Biosorption of Heavy Metals from Aqueous solution using Mangrove fern Acrostichum aureum L. leaf Biomass as a Sorbent

Lobo Soniya M. and Gulimane Krishnakumar

Department of Applied Botany, Mangalore University, Mangalagangothri-574199, Mangalore, Karnataka, INDIA

Available online at: www.isca.in, www.isca.me

Received 17th July 2015, revised 16th September 2015, accepted 25th October 2015

Abstract

Investigation was carried out to analyze the biosorption behavior of the Mangrove fern (Acrostichum aureum L.) leaf biomass, for the removal of heavy metals from the aqueous solution. The uptake for Cu (II), Pb (II) and Zn (II) were 46.86 ± 0.54 , 47.21 ± 0.65 and 47.81 ± 0.43 mg g⁻¹ respectively when 500 mg L⁻¹ of the metal solution was used. The Langmuir and Freundlich equilibrium adsorption isotherms were studied and data obtained from equilibrium experiment are found well fitting with the Freundlich model than the Langmuir model with correlation coefficient of 0.99. Kinetic studies indicated that biosorption of heavy metal by the mangrove fern followed well the pseudo second order. The presence of hydroxyl, carboxyl, and amino groups in the biomass and their role in metal ion adsorption process was confirmed by FTIR studies. By this study plentifully accessible a new biosorbent for the exclusion of heavy metals from the polluted water was identified.

Keywords: Acrostichum aureum, biosorption, heavy metal.

Introduction

Heavy metal pollution is endangering the aquatic systems and affecting the water quality. Studies on the sorption of metal ions from aqueous solution are vital in water pollution control. Increase in heavy metal pollution is associated with population growth and industrial progress. Heavy metal contamination is caused by industrial emissions, mining, smelting and application of fertilizers and sewage sludge contaminated with heavy metals¹. Heavy metals including copper (Cu), and zinc (Zn) at high concentrations induce phytotoxicities in plants^{2,3}. Heavy metals may cause haemolysis, gastrointestinal, skin, liver and kidney disorders⁴. The elimination of pollutant metal ions from the wastewaters is therefore important.

Ionic exchange, oxidation–reduction, membrane filtration, adsorption chemical precipitation and reverse osmosis, are cost-demanding methods used in pollution treatments^{5,6}. Biosorption, is the capacity of some biomaterials to remove heavy metals from the aqueous solutions. It is an alternative to the conventional technologies for heavy metal removal from the polluted effluents⁷. In the recent years various plant materials were tried for biosorption. Leaves of poplar, neem, peepal, maize, Ulmus, *Araucaria heterophylla* have been studied to find out metal removal potential⁸⁻¹³.

Biosorption studies on Tree fern for the removal of lead ions from aqueous solution bracken fern in the elimination of mercury and Lady fern in the removal of Zn and Cu from aqueous solution were carried out to investigate the potential of pteridophytes in bioadsorption ¹⁴⁻¹⁶. *Acrostichum aureum* L. is a mangrove fern growing luxuriantly in coastal regions of Karnataka, India. This fern shows clumped distribution and

produces golden yellow leaves; rhizome paste is used in wound treatment¹⁷. There are no reports on the major applications of leaves mainly in bioremediation. The present study investigates whether the mature leaf biomass of mangrove fern *Acrostichum aureum* can be used remove Lead, Copper and Zinc ions from aqueous solutions as a sorbent

Material and Methods

Effect of pH, Temperature, Contact Time and Initial Concentration on Metal Biosorption: Leaves of A.aureum was oven dried at 60°C, powdered, sieved and preserved in sealed polythene bags. Stock (1,000 mg L⁻¹) of Cu (II) from CuSO4, Pb (II) using Pb(NO₃)₂ and Zn (II) from ZnSO₄.6H₂O solutions in deionized water were prepared. The adsorption study was performed with 50 ml of metal solution (300 mg L⁻¹) in 100-ml conical flasks. Glassware's used for experimental purposes were washed in 20% nitric acid and rinsed with distilled water to eliminate any interference by other metals. The pH of solutions was adjusted from 2 to 6. The desired pH was adjusted with 1N NaOH and 1N HCl. Biomass of 0.5 g weight was taken in the each conical flask. The temperature effect of on biosorption was studied at 10°C, 20°C, 30°C, 40°C and 50°C. The metal ion concentration was determined after 100 minutes of incubation. To examine the effect of contact time on biosorption samples were withdrawn at different time intervals (20, 40, 60, 80 and 100 minutes) and the metal ion in the supernatant was determined using a flame atomized atomic absorption spectrophotometer (GBC 932 plus AVANTA spectrophotometer). Initial concentration was varied from 50 to 500 mg L⁻¹ of the metal ion to study the effect of on biosorption. Experiments were conducted in triplicates.

Int. Res. J. Environment Sci.

Vol. **4(11)**, 25-31, November **(2015)**

Adsorption Isotherms: The biosorption potential (q_{eq}) at equilibrium was calculated as follows:

$$q_{eq} = \frac{c_o - c_{eq}}{W} \times V \tag{1}$$

Where: C_o was the initial metal ion concentration (mg L^{-1}); C_{eq} the metal ion concentration at equilibrium (mg L^{-1}); V the volume of solution used (L); and W was weight of the dry leaf powder (g).

To investigate the sorption isotherm, two equilibrium models, the Langmuir and the Freundlich isotherm equations, were analysed. The Langmuir sorption isotherm is useful in describing sorption of a solute from a liquid solution¹⁸.

$$\frac{c_e}{q_e} = \frac{1}{\kappa_L q_{max}} + \frac{1}{q_{max}} C \tag{2}$$

Where K_L is the Langmuir constant linked to the adsorption energy and q_{max} is the maximum adsorption capacity (mg g⁻¹), Q_e is the equilibrium sorption capacity (mg g⁻¹), Q_e the equilibrium liquid phase concentration (mg L^{-1}). Values of Langmuir parameters q_{max} (monolayer sorption saturation capacity) and absorption constant K_L were obtained from the slope and intercept of the linear plot of $C_{e/qe}$ versus C_e .

The Freundlich isotherm¹⁹, which is the first known sorption isotherm, is expressed by the following equation

$$\log q_e = \log K_F + \frac{1}{\pi} \log C_e \tag{3}$$

Where: q_e is the equilibrium sorption capacity (mg g⁻¹), C_e the equilibrium liquid phase concentration (mg L⁻¹). K_F and n are isotherm constants.

Kinetic studies: The pseudo-first-order, pseudo-second-order and intraparticle diffusion equations were applied to the experimental data²⁰⁻²².

Linearized form of pseudo first order rate equation as described here is used.

$$Log(q_{eq} - q_t) = lnq_{eq} - K_1 t \tag{4}$$

Where q_{eq} and q_t are the metal ions adsorbed at equilibrium (mg g^{-1}) and at time t (min), respectively, and K_1 is the rate constant of the first order adsorption equation (min⁻¹). The sorption rate constants (K_1) can be determined by plotting ln (q_{eq} – q_t) versus t.

Experimental data was further analyzed using pseudo second order kinetic equation.

$$\frac{t}{qt} = \frac{1}{K_2 q_e^2} + \frac{1}{q_e} t \tag{5}$$

Where: K_2 is the rate constant of pseudo-second-order adsorption, t is the contact time (min), q_e (mg/g) and q_t (mg/g) are the amount of the solute adsorbed at equilibrium and at time t.

Weber and Morris²² proposed a model to examine the intraparticle diffusion.

$$q_t = K_{id} t^{(1/2)} (6$$

Where: q_t is the amount of adsorbed metal ions on the biomass at time t (mg g⁻¹). The intraparticle rate constant, K_{id} was obtained from the slope of the plot.

FTIR analysis: For FTIR analysis was carried out for control biomass sample and metal-loaded samples which was dried and blended with KBr. The FTIR spectra were collected at transmission mode (4,000–400 cm⁻¹) using a (IR Prestige-21Shimadzu) FTIR spectrophotometer.

Results and Discussion

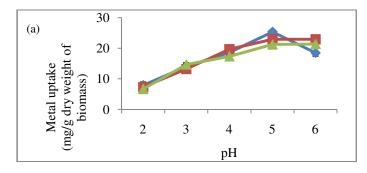
Effect of pH: The pH of the solution is the important parameters which influence the adsorption of metal ions²³. Uptake of Cu (II), Pb (II) and Zn (II) by biomass increased with an increase in pH of the solution (figure-2a). Studies on biosorption using neem and F. religiosa leaves also reported enhanced biosorption with increased pH ^{8, 9}. Biomass was most competent at pH 6.0 in removing Cu (II) and Zn (II) ions. Above this pH Cu (II) ions will precipitate ²⁴. In case of Pb (II) ions, the highest adsorption was noted at pH 5.0. With increasing pH, formation of lead hydroxide results in decreased adsorption. Functional groups get protonated at lower pH values which will avoid the positively charged metal ions to their surface⁹. Functional groups achieve negative charge at higher pH values and have a high attraction for the metal ions with positive charge, which may results in improved biosorption of the metal ions.

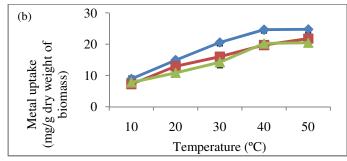
Effect of Temperature: Absorption rate increased from 7.45 to 20.34, 8.84 to 24.45, 7.19 to 19.5 mg/g for Cu, Pb and Zn respectively with temperature (figure-2b). Increased biosorption with temperature were also reported in tree fern biomass by Ho and Wang¹⁴. The temperature generally activates the process by increasing the heat or mass transport processes¹³. Temperature increase causes activation of adsorption sites²⁵. The biomass of *A. aureum* would include many types of binding sites and the effect of temperature on each binding site would differ and it will change the overall metal adsorption process.

Effect of contact time: From the experiment it was prominent that Cu (II) ions uptake was most quick compared to Pb (II) or Zn(II) ions. Smaller ion radius of the Copper ions may be responsible for quick adsorption²⁶. The equilibrium was attained within 1 h. Repulsive forces between the solute molecules on the adsorbent and aqueous solution may also contribute to the slowing down of uptake of metal at equilibrium¹³.

Effect of Initial Concentration of Metal Ions: The study has shown that biosorption capacities (q_{eq}) are influenced by initial concentration of the metal ions. The uptake was enhanced from 3.56 ± 0.43 , to 46.32 ± 1.02 , 4.37 ± 0.68 to 47.34 ± 1.43 , 3.83 ± 0.54 to 47.67 ± 1.23 mg g⁻¹ for, Pb (II), Zn (II) and Cu (II), respectively when initial concentration was varied from 50 to 500 mg L⁻¹. According to Lata et al.²⁷ initial metal

concentrations increase will acts as a positive driving force in overcoming mass transfer resistance of metal ions between the aqueous and solid phases. This would, in turn, enhance the uptake of metal ions. Similar observations were also made by earlier investigators in their studies on adsorption of metals on plant biomass ^{14,13}.





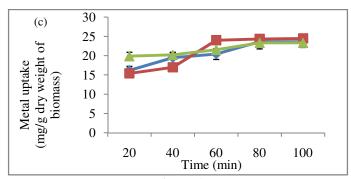


Figure-1

(a) Effect of initial pH on metal biosorption by *A. aureum* [Ci=300 mg L⁻¹, leaf powder =0.5g, time (t) = 80 min] (b) Effect of temperature on metal biosorption by A. aureum [Ci=300 mg L⁻¹, leaf powder =0.5g, t=80 min. and T=10°C, 20°C, 30°C, 40°C and 50°C] (c) Effect of contact time on metal biosorption [Ci=300 mg L⁻¹, leaf powder =0.5g, t=20, 40, 60, 80 and 100 min], [Cu (II) (square), Pb (II) (diamond) and Zn (II) (triangle)]

Equilibrium isotherm: According to Langmuir isotherm finite numbers of binding sites are available on the solid surface of the adsorbent with uniform energies for the adsorption. In a monolayer adsorption there will be no interaction between the metal ions adsorbed on the adsorbent surface. The correlation coefficients for Langmuir isotherm were 0.96 for Pb (II) and Zn

(II), 0.98 for Cu (II) respectively (table-1). Metal sorbing capacity of sorbent varies with various sorbents and sources. For example q_{max} of Pb determined using biomass of tree fern is 40mg/g and for Zn and Cu using lady fern was 0.073, 0.051 mg/g respectively 14,16 . However q_{max} reported in the present study is lower than the tree fern and significantly higher than lady fern.

The Freundlich isotherm suggests that on the heterogeneous surfaces of the adsorbent binding sites are not equivalent. It also indicates that sorption energy exponentially reduces on completion of the sorptional centers of an adsorbent. For a linear adsorption n = 1, for a chemical process in absorption n < 11 and n > 1 if it is a physical process. In this study Freundlich constant n was greater than 1, indicating that adsorption process here is a physical process and sorbate intensity on the surface of the sorbent was high, revealing favourable sorption even at high metal concentration. The R^2 values show fitness of the experimental data to the isotherm models. This also indicates high adsorption capacity of the sorbent and metal ions could be separated from its aqueous solution efficiently by the sorbent. The correlation coefficients value for Freundlich isotherm were 0.99 this indicating that the heavy metal sorption data fitted well to the Freundlich model.

Adsorption kinetics: For the pseudo first-order model ln $(q_{eq}-q_t)$ versus t was plotted. The correlation coefficients (R^2) for Cu (II), Pb (II), and Zn (II) adsorption onto biomass were 0.93, 0.90, and 0.83, respectively. The linear plots of t/qt versus t (figure-5) of the pseudo second- order for the adsorption of Cu (II), Pb (II), and Zn (II) ions onto biomass was plotted. The rate constants (K_2) , the q_{eq} and the R^2 values are given in the table-2. The correlation coefficients obtained was 0.99 for Cu (II), Pb (II), and Zn (II) adsorption. The R^2 values suggest that the adsorption fits well to the pseudo-second-order kinetic equation.

The results of intrapartical diffusion can be represented by a linear relationship, but the linear plot did not pass through the origin and the curve is dual natured due to variation in initial and final adsorption experiment.

FTIR analysis: Functional groups of the biomass were studied through FTIR spectra, which is a valuable tool for functional group analysis. Figure-2 to 5 shows the functional groups. IR peak observed at 3,421 cm⁻¹ due to stretching vibration of OH group indicated the involvement of hydroxyl group in metal binding ²⁶. Shifting of 2,372 cm⁻¹ to 2,374 cm⁻¹ for Pb, 2,376 cm⁻¹ for Zn is due to asymmetric stretching of -N=C=0 group. Shift in the wave number from 2,339 to 2,341 cm⁻¹ for Cu, Pb and Zn is due to the stretching vibration of N-H or C=O group. This indicates that oxygen and nitrogen are involved in metal adsorption. C-H stretching of methyl, methylene and methyne group was observed when band in region of 2,926 cm⁻¹ shifted to 2922 cm⁻¹ in Cu and Pb, 2,924 cm⁻¹ in Zn treated leaf powder. Broadening and stretching of the bond characteristic of C=O groups of amide linkage at 1,654 to 1,651 cm⁻¹ indicates the

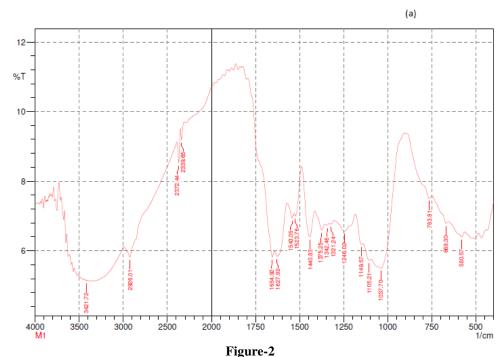
interaction of metals with C=O groups of the biomass. IR peak in the region of 1,440 shifted to 1,444 cm⁻¹ for Pb and Cu loaded leaf powder showed the participation of aliphatic or aromatic (C-H) group in the plane deformation vibration of methyl, methylene and methoxy groups. Peak shift in the region 1,321 to 1,317 cm⁻¹ indicate the aromatic amines involvement in

adsorption. Band in the region 669 cm⁻¹ shifted to 665 cm⁻¹ indicating the involvement of amide nitrogen in biosorption. After the binding of metal ion on the biomass, changes in intensity and shift in the position of peaks occur indicating the removal of metal ions by leaf biomass.

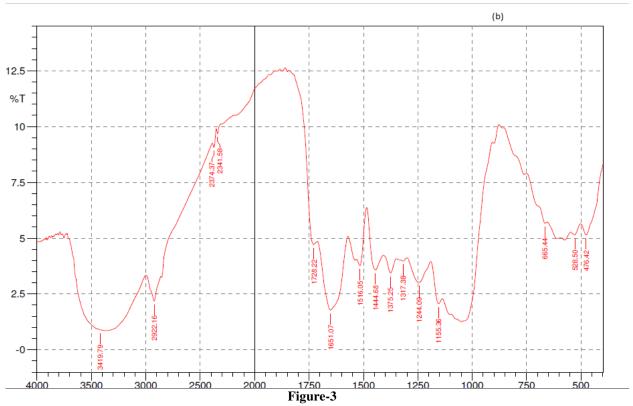
Metal	Langmuir isotherm			Freundlich isotherm		
	q m (mg g ⁻¹)	K L(L mg g ⁻¹)	\mathbb{R}^2	KF(mg g ⁻¹)	n	\mathbb{R}^2
Pb(II)	28.571	0.39	0.966	2.12	3.69	0.994
Cu(II)	27.77	0.51	0.983	2.15	3.703	0.993
Zn(II)	37.03	0.211	0.966	2.37	3.816	0.993

Table-2
Pseudo-second-order and intraparticle diffusion parameters for the biosorption of metal ions onto A. aureum leaf powder

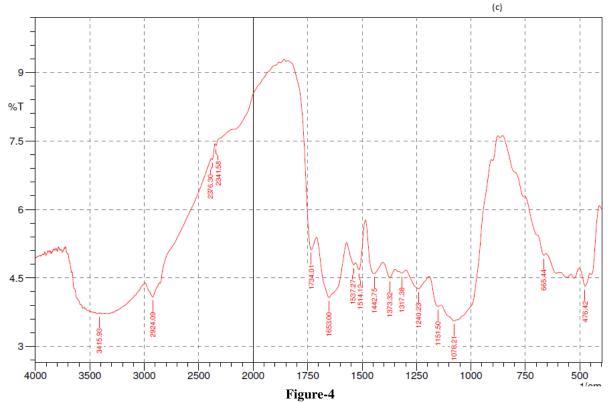
Metal	Pseudo-second-order			Intraparticle diffusion		
	\mathbf{K}_2	$ m q_{eq}$	R^2	K _i (mg/g min ^{-1/2})	Intercept (mg g ⁻¹)	R^2
Pb(II)	2	27.77	0.999	0.19	14.98	0.926
Cu(II)	3	28.57	0.955	0.1	18.63	0.925
Zn(II)	3	27.77	0.999	0.25	13.39	0.815



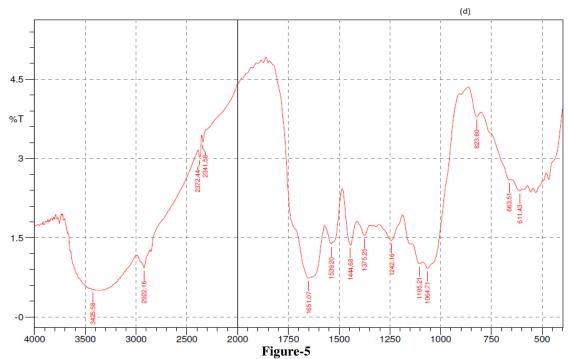
FTIR spectra of A. aureum leaf powder before the biosorption of heavy metals



FTIR spectra of A. aureum leaf powder after the biosorption of Pb (II)



FTIR spectra of A. aureum leaf powder after the biosorption of Zn (II)



FTIR spectra of A. aureum leaf powder after the biosorption of Cu (II)

Conclusion

Biosorption of heavy metals was investigated. *A. aureum* is a suitable sorbent for the removal of Cu(II), Pb(II) and Zn(II) from the aqueous solution. The study indicated that the maximum uptake for Cu(II), Pb(II) and Zn(II) were 46.86 ± 0.54, 47.21± 0.65 and 47.81± 0.43 mg g⁻¹ respectively at pH 5 when 500mg L⁻¹ of the metal solution were used at 40°C. Metal removal was a function of pH, temperature, time and initial concentration of metal ions in the solution. Freundlich isotherm fitted well to the adsorption data rather the Langmuir isotherm. FTIR studies revealed the presence of hydroxyl, carboxyl, and amino groups in the biomass and proved possible involvement of these groups in metal ion adsorption process. By this study the potential of a new cost-effective and plentifully available bioadsorbent for the removal of heavy metal ions from aqueous solutions has been identified.

Acknowledgement

The author (Soniya Marsi Lobo) would like to thank Department of Science and technology, New Delhi for the financial support through Inspire Fellowship.

References

- 1. Anthony G. K., Balwant S. and Naveen P. B., Heavy metal tolerance in common fern species, *Austral. J. Bot.*, **55**, 63–73 (2007)
- **2.** Chatterjee J. and Chatterjee C., Phytoxicity of cobalt, chromium and copper in Cauliflower, *Environ. Pollut.*,

109. 69–74 (**2000**)

- **3.** Cuypers A., Vangronsveld J. and Clijsters H., Peroxidases in roots and primary leaves of Phaseolus vulgaris Copper and Zinc Phytotoxicity: A comparison, *J. Plant Physiol.*, **159**, 869–876 (**2002**)
- **4.** Rengaraj S., Kim Y., Joo C.K., and Yi J., Removal of copper from aqueous solution by aminated and protonated mesoporous aluminas: Kinetics and equilibrium, *J. Colloid. Interface. Sci.*, **273**, 14–21 (**2004**)
- 5. Chong K.H. and Volesky B., Description of two-metal biosorption equilibria by Langmuir-type models, *Biotechnol. Bioeng.*, 47, 1–10 (1995)
- **6.** Elouear Z., Bouzid J., and Boujelben N., Removal of nickel and cadmium from aqueous solutions by sewage sludge ash: Study in single and binary systems, *Environ. Technol.* **30**, 561–570 (**2009**)
- 7. Davis T.A., Volesky B. and R.H.S.F. Vieira, Sargassum seaweed as biosorbent for heavy metals, *Water Res.* 34 (17), 4270 4278 (2000)
- **8.** Arshad M., Zafar M. N., Younis S. and Nadeema R., The use of neem biomass for the biosorption of zinc from aqueous solutions, *J. Hazard. Mater.* **157**, 534–540 **(2008)**
- Qaiser S., Saleemi A.R. and Umar M., Biosorption of lead from aqueous solution by Ficus religiosa leaves: Batch and column study, *J. Hazard. Mater.* 166, 998– 1005 (2009)

- **10.** Al-Masri M.S., Amin Y., Al-Akel B. and T. Al-Naama, Biosorption of Cadmium, Lead, and Uranium by Powder of Poplar Leaves and Branches, Appl. *Biochem. Biotechnol.*, **160(4)**, 976–87 (**2009**)
- **11.** Babarinde N. A. A., Oyesiku O.O., Babalola J.O. and Olatunji J.O., Isothermal and thermodynamic studies of the biosorption of zinc (II) ions by *Calymperes erosum* I., *Journal of Applied Sciences Research*, **4**, 716–721 (**2008**)
- **12.** Mahvi A.H., Gholami F. and Nazmara S., Cadmium biosorption from wastewater by ulmus leaves and their ash, *Eur. J. Scient. Res.* **23**, 197–203 (**2008**)
- 13. Sarada B., Krishna P.M., Kishore K.K. and Murthy Ch,V.R., Potential use of leaf biomass, *Araucaria heterophylla* for removal of Pb⁺², *Int. J. Phytorem.*, 15(8), 756–773 (2013)
- **14.** Ho Y.S. and Wang C.C., Pseudo-isotherms for the sorption of cadmium ion onto tree fern, *Process Biochem*, **39**, 761–765 (**2004**)
- **15.** Leticia C., Vasilism A., Pablo L., Jos´e L.B., Roberto H., and Manuel E.S.V., A dynamic proof of mercury elimination from solution through a combined sorption–reduction process. *Bioresour. Technol.*, **101**, 8969–8974 (**2010**)
- **16.** Asiagwu A. K., Owamah I. H. and Otutu J.O., Kinetic Model for the Sorption of Cu (ii) and Zn (ii) Using Lady Fern (*Athyrium Filix Femina*) Leaf Waste Biomass from Aqueous Solution, *Chemical and Process Engineering Research*, **3**, 1–13 (**2012**)
- **17.** Das S., Usefulness of Pteridophytes in India with Special reference to Medicine and Conservation, *J. Econ. Taxon. Bot.*, **27**, 7–16 (**2003**)
- **18.** Langmuir, The constitution and fundamental properties of solids and liquids., *J. Am. Chem. Soc.*, **38(11)**, 2221–2295 (**1916**)
- **19.** Freundlich H., U" ber die adsorption in lo"sungen, Zeitschrift fu"r Physikalische Chemie, **57**, 385–470, (**1926**)

- 20. Lagergren S., Zur theorie der sogenannten adsorption gelo"ster stoffe. Kungliga Svenska Vetenskapsakademiens Handlingar. Band., 24(4), 1-39 (1898)
- 21. Ho Y.S., Wase D.A., and C.F. Forster, Batch nickel removal from aqueous solution by sphagnum moss peat, *Water Res.*, 29(5), 1327–1332 (1995)
- **22.** Weber W.J. and Morris J.C., Kinetics of adsorption on carbon from solution, *J. Sanit. Eng. Div. Proceed. Am. Soc. Civil Eng.*, **89**, 31–59 (**1963**)
- 23. Uma M.K.N., Ashok V.B., Namdeo J.P., Balu P.K., and Smita S.Z., Equilibrium and Kinetic Studies on Biosorption of Heavy Metals by Leaf Powder of Paper Mulberry (*Broussonetia papyrifer*), Water Air Soil Pollut., 215,177–188 (2011)
- **24.** Gaballah, Goy D., Allain E., Kilbertus G. and Thauront J., Recovery of copper through decontamination of synthetic solutions using modified barks, *Metall. Mater. Trans.*, **28**, 3–23 (**1997**)
- **25.** Diwan S., Biosorption of Copper (II) from aqueous solution by non-living Spirogyra sp., *J. Environ. Res. Develop.*, **1(3)**, 227–231 (**2007**)
- **26.** Zhou D., Zhang L. and Guo S., Mechanisms of lead biosorption on cellulose/chitin beads, *Water Res.*, **39**, 3755–3762 (**2005**)
- **27.** Lata H., Garg V.K. and Gupta R.K., Sequestration of nickel from aqueous solution onto activated carbon prepared from *Parthenium hysterophorus* L., *J. Hazard. Mater.*, **157**, 503–509 (**2008**)
- 28. Dulcy E.C., Gunasekaran S.G. and Dharmendirakumar M., Preparation and Characterization of Chemically Modified Silk Cotton Hull Activated Carbon and Its Effects on Cd(II) Removal from Aqueous Solutions, *Biorem. J.*, 18 (2), 81–92(2014)