



## Review Paper

# Environmental impact of engineered nanomaterials

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## Abstract

*Advancements in the field of nanotechnology, have led to a concomitant rise in the incorporation of nanomaterials in consumer products. Engineered nanomaterials today are already being used in diverse commercial products in the fields of energy, sensing, food technology, electronics, pharmaceuticals, cosmetics, and material applications and have an estimated global market value of €20 billion. This has given rise to concerns about the undesirable effects of this technology on the environment. This review presents an overview of published studies about likely impact of nanoparticles in the ecosystem, their ecotoxicology, threat to human health and the environment and lack of sufficient data in the Indian context.*

**Keywords:** Engineered nanomaterials, toxicity.

## Introduction

Nanoparticles have been around for millions of years but our increasing ability to manipulate matter at the nanoscale has brought nanotechnology research into focus globally<sup>1,2</sup>. Roco et al predicted a global increase in the number of people working in the field of nanotechnology to an estimated total of 6 million by 2020<sup>3</sup>. This rapid development in nanotechnology has created new opportunities for high performance applications and new product innovation. However, there is a growing concern that the handling and disposal of engineered nanomaterials, may result in new, undesirable impacts on health and the environment<sup>4,6</sup>. Governments worldwide have started implementing policies and regulations for evaluating the risks of nanomaterials on health and environment<sup>7</sup>. Several reviews have focussed on the problems and risks associated with engineered nanomaterials<sup>6,8-10</sup>.

## Engineered Nanoparticles

The term 'nanomaterial' includes a range of materials that show unique properties due to their small size<sup>11</sup>. As per the definition adopted by the European Union "engineered nanomaterial" is any intentionally manufactured material, containing particles, in an unbound state or as an aggregate or as an agglomerate and where, for 50% or more of the particles in the number size distribution, one or more external dimensions is in the size range 1 nm to 100 nm<sup>12</sup>.

Extensive research has been carried out on the toxicology and health implications of nanoparticles, a majority of which is related to the impact of fullerenes, carbon nanotubes, quantum dots and ZnO and titania nanoparticles<sup>12-14</sup>. But data on nanoparticle toxicity is largely heterogeneous and also conflicting<sup>14,15</sup>. Understanding and interpretation of results is

complicated because different types of tests have been used to study toxicity<sup>16</sup>.

Difficulties in evaluating their potential impact arise because interactions of nanomaterials with the environment may often give indistinct signals<sup>17,18</sup>. Although characterization of nanomaterials with high degree of resolution is now achievable and some of these techniques maybe used to detect ENPs in water and soil samples but identification and quantification of ENPs in the environment still pose a challenge<sup>19,20</sup>. Further difficulties in detection of nanoparticles arise due to their interaction with other contaminants, influencing their ecotoxicity. Chemical transformations of nanoparticles in the environment and within living systems change their properties and reactivity<sup>21</sup>. Complications in characterization of engineered nanoparticles in natural environments arise also due to the fact that many engineered nanoparticles have their natural counterparts with the same composition.

Several papers outline the physicochemical principles that govern particle behaviour in an ecotoxicological context and some techniques that may be used to trace and quantify ENPs in various biological and chemical matrices<sup>22,23</sup>. Roth et al, and White et al have used Hyperspectral imaging (HSI) to evaluate the interaction of optically active nanoparticles in biological media and cells<sup>24,25</sup>.

## Toxicity of Engineered Nanoparticles

Small size, large surface area and enhanced reactivity are some of the features that are responsible for the widespread applications of nanomaterials but these very features also increase the possibility for them to cross cell membranes and enter inside cell organelles unlike bulk materials of the same

chemical composition<sup>26,27</sup>. Results of various in vivo and in vitro studies carried out to compare the behaviour of nanoparticles with their bulk counterparts point to their enhanced toxicity. Several studies focus on the absorption, incorporation and translocation of nanoparticles and its effects in organisms<sup>28</sup>. Metal nanoparticles, carbon-based nanomaterials and quantum dots are some of the most widely studied ENPs.

Formation of reactive oxygen species (ROS) is found to be an important route of nanoparticle toxicity<sup>29</sup>. Generation of free radicals and ROS such as hydroxyl radicals leads to oxidative stress<sup>30-32</sup>.

Large surface area of NPs increases its inherent toxicity due to enhanced interaction of organisms with these surfaces<sup>33-35</sup>.

### Ecotoxicity of Engineered Nanoparticles

To assess the impact of nanomaterials both toxicity and ecotoxicity need to be taken into account. Whereas toxicity deals with the harmful effects of a compound after its uptake, ecotoxicity looks at absorption mechanisms, bioavailability and its dependence on environmental factors. Transformations brought about by factors like pH, temperature, and salinity can alter ecotoxicity.

There are concerns that pollution could alter the path of natural nano-scale processes. Interaction of nanoparticles with other species could lead to their being rendered less harmful or less easily absorbed by organisms. Engineered nanomaterials have a tendency for aggregation, and agglomeration which depends on shape, size and surface area of particles<sup>36,21</sup>.

Even though ecotoxicity studies of engineered nanoparticles are insufficient, data available from several studies indicate that several engineered nanoparticles are toxic to organisms even in low concentrations. To assess the threat posed by ENPs to organisms, data regarding reactivity, transport, absorption mechanisms and persistency in the environment is required.

### Ecotoxicity of Carbon based nanoparticles

One of the first engineered nanoparticles to be investigated for their ecotoxicity is C60 fullerenes. Oberdörster was one of the first to examine that C60 even in concentrations as low as 0.5 mg/l can cause oxidative damage in fish (large mouth bass; *Micropterus salmoides*)<sup>37</sup>. Although several publications attributed the toxic products to a THF degradation product. Adverse health effects of C-60 have since been reviewed and debated. Oxidative, genotoxic, and cytotoxic responses have been attributed to C-60<sup>38,39</sup>. These effects vary according to the form and chemical modifications of fullerenes<sup>40</sup>. Although in another contradicting report Gharbi et al show C-60 to be a powerful anti oxidant. The study involved use of C(60) in acute carbon tetrachloride intoxication in rats pointing to the fact that in the absence of a polar organic solvent C(60) could help protect the livers of rats from free-radical damage<sup>41</sup>.

Liu Y, et al highlight variations in CNT toxicity through alterations in factors such as size, shape, impurities, surface charge and agglomeration. They discuss how CNT toxicity proceeds through oxidative stress, DNA damage mutation and progression towards malignancy and how routes of exposure routes influence its behavior<sup>42</sup>. Several studies highlight the dependence of CNT toxicity on exposure conditions, type of carbon nanotubes, structural characteristics, surface properties, chemical composition dispersion state and concentration<sup>43-45</sup> although more studies would help to gain a more comprehensive interpretation of the toxicity of CNTs<sup>38</sup>.

### Ecotoxicity of Inorganic Nanoparticles

Inorganic ENPs mainly Ag<sup>0</sup>, titania, ceria, zinc oxide, and iron oxides present in various manufactured products are associated with biota. Bottero et al found a strong relationship between biological malfunctions and the physico-chemical characteristics of very small nanoparticles. Ag<sup>0</sup> and CeO<sub>2</sub> are found act even at very low concentrations. TiO<sub>2</sub> being photo-active induced toxicity through ROS production<sup>46</sup>.

Cytotoxic effects of TiO<sub>2</sub>, ZnO, and CeO<sub>2</sub>, were compared by Xia et al to explain the relationship between physicochemical properties and cellular uptake and translocation. ZnO acts via formation of reactive oxygen species (ROS), inflammation, and apoptosis, CeO<sub>2</sub> nanoparticles did not cause inflammation or cytotoxicity on absorption by cells but suppressed the formation of reactive oxygen species whereas TiO<sub>2</sub> even though followed the same uptake mechanism as CeO<sub>2</sub> but did not exhibit either protective or harmful impact. This leads to the conclusion that metal oxide nanoparticles could induce varied biological responses ranging from cytoprotective to cytotoxic<sup>47</sup>.

ZnO NPs which have widespread applications in electronics, clothing, paints, cosmetic products, catalysts as well as in biosensors and medical devices are shown to have cytotoxic and genotoxic potential<sup>48,49</sup>. Various studies showed that ZnO NPs were cytotoxic for human lung epithelial cells. They are shown to upset glucose metabolism in such cells cause mitochondrial dysfunction, and induce apoptosis<sup>50,51</sup>. Han Z et al carried out in-vitro studies on mice injected with single doses of ZnO nanoparticles which were taken up by Leydig and Sertoli cells, causing cytotoxicity due to DNA damage brought about by increase in reactive oxygen species<sup>52</sup>. Johnson et al showed that the exposure to ZnO NPs led to autophagocytosis of immune cells, due to rise in the levels of the autophagosome protein LC3A reactive oxygen species (ROS) production<sup>53</sup>. Several others reported DNA damage in human epidermal cells, and apoptosis in pulmonary epithelial cell lines through ROS and oxidative stress even when exposure to ZnO NPs was very low<sup>49,54,55</sup>.

Experimental studies with nanotitania (having applications in self cleaning glass, low cost solar cells and cosmetics) have shown genotoxicity in aquatic organisms<sup>56,57</sup> and documented

lung injury and inflammation, toxicity via oxidative stress in human cells<sup>58-60</sup>.

A study investigating the impact of short-term exposures on workers in TiO<sub>2</sub> manufacturing units, documented increased oxidative stress, DNA and protein damage, as well as lipid peroxidation in their exhaled breath condensate (EBC) in comparison to unexposed control groups<sup>61</sup>.

Silver nanoparticles are another example of nanoparticles commercialized into various products<sup>62</sup> and a large amount of AgNPs enter into aquatic ecosystems<sup>63-65</sup>, are bioaccumulated<sup>66-68</sup> and liberate Ag<sup>+</sup> and induce formation of ROS<sup>69,70</sup>. Through observation of gene expression pattern Kwok et al suggested that disruption of sodium regulation was responsible for toxicity of AgNPs. They found that dissolution of silver nanoparticles depends on particle size, but coating materials influence agglomeration and toxicity<sup>71</sup> whereas Guo et al reported that particle size plays an important role in the genotoxicity of AgNPs. Comparative study of genotoxicities of silver acetate and silver nitrate has been done, to see the effect of nanosilver vis a vis ionic silver<sup>72</sup>.

QDs with applications in semiconductor devices, lasers, TV and computer displays and for drug delivery may also pose a threat to human health. The uptake, transport and toxicity of QD are determined by environmental factors as well as properties such as size, charge, outer coating of quantum dots, and their oxidative and photolytic stability<sup>73,74</sup>.

## Environmental Hazards and Risks

A lot of the data available from toxicity studies is based on experiments performed on organisms in laboratory settings but the same results are not always obtained when the experiment is carried out in actual environments<sup>75</sup>. Despite a huge number of publications on nanotoxicology, it is still unclear how significant the potential adverse impacts are, and whether the benefits from nanotechnological developments outweigh the risks involved. The current literature on ecotoxicity of nanomaterials lacks much of the information needed to evaluate potential hazards and quantify exposure with hazard<sup>27</sup>. Assessment of hazard requires evaluation of the form of nanomaterial in the environment, transport, transformation bioavailability and bioaccumulation. Gebel, et al suggest categorisation of nanoparticles based on exposure pathway and mode of action, to evaluate hazards<sup>76</sup>. A large number of ongoing studies utilize different models to evaluate toxicology of diverse NPs but there is often a gap between modeled values with actual measured values.

There is a need to study sets of ENPs to allow for quantification of characteristics such as physicochemical properties and biological response outcomes which would help to build dose-response relation data<sup>77,78</sup>.

## Regulations

A study<sup>79</sup> on nano titania reported the surface water concentrations of TiO<sub>2</sub> nano particles between 0.012–0.057 µg/L<sup>-1</sup> in Europe, 0.002–0.010 µg/L<sup>-1</sup> in the U.S., and 0.016–0.085 µg/L<sup>-1</sup> in Switzerland, but such data is hardly available for Indian environments<sup>80</sup> even though, much higher concentrations of nanoparticles due to rapid industrialization and lack of proper regulations may be expected<sup>81</sup>.

Some studies have focussed recently on the presence of nanoparticles in commercial products in India and their distribution and prevalence in the environment<sup>62</sup> and risk management.

A few organizations, such as Indian Institute of Toxicology Research (IITR) and National Institute of Pharmaceutical Education and Research (NIPER), CSIR etc. are involved with nanotechnology related risk research in India.

However, there are no nanotechnology specific regulations in India currently, though there is an entire range of regulations that can extend to nanotechnology applications<sup>82,83</sup>. India is making rapid and noteworthy progress in nanotechnology but research on toxicity is not in pace with the rate of introduction of new materials<sup>84-86</sup>.

## Conclusion

Nanotechnology is a field with immense potential and applications in all walks of life. In a context of uncertainty about the risks there is a need for research to monitor the environmental occurrence and hazards related to nanoparticles so that benefits from nanotechnology and nanoscience outweigh the risks associated with it.

## References

1. Guzman K.A.D., Taylor M.R. and Banfield J.F. (2006). Environmental Risks of Nanotechnology: National Nanotechnology Initiative Funding, 2000–2004. *Environ. Sci. Technol.*, 40(5), 1401-1407. doi: 10.1021/es0515708
2. UNESCO Science Report (2015). towards 2030.
3. Roco M.C., Mirkin C.A. and Hersam M.C. (2011). Nanotechnology research directions for societal needs in 2020: summary of international study. *J. Nanopart. Res.*, 13, 897-919. doi:10.1007/s11051-011-0275-5
4. Nowack B. and Bucheli T.D. (2007). Occurrence, behavior and effects of nanoparticles in the environment. *Environ. Pollut.*, 150, 5-22. doi:10.1016/j.envpol.2007.06.006
5. Wiesner Mark R., Lowry Greg V., Alvarez Pedro, Dionysiou Dianysios and Biswas Pratim (2006). Assessing the risks of manufactured nanoparticles. *Environ. Sci. Technol.*, 40(14), 4336-4345. doi: 10.1021/es062726m

6. Maynard A.D. (2007). Nanotechnology: Overviews and Issues. *Nanotechnology-Toxicological Issues and Environmental Safety*, Springer Publications, Netherlands, 1-14,
7. Roco M.C., Harthorn B., Guston D. and Shapira P. (2011). Innovative and responsible governance of nanotechnology for societal development. *Nanotechnology Research Directions for Societal Needs in 2020*, Springer Netherlands, 561-617.
8. Grieger K.D., Hansen S.F. and Baun A. (2009). The known unknowns of nanomaterials: describing and characterizing uncertainty within environmental, health and safety risks. *Nanotoxicology*, 3(3), 222-233. doi: 10.1080/17435390902944069
9. Hristozov D. and Malsch I. (2009). Hazards and risks of engineered nanoparticles for the environment and human health. *Sustainability*, 1(4), 1161-1194. doi: 10.3390/su1041161
10. Savolainen K., Alenius H., Norppa H., Pylkkanen L., Tuomi T. and Kasper G. (2010). Risk assessment of engineered nanomaterials and nanotechnologies-a review. *Toxicology*, 269(2-3), 92-104. doi:10.1016/j.tox.2010.01.013
11. Pettitt M.E. and Lead J.R. (2013). Minimum physicochemical characterization requirements for nanomaterial regulation. *Environ. Int.*, 52, 41-50. doi: 10.1016/j.envint.2012.11.009.
12. Oberdörster G., Oberdörster E. and Oberdörster J. (2005). Nanotoxicology: an emerging discipline evolving from studies of ultrafine particles. *Environmental Health Perspectives*, 113, 823-839. doi:10.1289/ehp.7339
13. Kreyling W.G., Semmler-Behnke M. and Möller W. (2006). Health Implications of Nanoparticles. *J Nanopart Res.*, 8(3), 543-562. doi:10.1007/s11051-005-9068-z
14. Net Andre, Tian Xia, Mädler Lutz and Ning Li (2006). Toxic Potential of Materials at the Nanolevel. *Science*, 311(5761), 622-627 doi: 10.1126/science.1114397
15. Bergamaschi E. (2009). Occupational exposure to nanomaterials: present knowledge and future development. *Nanotoxicology*, 3(3), 194-201. doi:10.1080/17435390903037038
16. Maurer-Jones Melissa A., Gunsolus Ian L., Murphy Catherine J. and Haynes Christy L. (2013). Toxicity of engineered nanoparticles in the environment. *Analytical Chemistry*, 85(6), 3036-3049. doi: 10.1021/ac303636s
17. Hotze E.M., Phenrat T., Lowry G.V. (2010). Nanoparticle aggregation: Challenges to understanding transport and reactivity in the environment. *J. Environ. Qual.*, 39(6), 1909-1924.
18. Nowack B. (2009). The Behaviour and Effects of Nanoparticles in the Environment. *Environ Pollut*, 157(4), 1063-1064. doi: 10.1016/j.envpol.2008.12.019
19. Hassellöv M., Readman J.W., Ranville J.F. and Tiede K. (2008). Nanoparticle analysis and characterization methodologies in environmental risk assessment of engineered nanoparticles. *Ecotoxicology*, 17, 344-361. doi: 10.1007/s10646-008-0225-x..
20. Tiede K., Hassellöv M., Breitbarth E., Chaudhry Q. and Boxall A.B.A. (2009). Considerations for environmental fate and ecotoxicity testing to support environmental risk assessments for engineered nanoparticles. *J.Chromatogr. A*, 1216(3), 503-509. doi: 10.1016/j.chroma.2008.09.008.
21. Lowry G.V., Gregory K.B., Apte S.C. and Lead J.R. (2012). Transformations of nanomaterials in the environment. *Environ. Sci. Technol.*, 46(13), 6893-6899. doi: 10.1021/es300839e.
22. Handy R.D., von der Kammer F., Lead J.R., Hassellöv Martin, Owen Richard and Crane Mark (2008). The ecotoxicology and chemistry of manufactured nanoparticles. *Ecotoxicology*, 17, 287-314. doi:10.1007/s10646-008-0199-8.
23. Kammer Frank von der, Legros Samuel, Hofmann Thilo, Larsen Erik H. and Loeschner Katrin (2011). Separation and characterization of nanoparticles in complex food and environmental samples by field-flow fractionation. *Trends in Analytical Chemistry*, 30(3), 425-436.
24. Roth Gary A., Sahil Tahiliani, Neu-Baker Nicole M. and Brenner Sara A. (2015). Hyperspectral microscopy as an analytical tool for nanomaterials. *Wiley Interdisciplinary Reviews: Nanomedicine and Nanobiotechnology*, 7(4), 565-579
25. White Brittany, Strawbridge Andrew, Grabinski Christin M. and Hussain Saber M. (2013). Hyperspectral imaging (HSI) to evaluate the interaction of optically active nanoparticles in biological media and cells. *Bios*, 84(4), 210-217.
26. Auffan M., Rose J., Bottero J.Y., Lowry G.V., Jolivet J.P. and Wiesner M. (2009). Towards a definition of inorganic nanoparticles from an environmental, health and safety perspective. *Nat. Nanotechnol.*, 4, 634-641.
27. Wiesner M.R., Lowry G.V., Jones K.L., Hochella M.F., Di Giulio R.T., Casman E. and Bernhardt E.S. (2009). Decreasing uncertainties in assessing environmental exposure, risk, and ecological implications of nanomaterials. *Environ. Sci. Technol.*, 43(17), 6458-6462.
28. Zhao F., Zhao Y., Liu Y., Chang Xueling, Chen Chunying and Zhao Yuliang (2011). Cellular uptake, intracellular trafficking and cytotoxicity of nanomaterials. *Small*, 7(10), 1322-1337. doi: 10.1002/smll.201100001
29. Kovochich M., Xia T., Xu J., Yeh J.I. and Nel A.E. (2007). Principles and procedures to assess nanomaterial toxicity. *Environmental Nanotechnology: Applications and Impacts*

- of Nanomaterials, Mc Graw Hill, New York, 205-229 ISBN: 9780071477505.
30. Li J., Chang X., Chen X., Gu Zhanjun, Zhao Feng, Chai Zhifang and Zhao Yuliang (2014). Toxicity of inorganic nanomaterials in biomedical imaging. *Biotechnol Adv*, 32(4), 727-743. doi: 10.1016/j.biotechadv.2013.12.009
31. Luna-Velasco A., Field J.A., Cobo-Curiel A. and Sierra-Alvarez R. (2011). Inorganic nanoparticles enhance the production of reactive oxygen species (ros) during the autoxidation of L-3,4-dihydroxyphenylalanine (L-dopa). *Chemosphere*, 85(1), 19-25. doi: 10.1016/j.chemosphere.2011.06.053.
32. Shvedova A.A., Pietroiusti A., Fadeel B. and Kagan V.E. (2012). Mechanisms of carbon nanotube-induced toxicity: Focus on oxidative stress. *Toxicol Appl Pharmacol*, 261(2), 121-133. doi: 10.1016/j.taap.2012.03.023.
33. Choi O. and Hu Z. (2008). Size dependent and reactive oxygen species related nanosilver toxicity to nitrifying bacteria. *Environ Sci Technol.*, 42(12), 4583-4588. doi: 10.1021/es703238h.
34. Hussain S.M., Hess K.L., Gearhart J.M., Geiss K.T. and Schlager J.J. (2005). In vitro toxicity of nanoparticles in brl 3a rat liver cells. *Toxicol In Vitro*, 19(7), 975-983. doi: 10.1016/j.tiv.2005.06.034.
35. Zoroddu M., Medici S., Ledda A., Nurchi V., Lachowicz J. and Peana M. (2014). Toxicity of nanoparticles. *Current Medicinal Chem.*, 21, 3837-3853. doi: 10.2174/0929867321666140601162314.
36. Handy R.D., Owen R. and Valsami-Jones E. (2008). The ecotoxicology of nanoparticles and nanomaterials: current status, knowledge gaps, challenges, and future needs. *Ecotoxicology*, 17(5), 315-325. doi:10.1007/s10646-008-0206-0
37. Oberdörster Eva, Zhu Shiqian, Blickley T. Michelle, McClellan-Green Patricia and Haasch Mary L. (2006). Ecotoxicology of carbon-based engineered nanoparticles: effects of fullerene (C 60) on aquatic organisms. *Carbon*, 44(6), 1112-1120.
38. Dhawan A., Taurozzi J.S., Pandey A.K., Shan W., Miller S.M., Hashsham S.A. and Tarabara V.V. (2006). Stable colloidal dispersions of C60 fullerenes in water: evidence for genotoxicity. *Environ. Sci. Technol.*, 40(23), 7394-7401. doi: 10.1021/es0609708
39. Markovic Z., Todorovic-Markovic B., Kleut D., Nikolic N., Vranjes-Djuric S., Misirkic M., Vucicevic L., Janjetovic K., Isakovic A., Harhaji L., Babic-Stojic B., Dramicanin M. and Trajkovic V. (2007). The mechanism of cell-damaging reactive oxygen generation by colloidal fullerenes. *Biomaterials*, 28(36), 5437-5448. doi: 10.1016/j.biomaterials.2007.09.002
40. Zhang Leshuai W., Yang Jianzhong, Barron Andrew R. and Monteiro-Riviere Nancy A. (2009). Endocytic mechanisms and toxicity of a functionalized fullerene in human cells. *Toxicology Letters*, 191(2-3), 149-157. doi: 10.1016/j.toxlet.2009.08.017
41. Gharbi N., Pressac M., Hadchouel M., Szwarc H., Wilson S.R. and Moussa F. (2005). [60] Fullerene is a powerful antioxidant in vivo with no acute or subacute toxicity. *Nano Lett.*, 5(12), 2578-2585. doi:10.1021/nl051866b
42. Liu Y., Zhao Y., Sun B. and Chen C. (2013). Understanding the toxicity of carbon nanotubes. *Acc Chem Res*, 46(3), 702-713. doi: 10.1021/ar300028m
43. Jackson P., Jacobsen N.R., Baun A., Birkedal Renie, Kühnel Dana, Alstrup Jensen Keld, Vogel Ulla and Wallin Håkan (2013). Bioaccumulation and ecotoxicity of carbon nanotubes. *Chemistry Central Journal*, 7, 154. doi:10.1186/1752-153X-7-154
44. Johnston H.J., Hutchison G.R., Christensen F.M., Peters S., Hankin S., Aschberger K. and Stone V. (2010). A critical review of the biological mechanisms underlying the in vivo and in vitro toxicity of carbon nanotubes: The contribution of physico-chemical characteristics. *Nanotoxicology*, 4(2), 207-246. doi: 10.3109/17435390903569639.
45. Jing Wang Xu, Yuanzhi Yang, Zhi Huang, Renhuan Chen, Jing Wang, Raorao Lin and Yunfeng (2013). Toxicity of carbon nanotubes. *Current Drug Metabolism*, 14(8), 891-899(9).
46. Bottero Jean-Yves, Auffan Mélanie, Rose Jérôme, Mouneyrac Catherine, Botta Céline, Labille Jérôme, Masion Armand, Thill Antoine and Chaneac Corinne (2011). Manufactured metal and metal-oxide nanoparticles: Properties and perturbing mechanisms of their biological activity in ecosystems. *Comptes Rendus Geoscience*, 343(2), 168-176.
47. Xia Tian, Kovochich Michael, Liong Monty, Mädler Lutz, Gilbert Benjamin, Shi Haibin, Yeh Joanne I., Zink Jeffrey I. and Nel Andre E. (2008). Comparison of the Mechanism of Toxicity of Zinc Oxide and Cerium Oxide Nanoparticles Based on Dissolution and Oxidative Stress Properties. *ACS Nano*, 2(10), 2121-2134. doi: 10.1021/nn800511k
48. Sharma V., Singh S.K., Anderson D., Tobin D.J., Dhawan A. (2011). Zinc oxide nanoparticle induced genotoxicity in primary human epidermal keratinocytes. *J Nanosci Nanotechnol.*, 11(5), 3782-3788.
49. Sharma V., Shukla R.K., Saxena N., Parmar D., Das M. and Dhawan A. (2009). DNA damaging potential of zinc oxide nanoparticles in human epidermal cells. *Toxicol Lett.*, 185(3), 211-218.
50. Lai X., Wei Y., Zhao H., Chen Suning, Bu Xin, Lu Fan, Qu Dingding, Yao Libo, Zheng Jianyong and Zhang Jian (2015). The effect of Fe2O3 and ZnO nanoparticles on cytotoxicity and glucose metabolism in lung epithelial cells.

- J Appl Toxicol.*, 35(6), 651-664. doi: 10.1002/jat.3128.
51. Lin W., Xu Y., Huang C.C., Shannon K.B., Chen D.R. and Huang Y.W. (2009). Toxicity of nano- and micro-sized ZnO particles in human lung epithelial cells. *J Nanopart Res.*, 11, 25-39. doi: 10.1007/s11051-008-9419-7
52. Han Z., Yan Q., Ge W., Liu Zhi-Guo, Gurunathan Sangiliyandi, De Felici Massimo, Shen Wei and Zhang Xi-Feng (2016). Cytotoxic effects of ZnO nanoparticles on mouse testicular cells. *International Journal of Nanomedicine*, 11, 5187-5203. doi:10.2147/IJN.S111447.
53. Johnson B.M., Fraietta J.A., Gracias D.T., Jennifer L Hope, Stairiker Christopher J., Patel Prachi R., Mueller Yvonne M., McHugh Michael D., Jablonowski Lauren J., Wheatley Margaret A. and Katsikis Peter D. (2015). Acute exposure to ZnO nanoparticles induces autophagic immune cell death. *Nanotoxicology*, 9(6), 737-748. doi: 10.3109/17435390.2014.974709.
54. Ahamed M., Akhtar M.J., Raja M., Ahmad Iqbal, Javed Siddiqui Mohammad Kaleem, AlSalhi Mohamad S. and Alrokayan Salman A. (2011). ZnO nanorod-induced apoptosis in human alveolar adenocarcinoma cells via p53, surviving and bax/bcl-2 pathways: role of oxidative stress. *Nanomedicine*, 7(6), 904-913. doi: 10.1016/j.nano.2011.04.011.
55. Ng K.W., Khoo S.P., Heng B.C., Setyawati Magdiel I., Tan Eng Chok, Zhao Xinxin, Xiong Sijing, Fang Wanru, Leong David T. and Loo Joachim S.C. (2011). The role of the tumor suppressor p53 pathway in the cellular DNA damage response to zinc oxide nanoparticles. *Biomaterials*, 32(32), 8218-8225. doi: 10.1016/j.biomaterials.2011.07.036
56. Hund-Rinke K. and Simon M. (2006). Ecotoxic effect of photocatalytic active nanoparticles (TiO<sub>2</sub>) on algae and daphnids. *Environ Sci Pollut Res.*, 13(4), 225-232.
57. Federici G., Shaw B.J. and Handy R.D. (2007). Toxicity of titanium dioxide nanoparticles to rainbow trout, (*Oncorhynchus mykiss*): Gill injury, oxidative stress, and other physiological effects. *Aquat Toxicol*, 84(4), 415-430. doi: 10.1016/j.aquatox.2007.07.009
58. Sayes Christie M., Wahi Rajeev, Kurian Preetha A., Liu Yunping, West Jennifer L., Ausman Kevin D., Warheit David B. and Colvin Vicki L. (2006). Correlating Nanoscale Titania Structure with Toxicity: A Cytotoxicity and Inflammatory Response Study with Human Dermal Fibroblasts and Human Lung Epithelial Cells. *Toxicol. Sci.*, 92(1), 174-185. doi: 10.1093/toxsci/kfj197
59. Pierzchala K., Lekka M., Magrez A., Kulik A.J., Forró L. and Sienkiewicz A. (2012). Photocatalytic and phototoxic properties of TiO<sub>2</sub>-based nanofilaments: ESR and AFM assays. *Nanotoxicology*, 6(8), 813-24. doi: 10.3109/17435390.2011.625129.
60. Montiel-Dávalos A.I., Ventura-Gallegos J.L., Alfaro-Moreno E., Soria-Castro E., García-Latorre E., Cabañas-Moreno J.G., Ramos-Godínez M.P. and López-Marure R. (2012). TiO<sub>2</sub> nanoparticles induce dysfunction and activation of human endothelial cells. *Chem Res Toxicol.*, 25(4), 920-30. doi: 10.1021/tx200551u
61. Pelclova Daniela, Zdimal Vladimir, Kacer Petr, Fenclova Zdenka, Vlckova Stepanka, Syslova Kamila, Navratil Tomas, Schwarz Jaroslav, Zikova Nadezda, Barosova Hana, Turci Francesco, Komarc Martin, Pelcl Tomas, Belacek Jaroslav, Kukutschova Jana and Zakharov Sergey (2016). Oxidative stress markers are elevated in exhaled breath condensate of workers exposed to nanoparticles during iron oxide pigment production. *J. Breath Res.*, 10(1), 016004. doi:10.1088/1752-7155/10/1/016004
62. Sarma Deshpande S. (2011). Life Cycle of a Nanosilver Based Candle Filter: Examining Issues of Toxicity, Risks, Challenges and Policy Implications. *Journal of biomedical nanotechnology*, 7(1), 83-84.
63. Fabrega J., Luoma Samuel N., Tyler Charles R., Galloway Tamara S. and Lead Jamie R. (2011). Silver nanoparticles: Behaviour and effects in the aquatic environment. *Environ. Int.*, 37, 517-531. doi: 10.1016/j.envint.2010.10.012.
64. Choi, O., Deng Kathy Kanjun, Kim Nam-Jung, Ross Louis, Surampalli Rao Y. and Hu Zhiqiang (2008). The inhibitory effects of silver nanoparticles, silver ions, and silver chloride colloids on microbial growth. *Water Research*, 42(12), 3066-3074. doi: 10.1016/j.watres.2008.02.021.
65. Massarsky A., Trudeau V.L. and Moon T.W. (2014). Predicting the environmental impact of nanosilver. *Environ. Toxicol. Pharmacol.*, 38, 861-873. doi: 10.1016/j.etap.2014.10.006.
66. Hsiao I.L., Hsieh Yi-Kong, Wang Chu-Fang, Chen I-Chieh and Huang Yuh-Jeen (2015). Trojan-horse mechanism in the cellular uptake of silver nanoparticles verified by direct intra- and extracellular silver speciation analysis. *Environmental Science & Technology*, 49(6), 3813-3821. doi: 10.1021/es504705p.
67. Wang Zhe, Liu Sijin, Ma Juan, Qu Guangbo, Wang Xiaoyan, Yu Sujuan, He Jiuyang, Liu Jingfu, Xia Tian and Gui- Jiang Bin (2013). Silver nanoparticles induced RNA polymerase silver binding and RNA transcription inhibition in erythroid progenitor cells. *ACS Nano*, 7(5), 4171-4186. doi: 10.1021/nn400594s
68. Adjei I.M., Sharma B. and Labhasetwar V. (2014). Nanoparticles: Cellular uptake and cytotoxicity. *Advances in Experimental Medicine & Biology*, 811, 73-91. doi:10.1007/978-94-017-8739-0\_5
69. Yu S.J., Yin Y.G. and Liu J.F. (2013). Silver nanoparticles in the environment. *Environmental Science Processes & Impacts*, 15, 78-92. doi 10.1039/C2EM30595J
70. Marin Stefania, Vlasceanu George Mihail, Tiplea Roxana

- Elena, Bucur Ioana Raluca, Lemnaru Madalina, Marin Maria Minodora and Grumezescu Alexandru Mihai (2015). Applications and toxicity of silver nanoparticles: A recent review. *Current Topics in Medicinal Chemistry*, 15(16), 1596-1604. doi: 10.2174/1568026615666150414142209
71. Kwok Kevin W.H., Dong Wu, Marinakos Stella M., Liu Jie, Chilkoti Ashutosh, Wiesner Mark R., Chernick Melissa and Hinton David E. (2016). Silver nanoparticle toxicity is related to coating materials and disruption of sodium concentration regulation. *Nanotoxicology*, 10(9), 1306-1317. doi: 10.1080/17435390.2016.1206150
72. Guo Xiaoqing, Li Yan, Yan Jian, Ingle Taylor, Jones Margie Yvonne, Mei Nan, Boudreau Mary D., Cunningham Candice K., Abbas Mazhar, Paredes Angel M., Zhou Tong, Moore Martha M., Howard Paul C. and Chen Tao (2016). Size- and coating-dependent cytotoxicity and genotoxicity of silver nanoparticles evaluated using in vitro standard assays. *Nanotoxicology*, 10(9), 1373-1384. doi: 10.1080/17435390.2016.1214764
73. Chen Nan, He Yao, Su Yuanyuan, Li Xiaoming, Huang Qing, Wang Haifeng, Zhang Xiangzhi, Tai Renzhong and Fan Chunhai (2012). The cytotoxicity of cadmium-based quantum dots. *Biomaterials*, 33(5), 1238-1244. doi: 10.1016/j.biomaterials.2011.10.070.
74. Bottrill Melanie and Green Mark (2011). Some aspects of quantum dot toxicity. *Chem. Commun.*, 47(25), 7039-7050. doi: 10.1039/C1CC10692A
75. Kang S., Mauter M.S. and Elimelech M. (2009). Microbial cytotoxicity of carbon-based nanomaterials: Implications for river water and wastewater effluent. *Environ. Sci. Technol.*, 43(7), 2648-2653. doi: 10.1021/es8031506
76. Gebel Thomas, Foth Heidi, Damm Georg, Freyberger Alexius, Kramer Peter-Jürgen, Lilienblum Werner, Röhl Claudia, Schupp Thomas, Weiss Carsten, Wollin Klaus-Michael and Hengstler Jan Georg (2014). Manufactured nanomaterials: categorization and approaches to hazard assessment. *Archives of toxicology*, 88(12), 2191-2211. doi:10.1007/s00204-014-1383-7
77. Bouwmeester Hans, Lynch Iseult, Marvin Hans J.P., Dawson Kenneth A., Berges Markus, Braguer Diane, Byrne Hugh J., Casey Alan, Chambers Gordon, Clift Martin J.D., Elia Giuliano, Fernandes Teresa F., Fjellsbø Lise B., Hatto Peter, Juillerat Lucienne, Klein Christoph, Kreyling Wolfgang G., Nickel Carmen, Riediker Michael and Stone Vicki (2011). Minimal analytical characterization of engineered nanomaterials needed for hazard assessment in biological matrices. *Nanotoxicology*, 5(1), 1-11, doi: 10.3109/17435391003775266
78. Meng Huan, Xia Tian, George Saji and Nel Andre E., (2009). A Predictive Toxicological Paradigm for the Safety Assessment of Nanomaterials. *ACS Nano*, 3(7), 1620-1627. doi: 10.1021/nn9005973
79. Gottschalk F., Sonderer T., Scholz R.W. and Nowack B. (2009). Modelled environmental concentrations of engineered nanomaterials (TiO<sub>2</sub>, ZnO, Ag, CNT, Fullerenes) for different regions. *Environ. Sci. Technol.*, 43(24), 9216-9222. doi: 10.1021/es9015553.
80. Kumar Prashant, Kumar Arun and Lead Jamie R. (2012). Nanoparticles in the Indian environment: known, unknowns and awareness. *Environmental Science & Technology*, 46(13), 7071-7072. doi: 10.1021/es302308h
81. Kumar P., Gurjar B.R., Nagpure A. and Harrison R.M. (2011). Preliminary estimates of nanoparticle number emissions from road vehicles in megacity Delhi and associated health impacts. *Environ. Sci. Technol.*, 45(13), 5514-5521.
82. Jayanthi A.P., Beumer K. and Bhattacharya S. (2012). Nanotechnology: Risk governance in India. *Econ. Political Daily*, 47(4), 34-40.
83. Sarma Deshpande S. (2011). How Resilient is India to Nanotechnology Risks? Examining Current Developments, Capacities and an Approach for Effective Risk Governance and Regulation. *European Journal of Law and Technology*, 2(3),
84. TERI Report (2009). Report on Regulatory Challenges Posed by Nanotechnology Developments in India. The Energy and Resources Institute, New Delhi.
85. TERI Report (2009). Nanotechnology Developments in India: a Status Report. The Energy and Resources Institute, New Delhi.
86. Beumer K. and Bhattacharya S. (2013). Emerging technologies in India: Developments, debates and silences about nanotechnology. *Science and Public Policy*, 40(5), 628-643. doi:10.1093/scipol/sct016.