

Cowpea emergence response to cadmium stress

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Abstract

Cadmium been the utmost prominent and extensively distributed heavy metal in Nigeria soils. This study investigated the emergence of cowpea accessions (TVu-91,92,93,95 and 96) to cadmium stress in Nigeria soils. The experimental setup consisted of 3 treatments namely 0ESV, 2.5ESV and 5ESV (Ecological Screening Value) and the following parameters were taken; time to 50% emergence, height of emergence, hypocotyl length, terminal leaf breadth and length, fresh and dry weight matter. Cadmium caused a significant decrease in the studied parameter. In TVu-91, cadmium pollution resulted to 2-days delay in time to 50% emergence between the control and 5ESV. The emergence height of TVu-93 sown in Cd-5ESV was 10.67cm as compared to 12.10cm in the control, the hypocotyl length was also reduced from 3.7 to 2.3cm with increased concentration. Terminal leaf length and breadth followed similar trend of reduction. However, the various accessions responded differently as there were insignificantly difference observed in the emergent height of TVu-93 between the treatments. In conclusion, although cadmium pollution reduced the emergence productivity of cowpea accession, however its effect varied within the accessions. This suggests that the presence of the genetic makeup of the individual accessions have the ability to withstand cadmium stress.

Keywords: Pollution, heavy metal, germination, soil toxicity, plant physiology.

Introduction

The environmental and human health implications of heavy metal is owing to their prevalent existence, tenacious in ecosystems, and noxious properties¹. Some of this heavy metals e.g. copper and zinc are necessary elements in plants growth and development. Zinc is significant to plant for production of seeds and in disease resistibility while Cu is importance in the metabolic processes of most plants. Nonetheless, some heavy metal existing in little concentrations are detrimental to plants. One of such heavy metal is cadmium and is commonly found as a contaminant in Nigerian soils². Cadmium is extremely poisonous to all biota and is mainly one of the greatest toxic metal in plants' because of its great dissolvability in H₂O and phytotoxicity³. Cadmium is has no relevance in the nutrient cycle of plants and when in high amount impedes growth⁴; even when comparatively minute, it modifies plants metabolism⁵.

There are various natural and anthropogenic sources that adds heavy metals in to the environment. In natural processes, volcanic upsurges is one of the main source of Cd, however the quantity of the metal released is relatively in significant in contrast to anthropogenic processes. While anthropogenic sources includes; industrial waste, fossil fuel combustion, waste ignition, phosphate fertilizers, among others are the largest contribution of Cd to the environment. A great consideration has been given to cadmium in both plant nutrition and soil science because of its phytotoxic effect which varies from growth reduction⁴, wilting, chlorosis⁶ and cell death⁷.

They are ubiquitous, persistence, non-biodegradable and accumulate in the environment. This implies that living organisms are continuously exposed to Hms⁸ throughout their life cycle, resulting in toxicity⁹. Plants however, depend on some mixture of nutrients found in soil and air, as well as the solar energy, CO₂, and water for photosynthetic processes¹⁰. Food chain pollution by metal is a burning concern in recent years due to their impending build up in bio system through polluted soil, H₂O and air⁴. The existence of plants is hinged on their capacity to identify the impetus, transmit signals and initiate defense mechanisms that regulate their metabolism accordingly¹¹.

Cowpea, *Vigna unguiculata* (L.) Walp, also popularly called 'beans' in Nigeria is an essential food crop (legume) and resourceful plant, mainly nurtured in the dry savannas regions of tropical Africa, including south America and Asia, spanning over 12million ha with a yearly production surpassing 3milliontones¹². Around 64% of the cultivated areas used for cowpea is in Central and West Africa where the crop is mainly planted in combination with sorghum, millet and maize usually in Sahelian and Sudan savanna region. Cowpea serve as a source of cash income and protein supplement in the daily life of people in Nigeria.

Nutritionally, the pods, leaf and green peas are eaten as food, with the green including the dry haulms used as livestock fed. Nigeria been the biggest producer and consumer of cowpea in the world¹³.

In Nigeria, cowpea is mostly grown in the north which characterized with the savanna type of vegetation and light rainfall, with Borno State being the major producer¹⁴. Owing to several limitations the amount of cowpea produce in West Africa was shown to be below 358kg/ha¹⁵ however Singh *et al.*¹⁶ projected 240kg/ha cowpea produce as an average in Nigeria. Some of these constraints include; insect pests infestation,¹⁷ diseases¹⁸, heavy metals¹⁹, insufficient information of ideal ethnical practices and great yielding varieties causing reduced yields²⁰. Studies showed the efficiency of cowpea to produce in such structures can be improved over the use of better-quality varieties, suitable date of planting, sophisticated plant population, enhanced soil richness, and appropriate spatial arrangements²¹. Recently, the International Institute of Tropical Agriculture (IITA) Ibadan, Nigeria developed some enhanced cultivars of cowpea for the savanna ecological zones. These cultivars are said to be superior to the local cultivars in many aspects (high yielding, earlier maturity, resistant to major pests and diseases). Attention is now being given to developing cultivars that can resist environmental stress. Therefore, the aim of this study was to examine the effects of cadmium stress on emergence parameters of 5 cowpea accession (TVu91, TVu92, TVu93, TVu95 and TVu96) obtained from IITA.

Materials and methods

Seed collection: The cowpea seed and its respective accession namely; TVu91, TVu92, TVu93, TVu95 and TVu96 were procured from the International Institute of Tropical Agriculture (IITA), Ibadan, Nigeria.

Metal: The experimental cadmium used was cadmium chloride procured from Pyrex chem. Lab. in Benin, Nigeria.

Experimental setup: The experimental setup was performed in the Botanic farm of the Dept. Plant Biology and Biotech., UNIBEN, Benin city. The soils were also collated here from ten random spots and mixed together to obtain composite sample. Soil samples were collected randomly in the site at a 0–30cm depths by the use of a soil auger. The soil samples were mixed together into a composite sample, sun-dried, grind to pass through a 2mm sieve before sending to the laboratory for physic-chemical examination using to standard techniques. The soils were afterward contaminated with cadmium based on its ecological screening value (ESV). Three concentrations were obtained; 0 ESV, 2.5ESV and 5 ESV.

Sowing: This comprised of 3 blocks with 15 treatments per block, 1 plant per polybag resulting to 45 plants organized in a random complete block design (RCBD). The experimental setup was left for 14weeks and parameters taken. The height of emergence, hypocotyl length, petiole length, terminal leaf length and breadth were measure using a meter rule in centimeter while stem width was measured in millimeters, the fresh weight was by weighing in grams while the dry matter was oven dried at 70°C according to Ekpo and Ebeagwu²². Time to 50%

emergence (T50) is the days needed for 50% emergence of the total number of seeds²³.

Table-1: Treatment designations for metal concentrations.

Designations	Description	Replications
0 ESV	Control (unpolluted soil)	3
2.5 ESV	0.15g of cadmium chloride diluted in 2L of water and mixed in 15kg soil	3
5 ESV	0.30g of cadmium chloride diluted in 2L of water and mixed with 15kg soil	3

Statistical analysis: The data was subjected using statistics to one way analysis of variance (ANOVA). The least significant difference (LSD) were used to separate any difference in their means. Results were presented as means and standard error (n= 7). The significance of the results was ascertained at $p < 0.05$. The SPSS-20® software was employed to run analyses of data.

Results and discussion

Effects of cadmium on Time to 50% Emergence: In this study, the effects of cadmium on 50% emergence time of cowpea accession (TVu91, TVu92, TVu93, TVu95 and TVu96) have been reported (Figure-1). TVu-95 sown in the control soils had a 50% mean emergence time speed of 4.20 days as compared to a delay of 6.30 days in TVu-95 sown in Cd-5ESV soil. The prolonged delayed difference of 3.00 days also reported in time to 50% emergence of TVu-93 between the Cd-5ESV and control conforms to those observed by other researchers²⁴. For example, Rahman²⁵ detected a reduced germinated seed and seedling development in chickpea exposed to 50, 100, 200 and 400ppm of nickel and cobalt. Singh *et al.*²⁶ reported a decline in germination percent and initial growth period of wheat exposed to copper at different levels. This shows that cadmium polluted soils affects time to 50% emergent of cowpea accessions, though there was no significant difference ($P=0.17$). Metal toxicity is a vital issue governing sprouting and growth of plants. Energy generation is vital for seed emergent and any impediment can disturb protein, DNA and RNA production including mitosis due to the fact that energy is a prerequisite for these processes to occur²⁷.

Effects of cadmium on height of emergent: The effects of cadmium pollution on cowpea resulted in a reduction in emergent height (Figure-2). Germination and initial seedling sprouting evaluation is regarded as an elementary experiment for assessing the toxic effects of any heavy metal or element type on plants^{28,29}. There was a decrease in seedling emergence height with increased metal concentration. Take for instance, TVu-93 sown in Cd-2.5ESV and Cd-5ESV had an emergent height value of 11.47cm and 10.67cm respectively as compared to 12.10cm in the control. Seedling growth is agreed as a

relevant pointer of metal effect on plants' capacity to existence. However, TVu-92 was more resilient to Cd pollution than the other studied accessions. Hence, cadmium pollution was highly significantly different ($P=0.000$). Some researchers has stated the decline of emergence and seedling growth of difference crops exposed to metals toxicity. This was not far from the

present study were the emergence height and hypocotyl length reduced with increase in Cd concentration. Take for instance, TVu-91 reduced from 11.62 to 10.30cm with increased metal toxicity. This can be due to the noxious effects of ions on the emergence process^{30,31}.

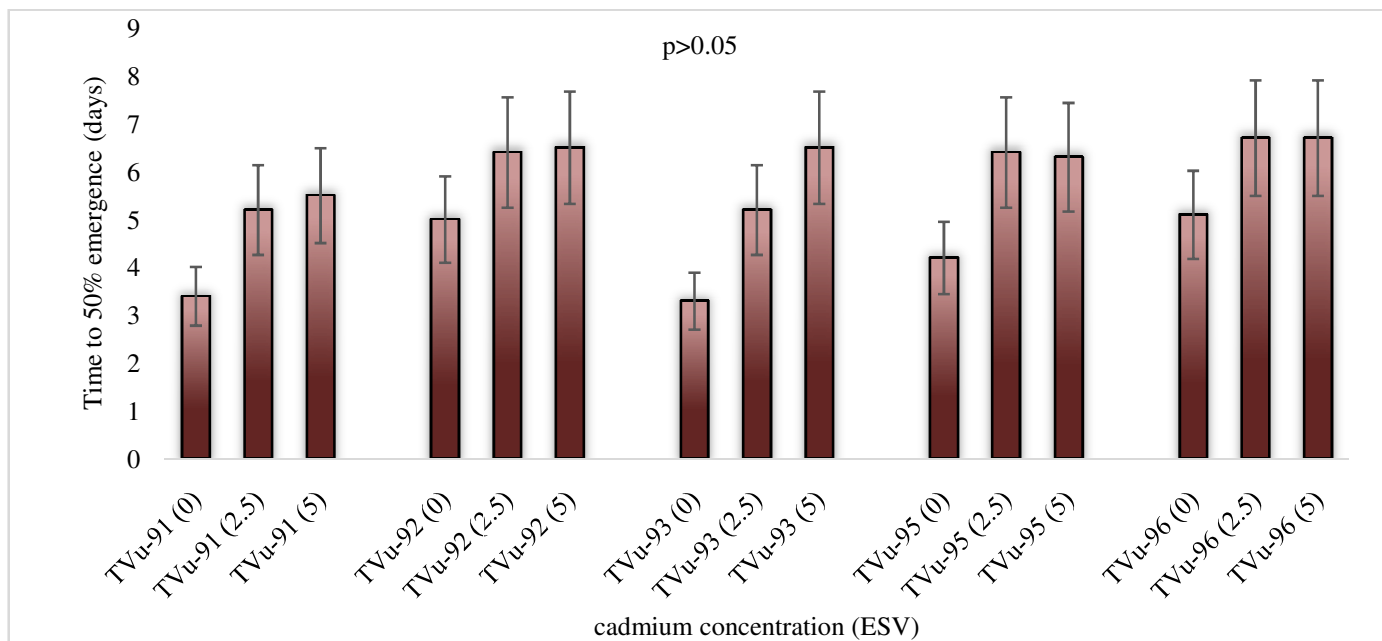


Figure-1: Time to 50% emergence (days) of cowpea.

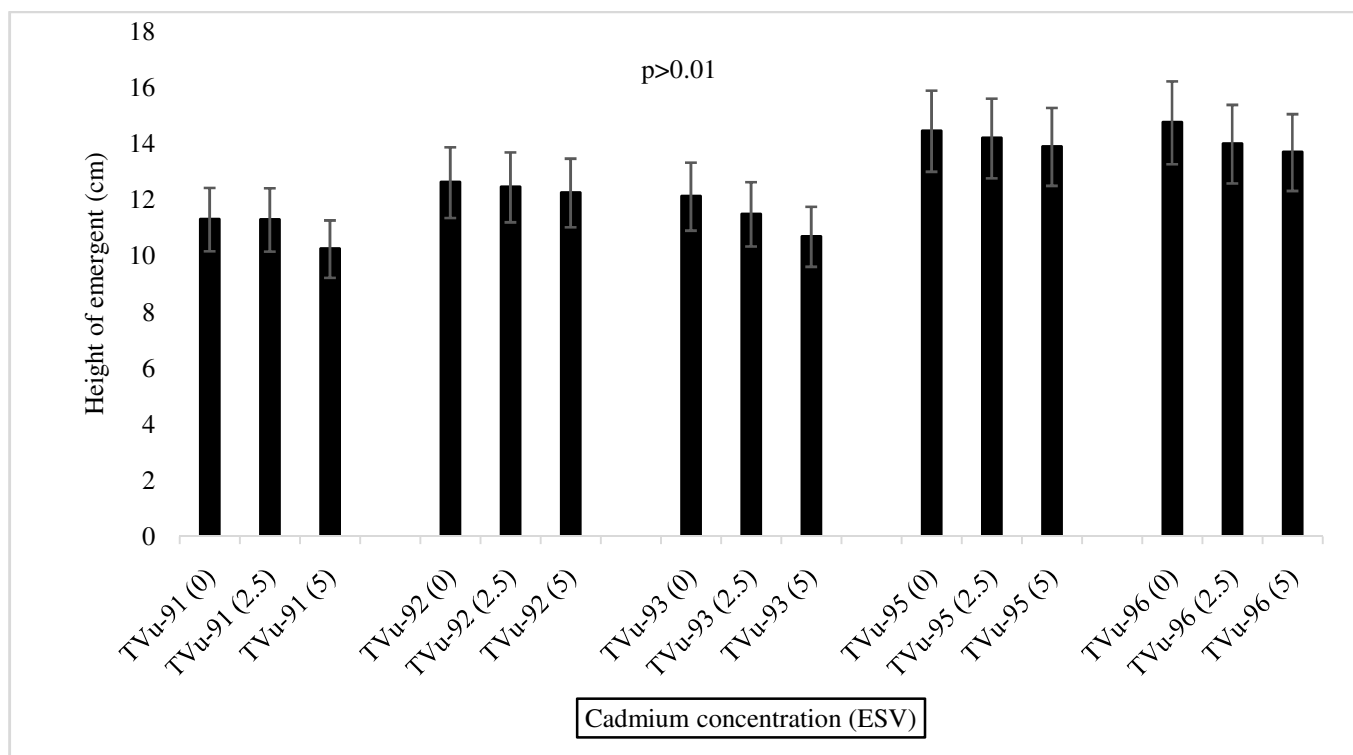


Figure-2: The height of emergent of cowpea.

Effects of cadmium on fresh and dry weight of emergent:

Figure-3 shows the fresh weight performance of emerged cowpea accessions to various concentration of cadmium. In this study, Cd significantly reduced both fresh and dry weight of emergence over the control. Fresh weight of emergent in TVu-91 sown in Cd-5ESV was 0.65g as compared to an increased 1.35g in TVu-91 in the control. The existence of inhibitory compounds in greater concentrations of the metal may be the cause for the various performance of the extracts and extreme reduction in the emergence parameter³². The plant dry weight matter followed similar trend of reduction with increase in metal concentration. TVu-91 was more susceptible to cadmium pollution than TVu-92. This decline in biomass can be credited to the inhibition of chlorophyll synthesis and photosynthesis by the metal. These results is in line with Singh *et al.*²⁶.

Effect of cadmium on hypocotyl length: The hypocotyl length of cowpea accession to cadmium pollution has been reported (Figure-4). Cadmium pollution resulted to a decrease of 1.4cm in the hypocotyl length of TVu96 between Cd-5ESV and control. This indicated that increased Cd concentration, decreases the hypocotyl length of the cowpea accessions. However, cadmium pollution has no significant difference (P=0.07) on the hypocotyl length of various cowpea accessions. The process of germination like emergence height and

hypocotyl length will be suppressed. This findings is in agreement with previous studies carried out^{4,6}.

Effects of Cd on Terminal leaf length and breadth: Figure-5 shows the consequence of cadmium pollution on the terminal leaf length and breadth of cowpea accessions. There was a reduction in terminal leaf breadth in all treatments with TVu-96 sown in Cd-5ESV having a value of 3.9cm as compared to 4.9cm in TVu-96 sown in the control soils. Growth decrease in Cd environments was observed for several species tested³³⁻³⁵. In this study, a significant reduction was reported for the hypocotyls, terminal leaf length and breadth particularly in the 50% emergence time radicles. Hence, the findings provide further backing to the previous research. Hence, a significant difference (P=0.009) exist on the terminal leaf breadth of the studied accessions to cadmium pollution.

Conclusion

From the findings, Cd pollution induce growth inhibition in cowpea seedlings, especially at 50% emergence time. However, the various accessions responded differently as there was little to no difference in the height of emergence of TVu-95. The observed variation in the cowpea seedlings response is strongly dependent on the intrinsic characteristics inherent within the accessions type.

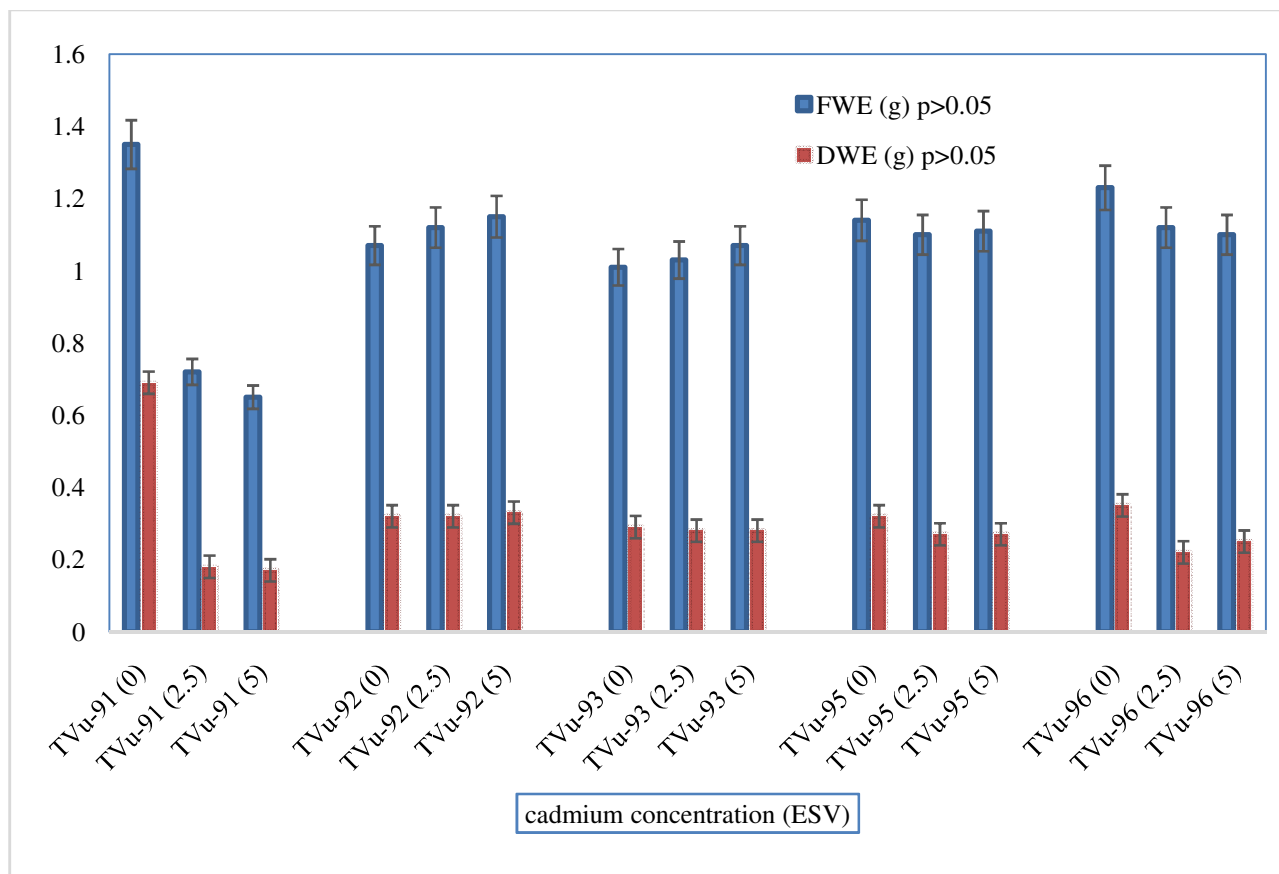


Figure-3: Fresh weight of emergent (FWE) and dry weight of emergent (DWE) of cowpea.

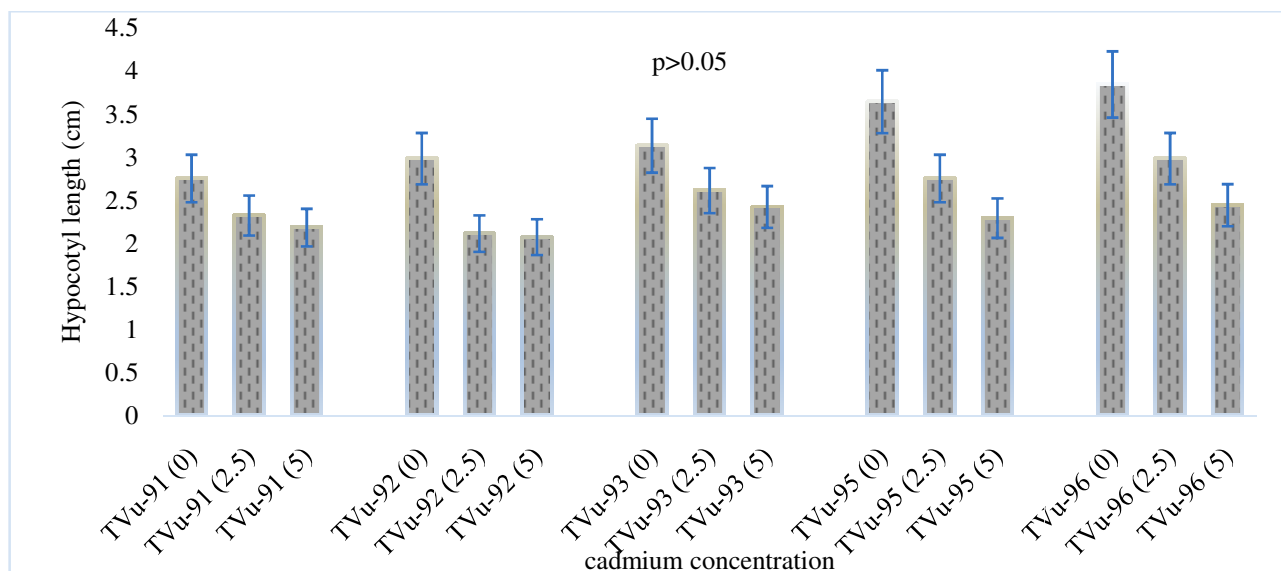
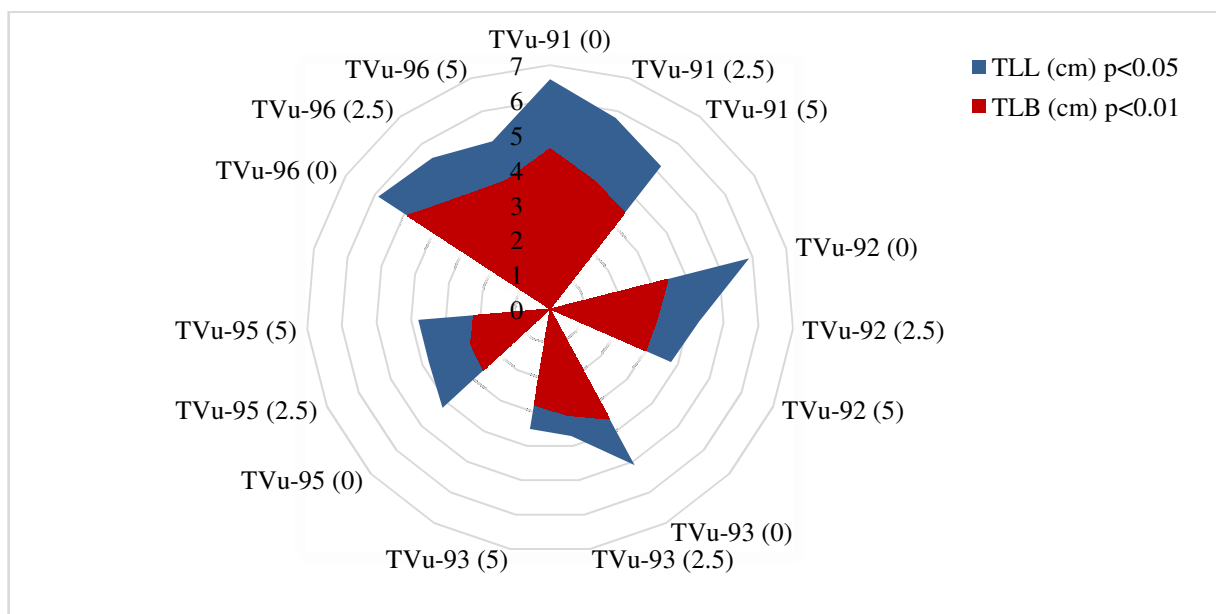


Figure-4: Hypocotyl length of cowpea accession in cadmium polluted soils 7 days after sowing.



NB: TLL - Terminal leaf length (cm), TLB - Terminal leaf breadth (cm)

Figure-5: TLL and TLB of cowpea accessions to cadmium polluted soils 14 days after sowing.

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References

1. Cui Y.J., Zhu Y.G., Zhai R.H., Huang Y., Qiu Y., Liang J.Z., Zengin F.K. and Munzuroglu O. (2005). Effects of some Heavy Metals on Content of Chlorophyll, Proline and some Antioxidant Chemicals in Bean (*Phaseolus vulgaris* L.) Seedlings. *Acta Biologica Cracoviensia Series Botanica*, 47(2), 157-164.
2. Galadima A., Garba Z.N., Leke L., Almustapha M.N. and Adam I.K. (2011). Domestic water Pollution among Local Communities in Nigeria - causes and consequences. *European J. Scientific Research*, 52(4), 592-603.
3. Ohanmu E.O. and Ikhajiagbe B. (2018). Enzymatic and Non-Enzymatic response of *Sphenostylis stenocarpa* to Cadmium Stress. *Asian J. Applied Sci.*, 11, 125-134.
4. Ohanmu E.O., Ikhajiagbe B. and Anoliefo G.O. (2017). Assessment of Growth and Yield responses African Yam

- Bean (*Sphenostylis stenocarpa*) to Cadmium Pollution. *Nig. J. Life Sci.*, 7(2), 166-180.
5. Van-Assehe F. and Clijsters C.P.H. (1990). Effects of Metals on Enzyme Activity in Plants. *Plant Cell Environmental*, 13(3), 195-206.
 6. Ikhajiagbe B., Anoliefo G.O., Ohanmu E.O. and Aliu E. (2018). Effects of different Cadmium Levels on the Growth and Yield Parameters of Wild *Vigna*. *Studia Universitatis Babeş-Bolyai Biologia, LXIII*, 2, 169-182.
 7. Gallego S.M., Pena L.B., Barcia R.A., Azpilicueta C.E., Iannone M.F. and Rosales E.P. (2012). Unravelling Cadmium Toxicity and Tolerance in Plants: Insight into Regulatory Mechanisms. *Environmental Experimental Botany*, 83, 33-46.
 8. Yeuka M., Sanele M. and Norah B. (2017). The Effects of Sub Lethal Levels of Lead on Acetylcholinesterase Activity in the Rock Pigeon (*Columba Livia*). *Zimbabwe J. Sci. Techn.*, 12, 1-7.
 9. Espin S., Martínez-Lopez E., Jimenez P., Maria-Mojica P. and García-Fernández A.J. (2014). Effects of Heavy Metals on Biomarkers for Oxidative Stress in Griffon Vulture (*Gyps fulvus*). *Environmental Research*, 129, 59-68.
 10. Ikhajiagbe B. (2016). Possible Adaptive Growth responses of *Chromolaena odorata* during Heavy Metal Remediation. *Ife Journal of Sci.*, 18(2), 403-411.
 11. Ohanmu E.O. and Ikhajiagbe B. (2018). Effect of Cadmium Pollution on Nitrogen Assimilation and Bioaccumulation of *Vigna unguiculata* L. *Asian J. Applied Sci.*, 11, 183-191.
 12. IITA. (1999). Cowpea-Cereals Systems Improvement in the Savannas. Annual Report of the International Institute of Tropical Agriculture, Ibadan, Nigeria.
 13. Singh B.B. (2007). Potential and Constraints of improved Cowpea Varieties in increasing the Productivity of Systems in the Dry Savannas of West Africa. 14.
 14. Kamara A.Y., Chikoye D., Omoigui L.O. and Dugje I.Y. (2007). Influence of Insecticide spraying Regimes and Cultivar on Insect-Pests and Yield of Cowpea in the Dry Savannas of North-Eastern Nigeria. *J. Food, Agriculture and Environment*, 5(1), 154-158.
 15. FAO. (2000). Site internet: <http://www.fao.org/statistics>.
 16. Singh B.B., Chamblis O.L. and Sharma B. (1997). Recent Advances in Cowpea Breeding. In Cowpea research, (ed.) Singh B.B., Mohan-Raj D.R., Dashiel K.E. and Jackai L.E.N. Co-publication of International Institute of Tropical Agriculture (IITA) and Japan International Research Centre for Agricultural Sciences (JIRCAS). IITA, Ibadan, Nigeria, 30.
 17. Thio I.G., Zida E.P., Sawadogo M. and Sérémé P. (2016). Current status of *Colletotrichum capsici* strains, causal agents of Brown blotch disease of cowpea in Burkina Faso. *African Journal of Biotechnology*, 15(5), 96-104.
 18. Abdulai M., Kusi F., Seini S.S., Seidu A., Nboyine J.A. and Larbi A. (2017). Effects of planting Date, Cultivar and Insecticide spray Application for the Management of Insect-Pests of Cowpea in Northern Ghana. *Crop Prot.*, 100, 168-176.
 19. Ohanmu E.O., Ikhajiagbe B. and Edegbai B.O. (2018). Nitrogen Distribution pattern in African Yam Bean (*Sphenostylis stenocarpa*) exposed to Cadmium Stress. *J. Applied Sci. Environ. Management*, 22(7), 1053-1057.
 20. Kanteh S.M., Norma J.E. and Sherman K.J. (2014). Effect of Plant Density and Weeding Regime on Population and Severity of Aphids (*Aphis craccivora* Koch) and Foliage Beetles (*Ootheca mutabilis* Sahl) on Cowpea in Sierra Leone. *Int. J. Agric. For.*, 4, 24-33.
 21. Kamai N., Kamara A.Y. and Omogui I.O. (2014). Varietal Trials and Physiological Basis for Yield Differences among Cowpea Varieties in Sudan Savanna of Nigeria. *International J. Agriculture Innovations and Research*, 2(5), 855-859.
 22. Ekpo M.A. and Ebeagwu C.J. (2009). The Effect of Crude Oil on Microorganisms and Dry Matter of Fluted Pumpkin (*Telfaria occidentalis*). *Sci. Research Essay*, 4(8), 733-739.
 23. Josep A.R. and Maria M. (2002). Seed Germination and Reproductive Features of *Lysimachia minoricensis* (Primulaceae), a Wild Extinct Plant. *Annals of Botany*, 89(5), 559-562.
 24. Talebi S., Nabavi-Kalat S.M. and Sohani-Darban A.L. (2014). The Study Effects of Heavy Metals on Germination Characteristics and Proline Content of Triticale (*Triticoseale Wittmack*). *International J. Farming and Allied Sci.*, 3(10), 1080-1087.
 25. Rahman K.M. and Mahmud K.M. (2010). Effect of varying Concentration of Nickel and Cobalt on the Plant Growth and Yield of Chickpea. *Australian J. Basic Applied Sci.*, 4(6), 1036-1046.
 26. Singh D., Nath K. and Kumar S.Y. (2007). Response of Wheat Seed Germination and Seedling Growth under Copper Stress. *J. Environmental Biology*, 28(2), 409-414.
 27. John M.K. and Van-Laerhoven C.J. (1976). Differential Effects of Cadmium on Lettuce Varieties. *Environmental Pollution*, 10(3), 163-173.
 28. Ahsan N., Lee D.G., Lee S.H., Kang K.Y., Lee J.J., Kim P.J., Yoon H.S., Kim J.S. and Lee B.H. (2007). Excess Copper induced Physiological and Proteomic changes in Germinating Rice Seeds. *Chemosphere*, 67(6), 1182-1193.
 29. Wang M. and Zhou Q. (2005). Single and joint Toxicity of Chlorimuronethyl, Cadmium and Copper acting on Wheat (*Triticum aestivum*). *Ecotoxicology and Environmental Safety*, 60, 169-175.

30. Khajeh-Hosseini M., Powell A.A. and Bingham I.J. (2003). The interaction between Salinity Stress and Seed Vigour during Germination of SOYBEAN seeds. *Seed Sci. Technology*, 31, 715-725.
31. Kiran Y. and Munzuroglu O. (2004). The effects of Lead on the Seed Germination and Seedling Growth of Lens (*Lens culinaris* Medic.). *Firat University J. Sci. Engineering*, 16(1), 1-9.
32. Shukla A.K., Prasad S., Srivastava S.K., Singh S.P. and Singh R.P. (2003). Allelopathic Effect of Thatch Grass (*Imperata cylindrica* L.) on various Kharif and Rabi Season Crops and Weeds. *Indian J. Weed Sci.*, 35, 163-166.
33. Scebba F., Arduini I., Ercoli L. and Sebastiani L. (2006). Cadmium Effects on Growth and Antioxidant Enzymes Activities in *Miscanthus sinensis*. *Biologia Plantarum*, 50(4), 688-692.
34. Abu-Muriefah S.S. (2008). Growth Parameters and Elemental Status of Cucumber (*Cucumis sativus*) Seedlings in response to Cadmium Accumulation. *International J. Agriculture and Biology*, 10(3), 261-266.
35. John R., Ahmad P., Gadgil K. and Sharma S. (2008). Effect of Cadmium and Lead on Growth, Biochemical Parameters and Uptake in *Lemna polyrrhiza* L. *Plant, Soil and Environment*, 54, 262-270.