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Antimicrobial Susceptibility Pattern of *Escherichia coli* isolated from urinary tract infections at Al-Quwayiyah General Hospital, Al- Quwayiyah, Kingdom of Saudi Arabia

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Abstract

Urinary tract infection (UTI) is the most common among all hospital acquired infections worldwide. In UTI, most common organism is E.coli. Organism responsible for the hospital acquired infection may have tendency to develop multiple drug resistance. E.coli acquired from the Al-Quwayiyah General Hospital source may differ in their resistant against antibiotics. This was a retrospective study conducted in an Al-Quwayiyah General hospital to know the prevalence and antibiogram of E.coli. Samples received include mid-stream clean catch urine, suprapubic aspirate and from Foley's catheter. All the plates were inspected for growth and the isolates were identified by observing colony morphology, Gram-stain characteristics and relevant biochemical tests. The isolates were tested for their antimicrobial susceptibility and the results were interpreted by vitek 2 methods, according to the guidelines of Clinical and Laboratory Standards Institute. Out of 96 samples tested, 53 (55.2%) were positive for E.coli. E.coli was highly resistant to Ampicillin (85.1%) followed by Piperacillin (66.03%) and least resistance was seen with Nalidixic acid (5.6%) and Nitrofurantoin (5.6%) followed by and Ciprofloxacin (11.3%). Knowledge of prevalence and antimicrobial susceptibility pattern of E. coli will help in selecting an appropriate antibiotic for empirical therapy. Formulation of hospital infection control committee and strict adherence to the guidelines of the committee will help in preventing the emergence of drug resistance.

Keywords: Antibiotic resistance pattern, urinary tract infection, E.coli.

Introduction

One of the commonest bacterial infections are the Urinary tract infections (UTIs), and the most common organism accounted for UTI is Escherichia coli $(E.coli)^{1,2}$. Despite the widespread availability of antibiotics, UTI is one of the most important causes of morbidity in general population and also the common cause of nosocomial infection among hospitalized patients³. Treatment of UTIs can be difficult because of recurrences and asymptomatic infections. The susceptibility patterns of uropathogens have been changing over the past years, both in community and nosocomial infections^{4, 5}.

Antibiotics are invariably used for the treatment of UTIs, though resistance to antibiotics has been reported all over the world, particularly in developing countries ⁶. Treatment of UTIs is a challenge due to the increasing level of antimicrobial resistance⁷. There is an increased emergence of antimicrobial resistance in the uropathogens, probably due to the empirical administration of anti bacterial therapy, even before the availability of the urine culture results, is a matter of concern worldwide⁸. The prevalence of antimicrobial resistance in

patients with UTI is increasing and can vary according to geographical and regional Location⁹.

For treatment of UTIs and prevention of antimicrobial resistance, knowledge of the common organisms responsible for UTIs and their antibiotic susceptibility patterns in specific geographical locations will help physicians in choosing an appropriate empirical treatment. There are only few studies of prevalence of UTI and antibiogram of E.coli in this part, hence the present study was undertaken to find out the prevalence and antibiogram of UTI due to E.coli. UTI is a serious health problem affecting millions of people each year. The prevalence of asymptomatic bacteriuria reported in Saudi Arabia was 5.3 $\%^{10}$. The most common organisms causing UTI are *E. coli* while Proteus, Klebsiella, Streptococcus and Staphylococcus epidermis also commonly the causative agents¹¹. The predominant organisms associated with UTI in Saudi Arabia are Gram negative bacteria which are highly resistant to commonly used oral agents¹². This present study was conducted to determine the Antimicrobial Susceptibility Pattern of Escherichia coli isolated from urinary tract infections at Al-Quwayiyah General Hospital, Al- Quwayiyah, Kingdom of Saudi Arabia.

Material and Methods

This was a retrospective study conducted in an Al-Quwayiyah General Hospital, Al-Quwayiyah, Saudi Arabia. Data was collected from Aug 2013 to Nov 2013 using the microbiological records of consecutive urine samples received in the laboratory during the study. All positive samples reports from both communities acquired and nosocomial UTI was included from various specialties. Urine samples were cultured using a 1µm calibrated loop onto CLED agar, blood agar and MacConkey agar plates, incubated at 37°C for 18-24 hours and the number of colonies was counted. All specimens were inoculated from CLED agar (Media, Ministry of Health) by semi quantitative method. The specimen yielding more than or equal to 10^5 organisms/ml of urine was interpreted as significant. Samples received include mid-stream clean catch urine, suprapubic aspirate and urine obtained from Foley's catheter. All the plates were inspected for growth and the isolates were identified by observing colony morphology. Gram-stain characteristics and relevant biochemical tests by Vitek 2 Instrument.

The isolates were tested for their antimicrobial susceptibility and the results were interpreted by modified Vitek 2 method¹³, according to the guidelines of Clinical and Laboratory Standards Institute¹⁴. The antibiotics tested were Cephalothin, Augmentin, Ampicillin, Piperacillin, Gentamicin, Nalidixic acid, Nitrofurantoin, Ciprofloxacin, Trimethoprim Sulfate, Cefoxitin, Cefpodoxime, ATCC *E.coli* 25922 was inoculated was used as control strain (Vitek 2 cards from Ministry of Health).

Results and Discussion

The total of 96 urine samples were collected from outpatient, Pediatrics, Emergency department, gynecology ward, female and male medical ward, out of which 53 (55.2%) were positive for *E. coli*. The age, gender and wise data of prevalence revealed that gram negative *E.coli* infection was found to be more prevalent female candidates (table 1 and figure 1). Highest number of cases was in the age group of 20-50 years, and maximum cases were from females (table 2). Highest resistance was seen with Ampicillin and least resistance to Nitrofurantoin and Nalidixicacid (figure 2).

Table-1
Age, Sex and Nationality distribution of <i>E.coli</i> from isolated
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cuses						
Age groups (years)	Nationality					
	Saudi		Non-Saudi		Total	
	М	F	М	F		
0-10	2	7	1	1	11	
11-20	2	2	-	-	4	
21-30	5	6	2	1	14	
31-40	1	5	1	2	9	
41-50	3	2	4	1	10	
50-90	1	4	-	-	5	
Total	14	26	8	5	53	

 Table-2

 Antibiotic Resistance Pattern of E.coli

Antibiotic	Number	Percentage
Cephalothin	15	28.3%
Augmentin	27	51.1%
Ampicillin	45	85.1%
Piperacillin	35	66.03%
Gentamicin	10	19.1%
Nalidixic acid	3	5.6%
Nitrofurantoin	3	5.6%
Ciprofloxacin	6	11.3%
Trimethoprim	29	54.7%
Sulfate		
Cefoxitin	7	13.2%
Cefpodoxime	17	32.07%

Sex and Nationality-wise distribution of E.coli from Isolated cases



Figure-1 The ratio of *E.coli* infected patients



Discussion: Knowledge on local prevalence and antimicrobial resistance trends among urinary isolates is important in guiding clinicians appropriate for appropriate empirical treatment of UTI. E. coli is the most prevalent pathogen contributing to this infection, but resistance is seen nearly 70-80% of the strains to the commonly used antibiotics¹⁵. In the present study E. coli accounted for 62.21% of all the positive cases. This is in accordance with the recent studies where the predominant organism isolated was E. coli¹⁶. In the present study highest resistance was seen with Ampicillin (85.1%), followed by Piperacillin (66.03%) and least resistance to Nalidixic acid (5.6%) and Nitrofurantoin (5.6%). This is in accordance with other studies where a high percentage of E. coli isolates were resistant to Ampicillin and Ofloxacin^{17, 18}. Fluoroquinolones are considered highly effective in treatment of UTI because of concentrating ability in urine and high renal clearance¹⁹. However due to wide spread use of Fluoroquinolones, there has been a report of evolving bacterial resistance to Fluoroquinolones²⁰. *E.coli* showed varied resistance among Aminoglycosides (56% to Gentamicin and 21.9% to Amikacin) the finding differs from the other study where resistance to Gentamicin and Amikacin was high²¹, but another study showed 40% resistance to Gentamicin and resistance to Amikacin was only 9%²². Resistance to Trimethoprim Sulfate (55.5%), Augmentin (52.7%), Cephalothin (27.7%). The findings are similar with the other studies $^{23, 24}$. Tazobactam in combination with piperacillin has an excellent clinical efficacy in various infections²⁵. Tazobactam seems to be the most promising beta lactamase inhibitor, which has unlike clavulanic acid and sulbactam, its own antibiotic activity²⁶. Resistance to Nitrofurantoin was 31.5%. In some studies Nitrofurantoin was found to be the most effective drug for UTI ^{27, 28}. Antimicrobial resistance pattern varies with time which might increase or decrease²⁹. Antibiotic susceptibility studies will help in early detection of development of antibiotic resistance and preserve powerful antibiotics like Imipenem for the treatment of life threatening infections.

Conclusion

In the present study, *E.coli* showed resistance to commonly used antibiotics. Antimicrobial susceptibility patterns vary with time and region. Prevalence and antibiotic susceptibility studies need to be conducted regularly, which will help in developing guidelines for treatment of UTI. The present study emphasizes the need for constant monitoring of susceptibility of uropathogens in different regions in order to rationalize the antibiotic use.

References

- Sharma S., Current understanding of Pathogenic Mechanisms in UTIs, Ann Natl Acad Med Sci., 33, 31-8 (1997)
- 2. Stamm W.E., Urinary tract infections and pyelonephritis, Chapter 269, In: Harrison's Principles of Internal

Medicine, 16th ed. Kasper DL, Braunwald E, Fauci AS, Hauser SL, Longo DL, Jameson JL, Eds. (McGraw-Hill, New York), 1715-21 (**2005**)

- **3.** Ronald A.R. and Pattulo M.S., The natural history of urinary infection in adults, *Med Clin North Am*, **75**, 299-312 (**1991**)
- 4. Manges A.R., Natarajan P., Solberg O.D., Dietrich P.S. and Riley L.W., The changing prevalence of drug-resistant *E coli* clonal groups in a community: Evidence for community outbreaks of urinary tract infections, *Epidemiol Infect*, **134**, 425-31 (**2006**)
- 5. Kahan N.R., Chinitz D.P., Waitman D.A., Dushnitzky D. and Kahan E., Shapiro M., Empiric treatment of uncomplicated urinary tract infection with fluoroquinolones in older women in Israel: Another lost treatment option? Ann Pharmacother **40**, 2223-7 (**2006**)
- 6. Lamikanra A. and Okeke I.N., A study of the effect of the urban/rural divide on the incidence of antibiotic resistance in Escherichia coli, *Biomed Lett*, **55**, 91-7 (**1997**)
- 7. Belet N., Işlek I., Belet U., Sunter A.T. and Kuçukoduk S., Comparison of trimethoprim-sulfamethoxazole, cephadroxil and cefprozil as prophylaxis for recurrent urinary tract infections in children, *J Chemother*, **16**, 77-81 (**2004**)
- Oladeinde B.H., Omoregie R., Olley M. and Anunibe J.A., Urinary tract infections in a rural community of Nigeria, *N Am J Med Sci*, 3(2), 75-7 (2011)
- 9. Karlowsky J.A., Kelly L.J., Thornsberry C., Jones M.E. and Sahm D.F., Trends in antimicrobial resistance among urinary tract infection isolates of Escherichia coli from female outpatients in the United States, *Antimicrob Agents Chemother*, **46**, 2540-5 (**2002**)
- Omar E.E. and ElHaj A.J., Urinary tract infections in school children in Saudi Arabia, *Med. Dig.*, 18, 3-7 (1992)
- **11.** Bonadio M., Costarelli S., Morelli G. and Tartaglia T., The influence of diabetes mellitus on the spectrum of uropathogens and the antimicrobial resistance in elderly adult patients with urinary tract infection, *BMC Infect Dis*, 6:54. Doi: 10.1186/1471-2334-6-54 (**2006**)
- 12. Al-Harthi A.A. and Al-Fifi S.H., Antibiotic resistance pattern and empirical therapy for urinary tract infections in children, *Saudi Med J.*, **29(6)**, 854-8 (**2008**)
- 13. Vitek Method http://www.biomerieux-USA.com/servlet/srt/bio/usa/dynPage?node=APRL_NW S_CS2 (2013)
- 14. Clinical Laboratories Standards Institute (CLSI). Performance of standards for antimicrobial disk

susceptibility tests; approved standards, 10th ed. M02-A10. Vol 29. Wayne, PA: CLSI; (**2009**)

- **15.** Kapil A., The challenge of antibiotic resistance: Need to contemplate, *Indian J Med Res.*, **121**, 83-91 (**2005**)
- 16. Mandal J., Acharya N.S., Buddhapriya D. and Parija S.C., Antibiotic resistance pattern among common bacterial uropathogens with a special reference to ciprofloxacin resistant Escherichia coli, *Indian J Med Res.*, 136(5), 842-9 (2012)
- 17. Manjunath G., Prakash R., Vamseedhar Annam K.S., The changing trends in the spectrum of the antimicrobial drug resistance pattern of the uropathogens which were isolated from hospitals and community patients with urinary tract infections in Tumkur and Bangalore, *Int J Biol Med Res.*, 2(2), 504-07 (2011)
- Gupta N., Kundra S., Sharma A., Gautam V. and Arora D.R., Antimicrobial susceptibility of uropathogens in India, *J Infect Dis Antimicrob Agents*, 24, 13-8 (2007)
- **19.** Piddock L.J.V. and Wise R., Mechanism of resistance to quinolones and clinical prospective, *J Antimicrobial Chemother*, **23**, 427-83 (**1989**)
- **20.** Al- Tawfiq J.A., Increasing antibiotic resistance among isolates of Escherichia coli recovered from inpatient and outpatients in a Saudi Arabian hospital, *Infect Control Hosp Epidemiol*, **27**, 748-53 (**2006**)
- **21.** Mohanty S., Kapil A., Das B.K. and Dhawan B., Antimicrobial resistance profile of nosocomial Antibiotic Resistance Pattern of UTI uropathogens in a tertiary care hospital, *Indian J Med Sci.*, **57**, 148-54 (**2003**)

- Gupta V., Yadav A., Joshi R.M., Antibiotic resistance pattern in uropathogen, *Indian J Med Microbial*, 20, 96-8 (2002)
- Anuradha K., Sailaja V.V., Umabala P., Satheesh T. and Lakshmi V., Sensitivity pattern of gram negative bacilli to three β- lactam/ β- lactamase inhibitor combination using the automated API system, *Indian J Med Microbial*, 25, 203-8 (2007)
- 24. Akram M., Shahid M., Khan A.U., Etiology and antibiotic resistant patterns of community acquired urinary tract infections in JNMC Hospital Aligarh, India, *Ann Clin Microbiol Antimicrob*, 6, 4 (2007)
- 25. Niki Y., Fundamental and clinical studies on betalactamase inhibihitors, *Nippon Rinssho*, 59, 771-6 (2001)
- 26. Blahova J., Hupkova M. and Kremery V., Sr. The effectivesness of so called potentiated penicillins (augmentin and tazobactam) in vitro. Cas Lek cesk, 134, 558-61 (1995)
- 27. Sahm D.F., Thornsberry C., Mayfield D.C., Jones M.E., Karlowsky J.A., Multidrug- resistant urinary isolates of Escherichia coli: prevalence and patient demographics in the United States in 2000, Antimicrob Agents Chemother, **45**, 1402-6 (**2001**)
- **28.** Sunil Kumar Biradar, Srikanth, Praveen Kumar Doddamani, Prevalence and anti-biogram of uropathogens in a tertiary care hospital, *World Journal of Pharmaceutical research*, **2**(**5**), 1534-43 (**2013**)
- 29. Dyer I.E., Sankary I.M., Dawson J.O., Antibiotic Resistance in Bacterial Urinary Tract Infections, 1991 to 1997, *WJM*, 169(5), 265-8 (1998)