

Bioactive Compounds and Medicinal Value of two Spices-Zingiber officinale Rosc. and Curcuma longa Linn

Andzouana Marcel* and Makomo Hubert

Department of Chemistry, faculty of Sciences and technics, Marien NGOUABI, University, Brazzaville, Congo andzouanamarcel@gmail.com

Available online at: www.isca.in, www.isca.me

Received 23rd August 2016, revised 29th September 2016, accepted 9th October 2016

Abstract

Zingiber officinale Rosc. and Curcuma longa Linn are two medicinal plants widely used as spices and are known in folk medicine for their various uses. Qualitative and quantitative analyses of the two spices were undertaken in various solvents using different methods to determine the medicinal value of these spices. The spices were assessed for the following phytochemicals: glycosides, flavonoids, saponins, alkaloids, terpenoids, tannins, tannins, phenols, carotenoids, anthocyanins and steroids. The test compounds were present in most of the screened plant extracts. Morever phytochemicals were highly present in ethanol extracts of both plants than in ethyl acetat extracts. Anthocyanins, phenols and tannins were the most common phytochemicals present in the extracts. Quantitative analysis showed that the plants contained various concentrations of phytochemicals. The results also indicated that C.longa contained appreciable amounts (0.02±0.01-15.07±0.04%) of phytoconstituents other than ginger (0.02±0.03-22.41%). The spices contained different bioactive compounds with biological properties leading us to conclude that consumption of the two varieties of capsicum spices may be important in health care and could be recommended in nutritional supplementation either as source of drugs in folk medicine.

Keywords: Spices, Nutrition, Analysis, Phytochemical, Medicinal.

Introduction

Tropical plants in Africa are known for their multiple uses in nutrition as spices, or components of bloods and in folk medicines as drugs to prevent or to cure various diseases. Among them *Zingiber officinale* (Roscoe), Zingiberaceae family, is a perennial plant (herbaceous in form of rhizome) found in warm climatic regions like Nigeria, Bangladesh, Taiwan, India, Jamaica and the United States of America. The rhizome contains numerous active compounds with medicinal properties. These properties contributed to its various medical applications.

Ginger is commonly used as a spice^{1,2} and has also been used to treat diarrhea, nausea, asthma and respiratory disorders³. In addition ginger extracts are reported to be natural larvicidal agents⁴ and showed anti-inflammatory, antioxidant and anticancer effects. Shogaol can significantly attenuate a variety of neuro inflammatory responses in cortical astrocytes^{5,6}.

The chemical constituents of Z.officinale are numerous depending on such factors like the origin of the sample and the state of the plant part used (fresh or dry)⁷.

Curcuma longa Linn (Zingiberaceae) is a rhizomatous herbaceous perennial herb, cultivated extensively in south and southeast tropical Asia⁸, Belgium, France, Ceylon, south India,

Indonesia and Bengal⁹. As spice, turmeric is important plant rhizome mainly used in nutrition for its color and aroma².

C.longa has been used as a condiment, a healing remedy, textile dye and for regulation of serum cholesterol and lipid peroxide levels¹⁰. Literature reveals that it was used against inflammations, cholagogue, hepatic, asthma, stomach pains, its antitumor, antiprotozoal, carminative properties and its ability to purify blood and liver regulation¹¹. Numerous studies have shown good antioxidant properties of ginger^{1,12} and turmeric¹³.

Various literature reports were found on the chemistry of the ginger and turmeric rhizomes^{7,14-16} but no study was found on the species from Congo. The present investigation was aimed to determine the phytochemical profile of *G.officinale* and *C. Longa* in order to determine the medicinal importance of these spices.

Materials and Methods

Samples of *G.officinale* and *C. longa* were bought from the local 'Total' market in Bacongo, South –Brazzaville in July 2014. Identification and authentification of the plant samples were done by Nkouka Saminou from the national herbarium of the vegetal research centre of Brazzaville (ex-OROSTOM-Congo) where voucher specimens are deposited. The plant samples were dried in an oven at 100-105°C for 24 hours and

Res. J. Chem. Sci.

milled into a powder. The powder was stored under dry conditions before analysis.

Chemical analysis: Physico-chemical analysis: The extractive values of the samples were determined by using various methods of extraction in water. Physico-chemical analysis was performed on water extracts obtained by infusion, this being the highest yield method. Titratable acidity was determined by titration with a dilute sodium hydroxide solution (0.1M/L). The PH and the temperature values of the solutions were measured automatically with a ph-meter. The results are presented in Table-1.

Qualitative and quantitative phytochemical analysis: **Extraction procedure:** The powdered samples (15g) from each plant were soaked in 100ml of 98% ethanol solution for 72 hours. After filtration of the obtained solutions, the filtrates were evaporated at room temperature for five days and submitted to phytochemical screening. For C.longa the defatted material was used for extraction then 40g of the powder were delipidated twice with 100ml of diethyl ether for 24h.

Preliminary phytochemical screening: Phytochemical analysis was carried out to confirm the presence of the various phytochemicals in the samples. The phytochemical constituents of the spices were determined according to the described methods¹⁷⁻²². The phytochemicals tested were alkaloids, flavonoids, glycosides, Tannins, triterpeoids, steroids, saponins, phenols, carotenoids and anthocyanins, and the corresponding results are shown in Table-2.

Quantitative phytochemical determination: Quantitative evaluation of bioactive constituents was carried out using the classic methods²³⁻²⁷. These active compounds include phenols, alkaloids. flavonoids. cardiac glycosides, anthocyanins and carotenoids.

Statistical analysis: Data were determined as means±SD of triplicate determination.

Table-1 Physico-chemical characteristics of *Z.offinales* and *C.longa* extracts

Plant extract	Physico-chemical characteristics			Method of extraction and Yield(%)		
(in water)	Acidity(g/l)	PH	T°C	Ma	If	De
Z.officinale	0.45	2.30	28.20	2.00	2.20	1.80
C.longa	2.64	3.75	27.80	1.80	2.60	2.10

Ma=maceration; If=infusion; De=decoction

Table-2 Qualitative phytochemical analysis of *Z.officinale and C.longa* extracts

	Z.	officinale	C.longa	
Phytochemical	ethanol	Ethyl acetat	ethanol	Ethyl acetat
Alcaloides	+++	+	+	+
Flavonoides	+++	+	+	+
Glycosides	++	+	+	+
Tannins	++	+++	+	-
Terpenoids	+++	+	+++	+
Steroids	+ ++	+	+	+
Saponins	-	+++	-	-
Phenols	+	+	+++	+++
Anthocyanines	+	+	+++	+++
Carotenoids	-	-	+	+++

+++= high concentration ++= moderately present += trace concentration Table-3

Ouantitative phytochemical analysis of Z.officinale and C.longa

Phytochemical (%)	Plant specie		
Filytochemical (%)	Z.officinale	C.longa	
Alcaloides	11.28±2.05	7.33±0.18	
Flavonoides	15.07±0.04	8.5±0.12	
Glycosides	00.00	00.00	
Saponins	2.13±0.07	22.41± 1.33	
Phenols	0.20±1.96	11.33±0.18	
Anthocyanins	0.40±3.92	2.33±0.16	
Carotenoids	0.02±0.003	0.02±0.01	

Results and Discussion

Physico-chemical analysis of water extracts: The results of water extraction shown in Table-1 indicated that infusion was the best method of extraction for both samples with yields of 2.20% for *Z.officinale* and 2.60% for *C.longa*. These values fell within the range of 2.24-2.726% recorded in acetone⁸. They were higher when compared with those reported for *Z.officinale*(1.50%) ²⁸ in petroleum -ether but lower than the 2.90% in ethanol. A very high extract yield was recorded for turmeric (66.20%) and ginger (57.80%)¹⁴. The high yield observed for turmeric was similarly reported in the range of 2.88-3.14% ⁸. The extract yield may depend on many factors such as the conditions of extraction, the nature of the solvent and the plant part used. *Z.officinal* extract was more acidic than that of turmeric at the same temperature.

Phytochemical screening of plant rhizomes: The results of phytochemical screening of the samples are summarized in Table-1. Qualitative analysis revealed the plants contained phenols, flavonoids, steroids, cardiac glycosides, alkaloids, terpenoids, and anthocyanins in all solvent extracts investigated but in variable concentrations.

The results also revealed that alkaloids and flavonoids were observed to be highly present (+++) in ethanolic extract of *Z.officinale* while they were present in trace quantities (+) in all other extracts as observed for glycosides which contrarily to the first were moderately present (++) in ethanolic extract. Similarly Shipra²⁹ reported a high presence of alkaloids in ethanolic extract of *Z.officinale* but flavonoids were moderately present. Alkaloids were not detected in polar solvent extract of *C.longa*³⁰. These facts correlated with those observed in the present study.

Tannins showed qualitative variations in both plant extracts screened. They were found to be present either in high (+++)

concentrations or in moderate (++) amounts in ethyl acetate and ethanolic extracts of *Z.officinale* respectively. However, in ethanolic extract of *C.longa* they were present in trace quantities (+) and completely absent (-) in ethyl acetate extract of the sample. This fact was also reported for $C.longa^{31}$ and some selected spices 30 .

Terpenoids were found to be present in high concentration (+++) in ethanolic extract of Z. officinale and C.longa while in ethyl acetate extracts of both plants they were present as trace compound(+). Similar fact was observed in Z.officinale extract of this solvent²⁸ but they were completely absent in C.longa³⁰.

Steroids showed qualitatively the same trend as observed for alkaloids and flavonoids. They were found to be highly present in ethanolic extract of *Z.officinale*, however, in all the other extracts of both plants they were detected in trace quantities. In contrast to the present results they were not detected in polar solvent extracts of *Z.officinale*²⁹.

Saponins were not present in any of the screened solvent extracts of both plants, except in the ethyl acetate extract of *Z.officinale* in which they were highly present. Similarly, saponins were reported to be absent in polar solvent extract of *C.longa*³¹, however they were highly present even in polar solent extract of *Z.officinale*²⁹.

The C.longa extracts of ethanol and ethyl acetate showed the high presence of anthocyanins and phenols, while in Z.officinale extracts they were present as trace compounds. They were not found in non-polar solvent extract (petroleum)²⁸. This fact was also observed in the case of carotenoids, which were completely absent in both Z.officinale extracts. However, they were found in a high concentration in ethyl acetate extract of C.longa and as trace compounds in ethanolic extract.

The tested compounds were found to be present in all the tested solvent extracts. It was found that they were more highly present in ethanolic extract of *Z.officinale* when compared to all the others plant extracts. The screened solvent extracts showed variations in phytochemical content. This can be explained by the variability of such factors as solubility in the solvent, polarity, the plant part nature (location, growing period), and

Phytochemical screening indicated that these spices contained the phytonutrients including alkaloids, glycosides, tannins, terpenoids, steroids, phenols carotenoids, saponins, flavonoids and anthocyanins.

interaction with other molecules presents in the samples.

Quantitative analysis of the plant rhizomes: The results of quantitative analysis of the plants are shown in Table-2 and indicate that alkaloids and flavonoids were the most abundant compounds, followed by saponins and phenols.

G.officinale contained the highest alkaloid and flavonoid contents of $11.28\pm2.05\%$ and $15.07\pm0.04\%$) while *C.longa* had the lowest of 07.33 ± 0.18 and $08.50\pm0.12\%$ for both compounds respectively).

The high alkaloid content reporded in *Z.officinale* (11.21%)¹⁵ was in line with the value recorded for the plants studied. These concentrations were found to be higher when compared with those recorded in other spices like garlic (2.54%)¹⁵ and capsicum species(1.8 and 5.3)(unpublished data).

Alkaloids are found in the leaves, and also in the seeds in little amounts and consequently in rhizomes³². This is in contrast with the result of the present work. An increased of alkaloid content was considered as depending of such factors like the favorable environmental conditions and nutrient supply³³.

For instance alkaloids stimulate the nervous systems and play role in blood pressure regulation. Alkaloids are pain relievers, tranquilizers and have also antimicrobial properties³⁴.

The flavonoid percent of ginger remained higher than the reported 5.56% ¹⁵. The same trend was observed when comparing the flavonoid content of *C.longa* with those of others spices such as pepper and garlic (6.38 & 1.16%). However this flavonoid amount was lower than that for *C.longa* which showed a high level of these compounds⁷. The flavonoids concentrations in plants can be influenced by factors such as genotype (species and variety) environment and harvesting period³⁵. The high phytochemical contents of the plants recorded in this work justified the multiples utilities and applications of these spices reported in folk medicine due to the claimed properties of alkaloids and flavonoids.

As observed for ginger¹⁵, the result also showed a low saponin content (02.13±0.07%). In contrast to alkaloids and flavonoids which were found in lower amounts in *C.longa*, saponins

 $(22.41\pm1.33\%)$ and phenols $(11.33\pm0.18\%)$ were the most abundant compounds detected in the sample. The saponin content values were found to be much higher than those reported for others spices like ginger (0.80%), pepper (7.40%) and garlic $(4.60\%)^{15}$.

For instance saponins are known in reduction of glucose level and in hypocholesterolemic properties. This could help users in fighting against heart diseases³⁶.

Phenols were the second most abundant compounds in tumeric after saponins. This result correlated with the fact observed in other work⁷ indicating a high level of polyphenol in turmeric than in ginger (174 & 55mg/100g respectively). Recent studies on tumeric have confirmed the fact that the sample contained a high level of phenolic compounds¹⁶.

Phenols are reported to be the main contributors of antioxidant activity in plants². This may be valuable for turmeric only due to the lower amount of phenols in ginger.

Anthocyanin constituents (0.40±3.92%) were found to have a lower concentration in ginger than in *C.longa* (02.33±0.16%). The finding that anthocyanins are the most abundant flavonoids of the red fruits³⁷ agree with the results of the present study since these compounds in the rhizomes as non colored samples were found in lower concentrations.

Anthocyanins play a role in health-preventive effects and reduce risks of, for example, age-related macular degeneration³⁸, they also reduce the risk of cardio vascular disorders³⁹.

Carotenoids were detected as trace compounds in both the plant samples with the smallest concentrations of 0.02±0.003 to 0.02±0.01%. This fact correlates with that observed for Ginger¹⁵. Moreover, carotenoids were not detected in garlic. It have been reported that the high values of carotenoids (29.3 &132.9mg/100g) lie to the red color of plant like pepper varieties⁴⁰. As observed for anthocyanins such finding may justify the lower concentrations recorded in the present study as non colored samples.

Glycosides were not detected in any of the plants screened in the present study.

The variations in phytochemical levels may be explained by different factors such as environmental conditions, harvesting season, sampling techniques, the disparity of these compounds, genetic differences, and solvent used⁴¹⁻⁴⁴.

The presence of phytochemicals in *Capsicum annum* and *Capsicum frutescens* determine the medicinal value of these spices and their importance in nutrition due to the biological properties of these compounds.

Conclusion

The comparative study of the two spices, ginger (Zingiber officinale) and turmeric (Curcuma longa) indicated that they contained phytochemical constituents with beneficial health effects in treating common chronic diseases. Thus they could be considered as sources of useful drugs for improving the health status of consumers and as food need in nutrition as a result of the presence of various compounds that are vital for good health and could be recommended in food supplementation.

Acknowledgement

Authors are thankful to Professor Attibayeba and the personal of the laboratory of Biology for laboratory facility and also the members of the laboratory of physical chemistry (Faculty of Sciences & Technics) for help to carry out analysis.

References

- 1. Kim J.K., Kim Y., Na K.M., Surh Y.J. and Kim T.Y. (2007). [6]-Gingerol prevents UVB-induced ROS production and COX-2 expression *in vitro* and *in vivo*. *Free Radical Res.*, 41, 603-614.
- 2. Maizura M., Aminah A. and Wan Aida W.M. (2011). Total phenolic content and antioxidant activity of kesum (*Polygonum minus*), ginger (*Zingiber officinale*) and turmeric (*Curcuma longa*) extract. *Int. Food Res. J.*, 18, 529-534.
- **3.** Medoua G.N., Egal A.A. and Oldewage-Theron H.W. (2009). Nutritional value and antioxidant capacity of lunch meals consumed by elderly people of Sharpeville, South Africa. *Food Chem.*, 115(1), 260-264.
- **4.** Kalaivani K., Senthil-Nathan S. and Murugesan A.G. (2012). Biological activity of selected Lamiaceae and Zingiberaceae plant essential oils against the dengue vector Aedes aegypti L. (Diptera: Culicidae). *Parasitology Res.*, 110(3), 1261-1268.
- Karna P., Chagani S., Gundala S.R., Rida P.C.G., Asif G., Sharma V., Gupta M.V. and Aneja R. (2011). Benefits of whole ginger extract in prostate cancer. *British J. Nutri.*, 107 (4), 473-484.
- **6.** El-Ghorab A.H., Nauman M., Anjum F.M., Hussain S. and Nadeem M. (2010). A comparative study on chemical composition and antioxidant activity of ginger (*Zingiber officinale*) and cumin (*Cuminum cyminum*). *J. Agric. Food Chem.*, 58(14), 8231-8237.
- 7. Trinidad P.T., Sagum R.S., de Leon M.P., Mallillin A.C. and Borlagdan M.P. (2012). *Zingiber Officinale* and *Curcuma Longa* as Potential Functional Foods/Ingredients. *Food and Public Health*, 2(2), 1-4.

- **8.** Jothivenkata K. and Moscow S. (2013). Validated method for estimation of Curcumin from different varieties of *curcuma longa. Int. J. Pharm. Biol. Sci.*, 4(1),1004-1010.
- **9.** Singh R., Mehta A., Mehta P. and Shukla K. (2011). Anthalmintic activity of rhizome extracts of *curcuma longa* and *Zingiber officcinale* (Zingiberaceae). *Int.J. pharm. Pharm. Sci.*, 3, 236-237.
- **10.** Kuttan G. and Kumar K.B., Guruvayoorappan C. and Kuttan R. (2007). Anti-tumor, anti-invarion and antimetastatic effects of curcumin. *Adv.Exp. Med. Biol.*, 595, 173-184.
- **11.** Gantait A., Barrman T. and Mukherjee P. (2011). Validated method for determination of Curcumin in turmeric Powder. *Indian J.Traditional Knowledge*, 10(2), 247-250.
- **12.** Maiti K., Mukherjee K., Gantait A., Saha B.P. and Mukherjee P.K. (2007). Curcumin-phospholipid complex: Preparation, therapeutic evaluation and pharmacokinetic study in rats. *Int. J. Pharm.*, 330, 155-163.
- **13.** Tangkanakul P., Auttaviboonkul P., Niyomwit B., Lowvitoon N., Charoenthamawat P. and Trakoontivakorn G. (2009). Antioxidant capacity, total phenolic content and nutritional composition of Asian foods after thermal processing. *Int. Food Res. J.*, 16, 571-580.
- **14.** Fapohunda S.O., Mnom J.U. and Fakeye F. (2012). Proximate analysis, phytochemical screening and antibacterial potentials of bitter cola Cinnamon, Ginger and banana peels. *Academia Arena*, 4(8),8-15.
- **15.** Otunola G.A., Oloyede O.B., Oladiji A.T. and Afolayan A.J. (2010). Comparative analysis of the chemical composition of three spices *Allium sativum* L. *Zingiber officinale* Rosc. and *Capsicum frutescens* L. commonly consumed in Nigeria. *Afr.J. Biotech.*, 9, 6927-6931.
- **16.** Kang C. and Kim E. (2010). Synergistic effect of curcumin and insulin on muscle cell glucose metabolism. *Food Chem.Toxicol.*, 48(8-9), 2366-2373.
- **17.** Trease G.E. and Evans W.C. (1989). A textbook of pharmacognosy. London: Bailliere, Tindall, 12th edn, 388, 480, 502, 535, 546.
- **18.** Sofowara A. (1993). Medicinal plants and Traditional the leaves of Ocimum gratissimum. *Sci. Res. Essay*, Medicine in Africa, Ibadan: Spectrum Books Ltd., 2, 163-166.
- **19.** Harborne J.B. (1984). Phytochemical methods, A guide to modern techniques of plant analysis. London: Chapman and Hall) 2nd edn.
- **20.** Harborne J.B. (1998). Phytoch jmemical methods, A guide to modern techniques of plant analysis. New York: Chapman and Hall, 3rd edn.
- **21.** Kokate C.K. et. al. (2001). Pharmacognosy. India: Mumbai, Nirali Prakasham, 16th edn.

- **22.** Aguzue O.C., Temilola A.F., Abubakar T.M., Jamal K.M. and Habibu A.S. (2012). Comparative chemical constituents of some desert fruits in Northern Nigeria. *Arch.Appl.Sci.Res.*, 4(2), 1061-1064.
- **23.** Boham A.B. and Kocipai A.C. (1994). Flavonoid and condensed tannins from Leaves of Hawaiian *vaccininum vaticulum* and *vicalycinium*. *Pacific Sci.*, 48, 458-463.
- **24.** Harborne J.B. (1973). Phytochemical Methods, A guide to modern techniques of plant analysis. London, New York:Chapman and Hall, 49-88.
- **25.** Obadoni B.O. and Ochuko P.O. (2001). Phytochemical studies and Comparative efficacy of the crude extracts of some homeostatic plants in Edo and Delta States of Nigeria. *Global J. Pure Appl. Sci.*, 8, 203-208.
- **26.** Onyeka E.U. and Nwambekwe I.O. (2007). Phytochemical profile of some green leafy vegetables in South East, Nigeria. *Nig. Food J.*, 25(1), 67-76.
- **27.** Iqbal H., Ullah R., Ullah R., Khurram M., Ulla N., Baseer A., Khan F.A., Rehman M., Khattak UR., Zahoor M. Khan J. and khan N. (2011). Phytochemical analysis of selected medicinal plants. *Afr.J. Biotech.*, 10(38), 7487-7492.
- **28.** Venkata K.S.N., Santhosh D., Narasimba R.D., Sanjeeva K.A. and Charles M.A. (2011). Preliminary phytochemical screening & anti diabetic activity of *Z.officinale*. *Int.J.Pharm.Life Sci.*, 2(12), 1287-1292.
- **29.** Bhargava Shipra, dhabhai Kshipra, batra Amla, Sharma Asha and malhotra Bharti (2012). *Z. officinale*: Chemical and phytochemical screening and evaluation of its antimicrobial activities. *J. Chem. Pharm. Res.*, 4(1), 360-364.
- **30.** Harsha N., Sridevi V., Lakshmi M.V.V.C., Lakshmi C. and Sataya D. (2013). Phytochemical analysis of some selected spices. *Int. J. Innov. Res. Sci. Eng. Technol.*, 2(11), 6618-6621.
- **31.** Saxena J. and Sahu R. (2012). Evaluation of phytochemical constituent in conventional & non conventional species of *curcuma*. *Int.Res.J.Pharm.*, 3(8), 203-204.
- **32.** Zehra M., Banerjee S., Naqvi A.A. and Kumar S. (1998). Variation in the growth and alkaloid production capability of the hairy roots of *Hyoscyamus albus*, *H. muticus* and their somatic hybrid. *Plant Sci.*, 136, 93-99.
- **33.** Liu Y.H, Liang Z.S., Chen B., Yang D.F. and Liu J.L. (2010). Elicitation of alkaloids in in vitro PLB (protocormlike body) cultures of Pinella ternate. *Enzyme Microb. Tech.*, 46, 28-31.

- 34. Adesuyi A.O., Elumm I.K., Adaramola F.B. and Nwokocha A.G.M. (2012). Nutritional and Phytochemical Screening of *Garcinia kola*. Adv. J.Food Sci. and Technol., 4(1), 9-14.
- **35.** Shaik S., Singh N. and Nicolas A. (2011). Comparison of the Selected Secondary Metabolite Content Present in the Cancer-Bush *Lessertia* (Sutherlandia) Frutescens L. Extracts. *Afr. J. Tradit. Complement. Altern. Med.*, 8(4), 429-434.
- **36.** Price K.R., Johnson L.J. and Fenwick G.R. (1987). The chemical and biological significance of saponins in foods and feeding stuffs, C.R.C. Crit. *Rev. Food Sci. Nutri.*, 26, 27-135.
- **37.** Pazmino-Duran A.E., Giusti M.M., Wrostad R.E. and Gloria B.A. (2001). Anthocyanins from *Oxalis triangularis* as potential food colorants. *Food Chem.*,75, 211-216.
- **38.** Jang Y.P., Zhou J., Nakanishi K. and Spanfow J.R. (2005). Anthocyanins protect against A2E photooxidation and membrane permeabilization in retinal pigment, epithelial cells. *Photochem. & Photobiol.*, 81, 529-536.
- **39.** Mazza G. (2005). Anthocyanins and hearth health. *Ann. Ist. Super. Sanita*, 45, 369-374.
- **40.** Vera-Guzmán A.M., Chávez-Servia J.L., Carrillo-Rodriguez J.C. and López M.G. (2011). Phytochemical Evaluation of Wild and Cultivated Pepper(*Capsicum annum L.* and *C.pubescens* Ruiz &Pav. from Oaxaca, Mexico, *Chilean. J.Agric.Res.*, 71(4), 578-585.
- **41.** Santos-Gomes P.C., Seabra R.M., Andrade P.B. and Fernandes-Ferreira M. (2002). Phenolic antioxidant compounds produced by in vitro shoots of sage (*Salvia officinalis L.*). *Plant Sci.*, 162, 981-987.
- **42.** Biglari F., AlKarkhi A.F.M. and Easa A.M. (2008). Antioxidant activity and phenolic content of various date palm (*Phoenix dactylifera*) fruits from Iran. *Food Chem.*, 107, 1636-1641.
- **43.** Jiang P., Burczynski F., Campbell C., Pierce G., Austria J.A. and Briggs C.J. (2007). Rutin and flavonoid contents in three buckwheat species *Fagopyrum esculentum*, *F. tataricum*, and *F. homotropicum* and their protective effects against lipid peroxidation. *Food Res. Int.*, 40, 356-364.
- **44.** Tijjani M.A., Abdurahaman F.I., Abba Y.S., Idris M., Baburo B.S.I., Mala G.A., Dungus M.H.N., Aji B.M. and Abubakar K.I. (2013). Evaluation of proximate and phytochemical Composition of leaves of *Annona Senegalensis Pers. J. Pharm. Sci. Innovat.*, 2(1), 7-9.