



Review Paper

Natural products as corrosion inhibitors for mild steel in H₂SO₄ solution: A Review

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Abstract

Corrosion is the destructive attack of a metal by chemical or electrochemical reaction with its environment. A percentage of inhibition was carried out by various methods like WL and electrochemical methods such as PDP and EIS. Natural products were used for the protection of MS materials from corrosion as they are biodegradable and eco-friendly. Nature of surface films produce on metal was studied by using various techniques like FT-IR, UV-Visible spectroscopy, EDX, AFM, GC-MS and SEM. The present review paper covers the research works (Period 2017-2018) carried out by various researchers on corrosion inhibition of MS in H₂SO₄ solutions by using a Natural products as green inhibitors.

Keywords: Mild Steel, H₂SO₄, Corrosion, Natural Products, PDP, EIS, SEM.

Introduction

Corrosion is a spontaneous physical or chemical attack on materials due to an interfacial reaction in an ambient environment resulting in the loss of material properties. A severe corrosion condition may result in tremendous economic loss as well as environmental impact to some extent. MS is also considered low carbon steel as it is defined as having a low carbon content of 0.05–0.25%. MS are extensively used in various industrial and manufacturing plants, including petroleum, chemical, petrochemical, construction, pulp and paper, and transportation industries due to its remarkable features such as low cost, easy availability, high strength, durability etc. MS corrosion occurs inevitably during the process of cleaning boilers, acid pickling, and de-scaling. H₂SO₄ is a strong acid and is used as a cleaner for rust, algae and scale from condensers and cooling tower¹. Sulfuric acid corrodes majority of most alloys and metals.

A corrosion inhibitor is generally referred to as a chemical substance that when applied in small quantities to a corrosive medium reduces the rate of corrosion of a metal or a metal alloy². The use of natural products as corrosion inhibitors is of enormous interest because they are environmentally friendly,

ecologically acceptable, readily available, inexpensive and can be used nowadays as ‘Green Inhibitors’. The inhibitor molecules can also adsorb on the mild steel surface based on donor-acceptor interactions between π -electrons of the aromatic/heterocyclic ring and vacant d-orbitals of surface iron.

In this review study indicates only Langmuir, Temkin, Freundlich adsorption isotherm found. The efficiency of these natural products extracts as corrosion inhibitors was calculated by using WL and electrochemical tests, such as PDP and EIS measurements. The objective of this review is to summarize the results regarding inhibition of MS corrosion in H₂SO₄ solution by using various Natural products.

Phytoconstituents present in Natural products

In this review, most of the Natural products found to contain active phytochemicals such as tannins, amino acids, flavonoids, saponins, phenols and alkaloid which have corrosion inhibitive effect.

Natural products used for inhibition of corrosion of MS in H₂SO₄ solutions are presented in Table-1.

Table-1: Natural products as corrosion inhibitor for MS in H₂SO₄ solution.

Inhibitor	Solution	Techniques(s)	Findings	I.E. _{max} (in %)	Ref. No.
<i>Acacia cyanophylla</i>	1 M H ₂ SO ₄	WL, EIS, AFM, FT-IR	Langmuir adsorption isotherm	83.80 WL, 86.95 EIS	3
<i>Adansonia digitata</i> (Baobab)	0.5 M H ₂ SO ₄	WL with time, EIS, PDP and SEM	Mixed inhibitor. Langmuir isotherm.	74.52 WL, 73.82 PDP, 73.97 EIS	4

<i>Aegle marmelos</i>	0.5 M H ₂ SO ₄	WL, PDP, EIS, SEM, AFM, FT-IR, UV-Vis, QCC	Mixed- type of inhibitor. Langmuir isotherm	83.08 WL, 83.08 PDP, 81.85 EIS	5
<i>Agave Americana</i>	0.5 M H ₂ SO ₄	PDP, EIS, SEM, FT-IR	Mixed type inhibitor. Langmuir adsorption isotherm	81.87 PDP	6
<i>Alkana tinctorial</i>	0.5 M H ₂ SO ₄	WL with temp., EIS, SEM, AFM, FT-IR, UV-Vis, QCC	Mixed inhibitor. Langmuir isotherm	90.46 WL, 91.63 PDP, 91.42 EIS	7
<i>Alstonia boonei</i>	0.5 M H ₂ SO ₄	WL with temp.	Langmuir isotherm.	83.0 WL	8
<i>Alternanthera bettzickiana</i>	5.0 M H ₂ SO ₄	WL with time and temp.	Langmuir and Temkin isotherm	Leaves 92.07 WL, Stem 89.95 WL, Root 88.41 WL	9
<i>Armoracia rusticana</i>	0.5 M H ₂ SO ₄	WL, PDP, EIS, SEM, AFM, FT-IR, UV-Vis SE, ATR, QCC	Mixed-type inhibitor. Langmuir adsorption isotherm	93.27 WL, 95.74 PDP, 94.04 EIS	10
<i>Artemisia vulgaris</i>	1 M H ₂ SO ₄	WL with time and temp., SEM, EDX	Langmuir adsorption isotherm	71.80 WL	11
<i>Asparagus racemosus</i>	0.5 M H ₂ SO ₄	WL, EIS, PDP, SEM, UV, AFM, FT-IR, QCC	Mixed inhibitor. Langmuir isotherm.	88.85 WL, 93.25 PDP, 90.56 EIS	12
<i>Aster koraiensis</i>	1 M H ₂ SO ₄	WL with temp., PDP, SEM, EDX, FT-IR	Mixed-type of inhibitor. Langmuir isotherm.	90.53 WL	13
<i>Azadirachta indica</i>	1 NH ₂ SO ₄	WL with temp., GC-MS, SEM	Langmuir isotherm.	65.6 WL	14
<i>Bauhinia tomentosa</i>	1 N H ₂ SO ₄	WL with temp., PDP, EIS, SEM, UV-Vis. and FT-IR	Mixed inhibitor. Temkin adsorption isotherm	86.63 WL, 84.17 PDP, 88.47 EIS	15
Black Liquor	0.5 M H ₂ SO ₄	WL with time, PDP, EIS, SEM	Cathodic-type of inhibitor	76.0 WL, 95.0 PDP, 89.0 EIS	16
<i>Bombax ceiba</i> Flower	0.5 M H ₂ SO ₄	WL, PDP, SEM.	Mixed inhibitor. Langmuir isotherm	90.84 WL, 96.29 PDP	17
<i>Butea monosperma</i>	0.5 M H ₂ SO ₄	WL, PDP, EIS, SEM, IR, AFM UV	Good inhibitor	87.45 WL, 97.94 PDP, 98.41 EIS	18
<i>Clerodendrum Splendens</i>	0.7, 1.2 and 2.2 M H ₂ SO ₄	WL with temp. & time, SEM, FT-IR	Langmuir adsorption isotherm	87.58 WL	19
<i>Corchorus olitorius</i>	0.5 M H ₂ SO ₄	WL with time and temp., PDP, EIS, GC-MS, SEM	Mixed-type of inhibitor. El-Awady adsorption isotherm.	93.0 WL	20
<i>Coreopsis tinctoria</i> (C. tinctoria)	0.5 M H ₂ SO ₄	WL, PDP, EIS, EDX, AAS, SEM, RS	Langmuir adsorption isotherm.	80.62 WL	21
<i>Coriandrum Sativum. L.</i>	0.5 M H ₂ SO ₄	OCP, PDP, EIS, FT-IR, SEM	Mixed inhibitor. Langmuir, Freundlich and Temkin adsorption isotherms.	96.7 PDP, 96.4 EIS	22
<i>Cuscuta reflexa</i>	0.5 M H ₂ SO ₄	WL, PDP, EIS, UV, FT-IR, AFM, SEM and DFT	Mixed-type of inhibitor. Langmuir adsorption isotherm.	95.47 PDP, 93.05 EIS	23
<i>Cyclea peltata</i>	1 N H ₂ SO ₄	WL	Langumir, Temkin and El-awady adsorption isotherm.	91.89 WL	24

<i>Delonix regia</i>	0.1 M H ₂ SO ₄	PDP, SEM, FT-IR, GC-MS	Mixed-type of inhibitor. Langmuir adsorption isotherm.	88.65 PDP	25
<i>Eriobotrya japonica</i> Lindl. (Loquat)	0.5 M H ₂ SO ₄	WL with temp. & time, PDP, EIS, FT-IR, HPLC, SEM	Cathodic inhibitor. Langmuir adsorption isotherm	96.2 WL, 96.3 PDP, 94.3 EIS	26
<i>Ervatamia coronaria</i>	1 M H ₂ SO ₄	WL, EIS, PDP, SEM, XRD	Mixed-type of inhibitor. Langmuir adsorption isotherm.	89.92 WL, 79.71 PDP, 71.91 EIS	27
<i>Fenugreek</i>	1 M H ₂ SO ₄	WL with temp., SEM, UV-Vis	Langmuir adsorption isotherm.	89.06 WL	28
<i>Ficus racemosa</i> (FR)	0.5 M H ₂ SO ₄	WL with temp., PDP, EIS	Mixed inhibitor. Langmuir isotherm.	--	29
<i>Ficus religiosa</i>	0.5 M H ₂ SO ₄	WL, PDP, EIS, SEM, AFM, FT-IR and UV-Vis, QCC	Mixed inhibitor. Langmuir isotherm.	88.29 WL, 92.26 PDP, 91.80 EIS	30
<i>Gardenia aqualla</i>	1 M H ₂ SO ₄	WL with temp.	Langmuir adsorption isotherm.	89.5 WL	31
<i>Gongronema latifolium</i>	5 M H ₂ SO ₄	WL with temp. and time, GM	Langmuir isotherm.	96.5 WL	32
<i>Ground nut</i>	1 M H ₂ SO ₄	WL with time and temp., RSM, SEM	Langmuir adsorption isotherm	85.9 WL	33
<i>Gum Tragacac powder</i>	1N H ₂ SO ₄	WL with temp, PDP, EIS	Mixed-type of inhibitor. Langmuir adsorption isotherm.	69.05 WL, 70.97 PDP, 86.89 EIS	34
<i>Houttuynia cordata</i>	0.5 M H ₂ SO ₄	WL with temp., PDP, EIS, SEM.	Mixed but predominantly cathodic inhibitor. Langmuir isotherm	89.39 WL, 94.4 PDP, 94.8 EIS	35
<i>Imperrata cylindrica</i>	1 M H ₂ SO ₄	WL with time and temp.	Good inhibitor.	76.58 WL	36
<i>Ipomoea batatas</i>	1.0 M H ₂ SO ₄	WL and GM	Langmuir adsorption isotherm	61.1 WL, 52.8 GM	37
<i>Jatropha tanjorensis</i>	1 M H ₂ SO ₄	WL with temp., HE.	Langmuir adsorption isotherm.	89.73 WL, 51.91 HE	38
<i>Lannea coromandelica</i>	1 M H ₂ SO ₄	WL with temp., PDP, EIS, XRD, FT-IR, SEM and AFM	Mixed-type of inhibitor. Langmuir isotherm.	89.0 WL, 88.5 PDP, 93.8 EIS	39
<i>Linum Usitatissimum</i>	1 M H ₂ SO ₄	WL with time and temp., PDP, EIS, SEM, FT-IR, LU Chromatogram	Mixed-type of inhibitor. Langmuir isotherm.	95.5 WL, 96.5 PDP, 96.5 EIS	40
<i>Luffa Aegyptiaca</i>	0.5 M H ₂ SO ₄	WL with time and temp., Colorometric, PDP, EIS, SE, FT-IR, Optical profilometry	Mixed-type of inhibitor. Langmuir and El-Awade isotherms.	94.29 WL, 92.84 PDP, 93.82 EIS	41
<i>Manihot esculenta</i> (Casava)	0.5 M H ₂ SO ₄	WL with temp., PDP, EDX, SEM, FT-IR	Mixed-type of inhibitor. Langmuir isotherm.	93.2 WL, 83.52 PDP	42
<i>Mentha spicata</i>	2.0 M H ₂ SO ₄	WL temp. & time, SEM, UV-Vis, FT-IR	Langmuir adsorption isotherm.	81.22 WL, 82.58 RN	43
<i>Microdesmis puberula</i>	1 M H ₂ SO ₄	WL with temp., HE	Freundlich adsorption isotherm.	83.25 WL, 82.82 HE	44

<i>Microdesmis puberula</i>	1 M H ₂ SO ₄	WL with temp.	Langmuir adsorption isotherm.	82.05 WL	45
<i>Mimosa pudica</i>	2.0 M H ₂ SO ₄	WL with temp., PDP, SEM, EDX	Freundlich isotherm.	72.89 WL	46
<i>Moringa oleifera</i>	1 N H ₂ SO ₄	WL with time, PDP, EIS, SEM, EDX, FT-IR	Mixed inhibitor.	96.00 WL, 56.57 PDP, 64.00 EIS	47
<i>Musa Sapientum</i>	2.0 M H ₂ SO ₄	WL	--	--	48
<i>Myristica fragrance</i>	0.5 M H ₂ SO ₄	WL, EIS, PDP, UV, IR, SEM, NMR, AFM and QCC	Mixed inhibitor.	83.27 WL, 87.42 PDP, 87.81 EIS	49
<i>Myristica fragrance</i>	0.5 M H ₂ SO ₄	WL, EIS, SEM, UV-Vis	Langmuir adsorption isotherm	87.81 WL	50
<i>Ocimum Canum</i>	0.5 M H ₂ SO ₄	WL with temp., GM	Langmuir isotherm.	89.2GM, 89.0 WL.	51
<i>Ocimum gratissimum (OG)</i>	0.5 M H ₂ SO ₄	WL, SEM, PDP, EIS, FT-IR	Mixed-type of inhibitor. Langmuir isotherm.	77.2 PP, 96.2 EIS	52
<i>Opuntia cochenillifera</i>	0.5 M H ₂ SO ₄	WL, EIS, PDP, SEM, FT-IR, EDX	Mixedinhibitor. Langmuir isotherm.	94.41 WL, 83.79 PDP, 85.18 EIS	53
<i>Pedaliium murex L.</i>	0.5 M H ₂ SO ₄	WL, PDP, EIS	Mixed inhibitor predomentely Cathodic. Langmuir isotherm.	90.47 WL, 87.51 PDP, 89.33 EIS	54
<i>Piper Guineense Squeezed</i>	2 M H ₂ SO ₄	WL with time and temp.	---	42.0 WL	55
<i>Pongamia Pinnata</i>	1 N H ₂ SO ₄	WL with time and temp., PDP, EIS, EDX, SEM, FT-IR, GC-MS	Mixed-type of inhibitor. Langmuir, Temkin and Freundlich isotherms.	---	56
<i>Poupartibirrea back</i>	0.5 M H ₂ SO ₄	WL with time	Efficient inhibitor.	70.0 WL	57
<i>Rice Husk</i>	1 M H ₂ SO ₄	WL with time, XRD.	Langmuir adsorption isotherms.	96.60 WL	58
<i>Saraca ashoka</i>	0.5 M H ₂ SO ₄	WL, EIS, PDP, SEM, UV-Vis., FT-IR, AFM, QCC	Mixed-type of inhibitor. Langmuir isotherm.	89.98 WL, 95.48 PDP, 93.09 EIS	59
<i>Sida acuta</i>	0.7, 1.2, 2.2 M H ₂ SO ₄	WL with time, ANN, FT-IR	--	96.37 WL	60
<i>Sida cordifolia</i>	0.5 M H ₂ SO ₄	WL, PDP, EIS, AFM, SEM	-	--	61
<i>Solenostemon Monostachyus</i>	1 M H ₂ SO ₄	WL with time and temp., HE	Freundlich isotherms.	80.35 WL, 70.53 HE	62
<i>Spathodea Campanulata</i>	1 N H ₂ SO ₄	WL with temp., PDP, SEM	Mixed inhibitor. Langmuir, Temkin and Freundlich isotherms.	82.74 WL	63
<i>Telfairia occidentalis Rind</i>	1 M H ₂ SO ₄	WL with temp., HE, SEM	Langmuir adsorption isotherms.	89.01 WL, 90.94 HE.	64
<i>Tephrosia villosa</i>	0.5 M H ₂ SO ₄	WL with temp., PDP, EIS	Mixed-type of inhibitor predominantly cathodic. Langmuir and Temkin adsorption isotherms.	97.64 WL, 94.12 PDP, 94.06 EIS	65
<i>Tribulus terrestris</i>	0.5, 0.75, 1.0 and 1.5 M H ₂ SO ₄	WL with temp.	Langmuir adsorption isotherms.	0.5N 92.81WL	66

<i>Valeriana wallichii</i>	0.5 M H ₂ SO ₄	WL, PDP, EIS, AFM, SEM, UV-Vis., FT-IR, QCC	Mixed inhibitor. Langmuir isotherm.	93.74 PDP	67
<i>Viscum album</i>	5.0 M H ₂ SO ₄	WL with temp., GM	Temkin isotherm.	99.99 WL	68
<i>Carcia papaya</i> (CP) and <i>Azadirachta indica</i> (AZ) (Neem) Leaf	1 and 2 M H ₂ SO ₄	WL with time	--	CP79.99WL, AZ75.49WL, CP+AZ 92.20WL	69
<i>Musa paradisiacal</i> (MP), <i>Azadirachta indica</i> (AZ), <i>Citrullus lanatus</i> (CL)	0.5 M H ₂ SO ₄	WL with time	--	MP68.73WL, AZ98.56WL, CL94.06 WL	70

Solutions: In this Review, H₂SO₄ solution was used as corrosive solutions.

Type of plant materials used as green inhibitors: Different parts of plant such as, Leaves^{3,8,13,15,19, 24,26,28,29,31,33,36-39,41,42,47,52, 54,60-63,65,69}, Fruits^{5,12,30,66}, Flower¹⁷, Root^{44,67}, Stem^{20,45,53}, Peels⁴⁸, Seeds^{4,22,40} and Plant extract⁵¹ were used as corrosion inhibitor.

Extracts used: Various solvents like Distilled water^{14,18,22,28,49, 54}, Double Distilled water^{3,15,24,39,65}, Ethanol^{5,7,9,12,20,23,30-32,42,44, 46-48,51,52,60,64,68,69}, Methanol^{4,11,43,57} and NaOH⁵⁸ were used for the preparation of plant extract.

Techniques: Various techniques like WL^{3,5,10,12,17,18,21,23,24,27,30, 37,48-50,52-54,59,61,67}, WL with time^{4,9,11,16,19,20,26,32,33,36,40,41,43,47,55-57, 58,60,62,69,70}, WL with temp.^{7-9,11,13-15,19,20,26,28,29,31,35} 36,38-46, 51, 52,55,56,62,63-66,149, GM^{32,37,51,68}, HE^{38,44,62,64}, PDP^{4-6,10,12,13,15,16-18,20-27,29,30,34,35,39-41,43,46,47,49,53,54,56,59,61,63,65,67} and EIS^{3-7,10,12,15,16,18,20-23, 26,27,29,30,34,35,40,41,47,49,50,52-54,56,59,61,65,67} were employed for calculation of percentage inhibition efficiency. Other techniques like OCP²², SE^{10,41}, RS²¹, Colorometry⁴¹, XRD^{27,39,58}, AFM^{3,5,7, 10,12,18,23,30,39,49,59,61,67}, RSM³³, DFT²³, HPLC²⁶, NMR⁴⁹, AAS^{21,60}, ATR¹⁰, ANN⁶⁰, QCC^{5,7,10,12,30,49,59,67} were also used.

Techniques for surface film analysis: Film formed on metal surface were studied by various techniques, like SEM^{4-7,10-15,16-23, 25-28,30,33,35,39,40,42,43,46,47,49,50,52,53,56,59,61,63,64,67}, FT-IR^{3,5-7,10,12,13,15,18, 19,22,23,25,26,30,39-43,47,49,52,53,56,59,60,67}, UV-Vis^{5,7,10,12,15,18,23,28,30,43,49,50, 59,67}, GC-MS^{14,20,25,56} and EDX^{11,13,21,42,46,47,53,56}.

Adsorption isotherms: In this review adsorption isotherms like Langmuir^{3-14,17,19,21-35,37-43,45,50,51-53,56,58,64-67}, Freundlich^{22,44,46,56,62, 63}, Temkin^{9,15,22,24,56,63,65,68} and El-Awaday^{20,24, 41} were suggested.

Conclusion

This review paper has summarized the research works carried out by various researchers on corrosion of MS in H₂SO₄ and its inhibition by using different natural plant extracts and techniques. Corrosion inhibition efficiency depends on many variables such as corrosion inhibitor, inhibitor concentrations, temp. and immersion time. In fact, plant extracts have numerous photochemical constituents that are capable to easily be

adsorbed onto metals, and thus inhibit their corrosion. Percentage I.E. of a corrosion inhibitor was evaluated by WL, PDP and EIS techniques. Other techniques such as SEM, FT-IR, AFM, GC-MS and EDX were also used to study film formed on surface of metal. Results obtained from weight loss data were in good agreement with PDP and EIS methods.

Abbreviations: AAS: atomic absorption spectroscopy, AFM: atomic force microscope, ANN: artificial neural network, ATR: alternate total reflection spectroscopy, CR: corrosion rate, DFT: density functional theory, EDS: energy-dispersive spectroscopy, EIS: electrochemical impedance spectroscopy, FT-IR: fourier-transform infrared spectroscopy, GC-MS: gas chromatography mass spectrometry, GM: gasometry method, H₂SO₄: sulfuric acid, HE: hydrogen evolution, HPLC: High pressure liquid chromatography, IEmax (%): maximum inhibition efficiency, KI: potassium iodide, MS: mild steel, NaCl: sodium chloride, NaOH: sodium hydroxide, NMR: nuclear magnetic resonance, OCP: open circuit potential, PDP: potentiodynamic polarization, QCC: quantum chemical calculations, QMC: Quantum mechanical calculations. RSM: response surface methodology, RS: Raman spectroscopy, RN: reaction number, SE: synergistic effect, SEM: scanning electron microscopy, UV-vis: ultraviolet-visible spectroscope or ultraviolet-visible spectrophotometry, WL: weight loss, XRD: X-ray diffraction.

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