme: Lysozyme is a lytic enzyme, functioning as antimicrobial agent, break the peptidoglycan component cell walls of bacteria. Enzyme-containing muramic acid is hydrolyses glycol chitin and effect on chitin, present in cell wall and chitin of Invertebrates²⁹. It was also reported that in addition to bacteria lysozyme inhibit viruses³⁰, parasites³¹. Lysozyme was reported in the skin mucus of five marine teleost fish by³² and three freshwater fish highlighting the variations in the considered fish to furnish important information for the aquaculture industry³³.

C-reactive proteins (CRPs): In this study (CRPs), C-Reactive Proteins are a group of proteins (pentraxins) and bind various ligands in a Ca^{2+} -dependent binding capability. C-reactive protein is a part of the innate immune defense activity of the complement pathways and recognition and clearance of apoptotic cells³⁴. Presence of pentraxin-like molecules was also reported in different teleost fish species including common wolfish (*Anarhichas lupus*), Atlantic salmon (*S. salar*), halibut (*H. hippoglossus*), cod (*Gadus morhua*), and Indian major carp (*Catla catla*)³⁵.

Complement System: Protein and non-protein elements of the complement system contain both adaptive immunity and innate. Approximately thirty- five plasma and membrane-bound proteins present in it³⁶. Immune-related complement proteins (C3, C7, and C1q) are reported in the skin mucosa of Atlantic halibut (*H. hippoglossus*), Siberian sturgeon (*Acipenser baerii*) and grass carp (*Ctenopharyngodon idella*), respectively^{37-39.}

Antifungal activity of Fish skin mucus: In this study we review the antifungal activity of skin mucus of various fish species against *Candida brusei*, *Saccharomyces cerevisiae Candida albicans*, *Candida tropicalis*, and *Issatchenkia orientalis*, and find *Gadus morhua* were most active against the fungi⁴⁰.

Antimicrobial activity of epidermal mucus extract of *Maculabatis gerrardi* and *Pastinachus sephen* was also reported againstthe fungal strains *viz.*, *Candida tropicalis, Aspergillus niger, Penicillium sp., Trichophyton mentagrophytes, Alternaria alternata, Candida albicans, Rhizopus sp., Mucor sp.,* and *Trichophyton rubrum* and showed a potent activity also against fungal pathogens⁴¹.

The antifungal activity catfish (*Clarias batrachus*) was tested against pathogenic fungi: *Aspergillus niger, Aspergillus nidulans, Fusarium moniliforme, Candida albicans,* and *Trichoderma koningi,* and maximum zone of inhibition was observed against both *A. niger* and *C. albicans;* followed by *A. nidulans* and *F. moniliforme.* While *T. koningi,* showed the minimum zone of inhibition⁴².

Antifungal activity of skin mucus extracts of Mastacembalus armatus against 6 fungal strains viz. Aspergillus niger, Aspergillus flavus, Candida albicans, Cryptococcus *neoformans*, and *Mucor sp* and it was observed that the acidic mucus extract inhibited the growth of *Mucor sp*. better than the organic extracts. However, the organic mucus extracts of the same fish were found to be more effective against *A. niger* and *A. flavus* than the acidic extracts⁴³.

The antifungal activity of skin mucus of fish *Dasyatis pastinaca* is a great inhibitor of *Candida albicans*, *Candida glabrata*, and *Candida tropicalis* was observed⁴⁴.

The antifungal properties of epidermal mucus of freshwater fish species viz., *Catla catla, Cyperinus carpio,* and *Heteropneustes fossilis* against fungal strains: *Aspergillus niger, A. clavatus, A. flavus, Candida albicans, C. tropicalis, C. auris,* and *Mucor ramosissimus* was also studied and observed that both the acidic and organic extracts showed a vast range of antifungal effects against all the selected pathogenic fungi. However, acidic extracts of all the specimens exhibited a higher antifungal effect against the selected fungal strains. It was considered that the basic peptides were responsible for antifungal action of all fishes. The skin mucus extracts of *C. catla* showed the highest antifungal effect out of the three⁴⁵.

Conclusion

As an alternative to antifungal drugs, it is necessary and also wants attention the animal sources of antifungal components from the above-mentioned studies, it is evident that the fish skin mucus possesses strong antifungal activities against pathogenic fungal strain, it could be noted as a source of novel antifungal components. The mucus is usually discarded from the fisheries and hence is wasted. This skin mucus can be efficiently employed for the antifungal agents, with a low risk of developing resistance, which would be beneficial in the case of drug resistance.

References

- Rajani, N., & Alka, M. (2015). To study the Ethnomedicinal importance of food fish used by localite of Durg. *IOSR J Environ Sci Toxicol Food Technol*, 16, 38-40.
- 2. Gupta, T., & Dey, M. (2017). Ichthyotherapy: use of fishes as medicine by ethnic Karbi people of Assam, India. *European Journal of Pharmaceutical and Medical Research*, 4(10), 341-343.
- **3.** Subramanian, S., MacKinnon, S. L., and Ross, N. W. (2007). A comparative study on innate immune parameters in the epidermal mucus of various fish species. *Comparative biochemistry and physiology. Part B, Biochemistry and molecular biology*, 148(3), 256–263.
- **4.** Cameron, A. M., and Endean, R. (1973). Epidermal secretions and the evolution of venom glands in fishes. *Toxicon*, 11(5), 401-410.
- 5. Dash, S., Das, S. K., Samal, J., and Thatoi, H. N. (2018). Epidermal mucus, a major determinant in fish health: a

review. Iranian journal of veterinary research, 19(2), 72-81.

- 6. Gomez, D., Sunyer, J. O., and Salinas, I. (2013). The mucosal immune system of fish: the evolution of tolerating commensals while fighting pathogens. *Fish and shellfish immunology*, 35(6), 1729–1739.
- 7. Austin, B., and McIntosh, D. (1988). Natural antibacterial compounds on the surface of rainbow trout, Salmo gairdneri Richardson. *Journal of Fish Diseases*, 11(3), 275-277.
- 8. Shephard, K. L. (1994). Functions for fish mucus. *Reviews in Fish Biology and Fisheries*, 4(4), 401-429.
- 9. Cone, R. A. (2009). Barrier properties of mucus. *Advanced Drug Delivery Reviews*, 61(2), 75-85.
- **10.** Shephard, K. L. (1993). Mucus on the epidermis of fish and its influence on drug delivery. *Advanced Drug Delivery Reviews*, 11(3), 403-417.
- **11.** Zaccone, G., Kapoor, B., Fasulo, S., and Ainis, L. (2001). Structural, histochemical, and functional aspects of the epidermis of fishes. *Advances in Marine Biology*, 253-348.
- Brinchmann, M. F. (2016). Immune relevant molecules identified in the skin mucus of fish using -omics technologies. *Molecular Bio. Systems*, 12(7), 2056-2063.
- **13.** Ingram, G. A. (1980). Substances involved in the natural resistance of fish to infection-A review. *Journal of Fish Biology*, 16(1), 23-60.
- Aranishi, F., Mano, N., and Hirose, H. (1998). Fluorescence localization of epidermal cathepsins L and B in the Japanese eel. *Fish Physiology and Biochemistry*, 19, 205-209.
- **15.** Dash, S., Samal, J., and Thatoi, H. (2014). A comparative study on innate immunity parameters in the epidermal mucus of Indian major carps. *Aquaculture International*, 22(2), 411-421.
- **16.** Al-Rasheed, A., Handool, K. O., Alhelli, A. M., Garba, B., Muhialdin, B. J., Masomian, M., Hani, H., and Daud, H. H. (2020). Assessment of some immune components from the Bioactive crude extract derived from the epidermal mucus of climbing perch *Anabas testudines*. *Turkish Journal of Fisheries and Aquatic Sciences*, 20(10), 755-766.
- **17.** Aranishi, F., and Nakane, M. (1997). Epidermal proteases of the Japanese eel. *Fish Physiology and Biochemistry*, 16, 471-478.
- **18.** Bhatnagar, A., and Rathi, P. (2021). Fish skin mucus as a putative bio-resource for the development of next-generation antibiotics. *Egyptian Journal of Aquatic Biology and Fisheries*, 25(5), 1063-1091.
- **19.** Tsutsui, S., Okamoto, M., Ono, M., Suetake, H., Kikuchi, K., Nakamura, O., Suzuki, Y., and Watanabe, T. (2011). A new type of lectin discovered in a fish, Flathead

(*Platycephalus indicus*), suggests an alternative functional role for mammalian plasma kallikrein. *Glycobiology*, 21(12), 1580-1587.

- **20.** Cordero, H., Brinchmann, M.F., Cuesta, A., Meseguer, J., and Esteban, M.A. (2015). Skin mucus proteome map of European sea bass (*Dicentrarchuslabrax*). *Proteomics*. 15, 4007-4020.
- Tsutsui, S., Yamaguchi, M., Hirasawa, A., Nakamura, O., & Watanabe, T. (2009). Common skate (Raja kenojei) secretes pentraxin into the cutaneous secretion: the first skin mucus lectin in cartilaginous fish. *Journal of biochemistry*, 146(2), 295-306.
- 22. Huan, Y., Kong, Q., Mou, H., and Yi, H. (2020). Antimicrobial Peptides: Classification, Design, Application and Research Progress in Multiple Fields. *Frontiers in microbiology*, 11, 582779.
- Valero, Y., Elena, CP., Jose, M., Maria, A.E., and Alberto, C. (2013). Biological role of fish antimicrobial peptides. In: Seong, MD and Hak, YI (Eds.), Antimicrobial peptides. (1st Edition.), New York, USA. *Nova Science Publishers Inc.*, 31-60.
- 24. Shai, Y. (1994). Pardaxin: channel formation by a shark repellant peptide from fish. *Toxicology*, 87, 109-129.
- 25. Lee, S. A., Kim, Y. K., Lim, S. S., Zhu, W. L., Ko, H., Shin, S. Y., Hahm, K. S., and Kim, Y. (2007). Solution structure and cell selectivity of piscidin 1 and its analogues. *Biochemistry*, 46(12), 3653–3663.
- **26.** Sung, W. S., Lee, J., and Lee, D. G. (2008). Fungicidal effect and the mode of action of piscidin2 derived from hybrid striped bass. *Biochemical and biophysical research communications*, *371*(3), 551–555.
- 27. Das, S.K., Samal, J., and Dash, S. (2013). Antimicrobial activity of skin mucus of fishes: a review. In: Thatoi, HN and Mishra, BB (Eds.), *Advances in biotechnology*. (1st Edition.), USA, Studium Press. 491-506.
- **28.** Park, C. H., Valore, E. V., Waring, A. J., and Ganz, T. (2001). Hepcidin, a urinary antimicrobial peptide synthesized in the liver. *Journal of Biological Chemistry*, 276(11), 7806-7810.
- **29.** Wu, M., Maier, E., Benz, R., and Hancock, R. E. (1999). Mechanism of interaction of different classes of cationic antimicrobial peptides with planar bilayers and with the cytoplasmic membrane of *Escherichia coli*. *Biochemistry*, 38(22), 7235-7242.
- 30. Lee-Huang, S., Huang, P. L., Sun, Y., Huang, P. L., Kung, H., Blithe, D. L., and Chen, H. (1999). Lysozyme and RNases as anti-HIV components in β-core preparations of human chorionic gonadotropin. *Proceedings of the National Academy of Sciences*, 96(6), 2678-2681.
- **31.** Leon-Sicairos, N., Lopez-Soto, F., Reyes-Lopez, M., Godinez-Vargas, D., Ordaz-Pichardo, C., and De la

Garza, M. (2006). Amoebicidal activity of milk, apolactoferrin, sIgA and lysozyme. *Clinical Medicine and Research*, 4(2), 106-113.

- **32.** Guardiola, F. A., Cuesta, A., Arizcun, M., Meseguer, J., and Esteban, M. A. (2014). Comparative skin mucus and serum humoral defense mechanisms in the teleost gilthead seabream (*Sparus aurata*). *Fish and Shellfish Immunology*, 36(2), 545-551.
- **33.** Sridhar, A., Manikandan, D. B., Palaniyappan, S., Sekar, R. K., and Ramasamy, T. (2021). Correlation between three freshwater fish skin mucus Antiproliferative effect and its elemental composition role in bacterial growth. *Turkish Journal of Fisheries and Aquatic Sciences*, 21(05), 233-244.
- **34.** Nauta, A. J., Daha, M. R., Kooten, C. V., and Roos, A. (2003). Recognition and clearance of apoptotic cells: A role for complement and pentraxins. *Trends in Immunology*, 24(3), 148-154.
- **35.** Lund, V., and Olafsen, J. A. (1998). A comparative study of pentraxin-like proteins in different fish species. *Developmental and Comparative Immunology*, 22(2), 185-194.
- **36.** Boshra, H., Li, J., and Sunyer, J. (2006). Recent advances on the complement system of teleost fish. *Fish and Shellfish Immunology*, 20(2), 239-262.
- **37.** Magnadottir, B., Lange, S., Gudmundsdottir, S., Bogwald, J., and Dalmo, R. (2005). Ontogeny of humoral immune parameters in fish. *Fish and Shellfish Immunology*, 19(5), 429-439.
- **38.** Shen, Y., Zhang, J., Xu, X., Fu, J., and Li, J. (2012). Expression of complement component C7 and involvement in innate immune responses to bacteria in grass carp. *Fish and Shellfish Immunology*, 33(2), 448-454.
- **39.** Fan, C., Wang, J., Zhang, X., and Song, J. (2015). Functional C1q is present in the skin mucus of Siberian sturgeon (*Acipenser baerii*). *Integrative Zoology*, 10(1), 102-110.
- **40.** Hellio, C., Pons, A. M., Beaupoil, C., Bourgougnon, N., and Gal, Y. L. (2002). Antibacterial, antifungal, and cytotoxic activities of extracts from the fish epidermis and epidermal mucus. *International Journal of Antimicrobial Agents*, 20(3), 214-219.
- **41.** Vennila, R., Kumar, K. R., Kanchana, S., Arumugam, M., Vijayalakshmi, S., and Balasubramaniam, T. (2011). Preliminary investigation on the antimicrobial and proteolytic property of the epidermal mucus secretion of marine stingrays. *Asian Pacific Journal of Tropical Biomedicine*, 1(2), S239-S243.
- **42.** Loganathan, K., Muniyan, M.K., Prakash, A.A., Raja, P.S., and Prakash, M.S. (2011). Studies on the role of mucus from *Clarias batrachus* (Linn) against selected microbes.

International Journal of Pharmaceutical Applications, 2(3) 2011, 202-206.

- **43.** Uthayakumar, V., Ramasubramanian, V., Senthilkumar, D., Priyadarisini, V. B., and Harikrishnan, R. (2012). Biochemical characterization, antimicrobial and haemolytic studies on skin mucus of freshwater spiny eel *Mastacembelus armatus. Asian Pacific Journal of Tropical Biomedicine*, 2(2), S863-S869.
- **44.** Fuochi, V., Li Volti, G., Camiolo, G., Tiralongo, F., Giallongo, C., Distefano, A., Petronio Petronio, G.,

Barbagallo, I., Viola, M., Furneri, P., Di Rosa, M., Avola, R., and Tibullo, D. (2017). Antimicrobial and antiproliferative effects of skin mucus derived from *Dasyatispastinaca* (Linnaeus, 1758). *Marine Drugs*, 15(11), 342.

45. Kumari, S., and Yadav, S. (2020). Study of Antifungal Activity of Epidermal Mucus of Three Fresh Water Fishes. *Annals of Biology*, 36(1), 75-80.