



## Studies on $\beta$ -Hemolysin producing *Aeromonas hydrophila* MTCC 646 source Antibacterial activity

Ananth kumar.Y<sup>1\*</sup>, Nirmal Kumar.N<sup>2</sup> and Pothiraj.R<sup>3</sup>

<sup>1</sup>Department of Zoology, St.Xavier's College, Palayamkottai, Tirunelveli, Tamil Nadu, INDIA

<sup>2</sup>Department of Botany, VHNSN College, Virudhunagar, Tamil Nadu, INDIA

<sup>3</sup>Department of Bioengineering, VHNSN College, Virudhunagar, Tamil Nadu, INDIA

Available online at: [www.isca.in](http://www.isca.in), [www.isca.me](http://www.isca.me)

Received 27<sup>th</sup> June 2013, revised 30<sup>th</sup> October 2014, accepted 5<sup>th</sup> January 2015

### Abstract

*Aeromonas hydrophila* MTCC646 was purchased from Institute of microbial Technology (IMTECH) Chandigarh, and it was investigated for the hemolytic, Proteolytic and antibacterial activities. The ability of the *Aeromonas hydrophila* MTCC646 to exhibit the above activities was confirmed by the results of blood agar plates, Muller Hinton Agar (MHA) plates seeded with the bacterial pathogens.

**Keywords:** *Aeromonas hydrophila* MTCC646, antibacterial activity, hemolysin.

### Introduction

*Aeromonas hydrophila* MTCC 646 is a gram-negative, nonsporeforming, rod shaped bacteria in the family of Aeromonadaceae. *Aeromonas hydrophila* can survive in aerobic and anaerobic environments. It causes infections in marine, fresh water and ornamental fish<sup>1,2</sup>. Sea foods have been contaminated (37%) by *Aeromonas* from the west coastal region of southern India<sup>3</sup>.

*Aeromonas hydrophila* was isolated from extra intestinal specimens from the coastal region of Southern Karnataka and it can cause serious infections among immune compromised as well as immune competent individuals<sup>4</sup>. *Aeromonas hydrophila* CRI14 produces siderophores. Although *A. hydrophila* CRI14 produced siderophores, production speed was not in accordance with its growth in fish sera. Simultaneously *A. hydrophila* protease compensates its adequacy of siderophore for iron<sup>5</sup>.

Haemolysin, protease and extracellular polysaccharides (EPS) from pathogenic *Aeromonas hydrophila* An4 shows antibacterial activity against some bacterial pathogens of marine ecosystem. Haemolysins are one the important bacterial virulence factors<sup>6</sup>. Extracellular polysaccharides also play a major role in virulence of pathogenic bacteria and it mediates the interaction between pathogenic bacteria and their environment through adhesion to the host<sup>7</sup>.

Nile tilapia is a popular fresh water aquaculture species in the Philippines. The study focused on the bacteriological examination of Nile tilapia (*Oreochromis niloticus*) during disease outbreaks in various aquaculture farms shows ampicillin resistant *Aeromonas hydrophila*. Consistent isolation of *A. hydrophila* from various tissues of diseased Nile tilapia during disease outbreaks shows a recurring septicemia<sup>8</sup>.

*Aeromonas hydrophila* has been isolated from wounds of five species of brackish water fish including species such as *Platosus anguillaries*, *Lates calcarifer*, *Epinephelus megachie*, *Labeo rohita*, and *Serotherodon nilotica*<sup>9</sup>. Aerolysin binds to specific glycoprotein receptors on the surface of eukaryotic cells before inserting into the lipid bilayer and forming holes. Such aerolysin and haemolysin genes were detected in *A. hydrophila* strains isolated from infected Koi carp (*Cyprinus carpio*)<sup>10</sup>.

The expression of extracellular proteinases and haemolysins under altered atmospheres were studied well and optimal conditions for the expression were determined<sup>11</sup>. More widespread recognition of the prevalence and pathogenic significance of this species have stimulated a number of investigations of the ability of *A. hydrophila* to grown under modern commercial refrigerated storage conditions<sup>12,13</sup>.

Mass mortality in *Misgurnus anguillicaudatus* (Korean cyprinid loach) was caused by *Aeromonas hydrophila*. Additionally it was proved that isolated *Aeromonas hydrophila* containing the *tetE* gene (i.e., Tetracycline resistant gene) exists in Korean aquaculture system and has virulence<sup>14</sup>. In Kolkata the multi-drug resistant *Aeromonas hydrophila* was isolated from surface waters. The *Aeromonas hydrophila* has been examined for their enteropathogenic potential in the sealed-adult -Mouse model using live bacterial cells. Exposure of Mouse intestine to the isolates of *Aeromonas hydrophila* caused epithelial damage of villi associated with architectural distortion of polymorphonuclear cells extending through the mucus<sup>15</sup>.

The activity of some  $\beta$ -lactum antibiotics upon 20 strains of *Aeromonas hydrophila* and some properties of their  $\beta$ -lactamases has been studied. The results show that resistance of

*Aeromonas hydrophila* to  $\beta$ -lactam antibiotics. The high degree of resistance to benzyl penicillin and lower resistance to ampicillin and cephaloridine was observed<sup>16,17</sup>. *Aeromonas hydrophila* was present in faces of buffaloes<sup>18</sup>. *Aeromonas hydrophila* was found to produce haemolytic and proteolytic exotoxin lethal to *Tilapia nilotica*, as well as heat stable unknown virulent factors that were responsible for 20% mortality. The lethality of ECP was decreased by heating and completely inactivated by boiling at 100°C for 10 mins<sup>19</sup>. *Aeromonas hydrophila* was isolated from (Tamilnadu) stool samples of children with acute diarrhea shows multiple antibiotic resistance character.

Some strains of *Aeromonas hydrophila* were isolated from infected skin from common fresh water fish in Bangladesh and tested for enterotoxin production, hemolysin production and proved that strains have resistant to ampicillin<sup>20</sup>. *Aeromonas hydrophila* is also reported as to produce pathogenesis in Gold fish (*Carrassius auratus*) and Koi (*Cyprinus carpio*)<sup>21</sup>.

The formation of extracellular haemolysin by *Aeromonas hydrophila* in relation to protease and staphylolytic enzyme was well studied<sup>22</sup>. Isolation rates of *Aeromonas hydrophila* from stool samples of symptomatic and asymptomatic individuals were examined for several common enteric media. Sheep blood agar with 10 $\mu$ g of ampicillin per ml, preceded by overnight enrichment in alkaline peptone water, yielded 2.6 times the number of isolates as the other media examined for isolation of *Aeromonas hydrophila* from humans<sup>23</sup>. The cytopathogenic response to the  $\alpha$ -hemolysin was reversible, whereas cells treated with small amounts of the  $\beta$ -hemolysin for only 1 minute invariably died within a few hours. Thus, the two hemolysins from *Aeromonas hydrophila*, despite dissimilarities in their interactions with cultured cells<sup>24</sup>.

## Material and Methods

**Source of organism:** *Aeromonas hydrophila* MTCC 646 was purchased from Institute of Microbial Technology (IMTECH), Chandigarh.

**Detection of Proteolytic activity:** *Aeromonas hydrophila* MTCC646 spot was inoculated on casein agar plate and stab inoculated in 10% gelatin agar in test tube. The proteolytic activities were observed in the form of zone clearance and liquefaction after 48 hours of incubation.

**Detection of Hemolytic activity:** Hemolytic activity was isolated by streaking the *Aeromonas hydrophila* MTCC 646 on blood agar base plate supplemented with 7% human blood. The Blood should be aseptically transferred to the media. After sterilizing the media, the blood is added, in order to avoid denaturation.

**Determination of Antimicrobial Activity:** *Aeromonas hydrophila* MTCC 646 was cultured in 100ml Nutrient Broth

(NB) medium at room temperature for 5 days in a rotary shaker. After five days the culture medium was centrifuged at 8000 rpm for 8 minutes and filtrate was screened for antimicrobial activity by agar- diffusion technique on Muller Hinton Agar media (MHA) that was previously seeded with test pathogens. The plates were incubated for 12hrs. Formation of any zone of inhibition was recorded<sup>25</sup>. The test pathogens as follows; *Klebsiella pneumonia*, *Bacillus subtilis*, *Streptococcus aureus*, *Salmonella typhi*, *staphylococcus aureus*, *Pseudomonas Species*

## Results and Discussion

In the proteolytic activity *Aeromonas hydrophila* MTCC 646 was spot inoculated on casein agar media and stab was inoculated in 10% gelatin agar in test tube, does not from clearance of zone and liquefaction. This shows that *Aeromonas hydrophila* MTCC 646 does not produce protease extracellularly. Hemolytic proteins are commonly isolated from pathogenic bacteria and  $\beta$ -hemolysin is one of the important bacterial virulence factors. Hemolysin and its related proteins containing cystathionine  $\beta$  synthase (CBS) domains are bacterial toxins that function by assembling identical subunits into a membrane-spanning pore<sup>26</sup>. It has been suggested that proteolytic enzymes of fish pathogen, *Aeromonas hydrophila*, play an important role in causing massive tissue damage in the host which may facilitate establishment of infection. In Hemolytic activity *Aeromonas hydrophila* strain MTCC 646 interestingly demonstrated a very clear  $\beta$ -hemolysin in the form of clearance zone along the streak on blood agar plate within 24 hours of incubation at room temperature (figure-1). We have demonstrated the hemolytic activity of the *Aeromonas hydrophila* MTCC 646 by using blood agar supplemented with 7% human blood. In antimicrobial activity shows that, filtrate obtained from NB inoculated with *Aeromonas hydrophila* MTCC 646 has antimicrobial activity against some test organisms such as *Klebsiella pneumonia*, *Bacillus subtilis*, *Streptococcus aureus*, *Salmonella typhi*, *staphylococcus aureus*, *Pseudomonas Species*. Attention has been given to the production of haemolysin by *Aeromonas hydrophila*, and it regarded as indication of pathogenic potential, though non-haemolytic aeromonads have also been considered as human pathogen<sup>27</sup>. We have investigated that the *Aeromonas hydrophila* MTCC 646 for proteolytic activity. The results show that the strain *Aeromonas hydrophila* MTCC 646 does not exhibit proteolytic activity. In previous study<sup>6</sup> shows that *Aeromonas hydrophila* An4 have antibacterial activity against marine bacterial species.

## Conclusion

The present study concluded that, the *Aeromonas hydrophila* MTCC 646 shows antibacterial activity against some bacterial pathogens. It suggested that further studies are required to determine the antimicrobial activity of *Aeromonas hydrophila* MTCC 646.

## Reference

- Ramasamy harikrishnan, Chellam Balasundaram, Manchul kim, Ju-sang kim and Moon- Soo Heo., Effective administration route of Azadirachitin and its impact on hematological and Biochemical parameters in gold fish (*carassius Auratus*) infected with *Aeromonas hydrophila*, *Bull vet Inst pulawy*, 613-619 (2009)
- Alagappan K.M., Devasigamani B., Kumaran S. and Sakthivel K., Histopathological alterations in Estuarine catfish (*Arius maculatus*; Thunberg,1792) due to *Aeromonas hydrophila* infection, *World J. of Fish and Marine Sciences*, **1(3)**, 185-189 (2009)
- Thayumanavan T., Vivekanandhan G., Savithamani K., Subash Kumar R. and Lakshmana Perumalsamy P., Incidence of haemolysin positive and drug-resistant *Aeromonas hydrophila* in Freshly Caught finfish and prawn Collected from major commercial fish of coastal south India, *FEMS Immunol Med Microbiol*, **36**, 41-45 (2003)
- Mukukhopadhyay C. and Chawla K. Bairy, Emerging extra: Intestinal infections with *Aeromonas hydrophila* in coastal region of Southern Karnataka, *J post grad Med.*, **3**, 199-202 (2008)
- Tang F., Zhu X., Long H. and Zeng Y., The role of *Aeromonas hydrophila* protease in the utilization of fish Serum Iron invitro, *Asian Fisheries Science*, **10**, 317-321 (1998)
- Anju pandey, Milind Naik and Santosh Kumar Dubey, Hemolysin, Protease and EPS Producing pathogenic *Aeromonas hydrophila* Strain An4 Shows antibacterial activity against Marine Bacterial fish Prathogens, *Journal of Marine Biology*, Hidawi Publishing Corporation, Article ID563205 (2010)
- Vuong C., Kocianova S. and Voyich J.M., A Crucial role for EPS modification in Bacterial Biofilm formation, Immune evasion, and Virulence, *The Journal of Biological chemistry*, **279(52)**, 54881-54886 (2004)
- Yambot A.V., Isolation of *Aeromonas hydrophila* from philippines *Oreochromis niloticus* during fish disease out breaks in Philippines, *Asian Fisheries science*, **10**, 347-354 (1998)
- Rahim Z., Aziz K.M.S., Huq M.I. and Saeed H., Isolation of *Aeromonas hydrophila* from the Wounds of Five Species of brackish water fish of Bangladesh, *Bangladesh J. Zoo.*, **13**, 37-42 (1985)
- Uma A., Rebecca G., Meena S. and Saravanabava K., PCR detection of putative Aerolysin and haemolysin genes in an *Aeromonas hydrophila* isolate from infected koi carp (*Cyprinus carpio*), *Tamilnadu J. veterinary and Animal Sciences*, **6(1)**, 31-33 (2010)
- McMahon M.A.S., The expression of proteinases and hemolysins by *Aeromonas hydrophila* under modified atmospheres, *Journal of Applied Microbiology*, **89**, 415-422 (2000)
- Hudson J.A. and Avery S.M., Growth of *Listeria Monocytogenes*, *Aeromonas hydrophila* and *Yersinia enterocolitica* on cooked muscle tissue and refrigeration and mild temperature abuse, *Journal of Food Safety*, **14**, 41-52 (1994)
- Davies A.R., and Slade A., Fate of *Aeromonas* and *Yersinia* on modified atmosphere packaged (MAP) cod and trout, *Letters in Applied Microbiology*, **21**, 354-358 (1995)
- Jin Woo Jun, Ji Hyung Kim, Casiano H and Se chang Park, Occurrence of tetracycline: Resistant *Aeromonas hydrophila* infection in Korean Cyprinid loach (*Misgurnus anguillicaudatus*), *African Journal of Microbiology Research*, **4(9)**, 849-855 (2010)
- Rathinasamy Subash Kumar, Thangavelu Thayumanavan, Govindasamy Vivekanandhan and Perumalsamy Lakshmana Perumalsamy, Occurrence of *Aeromonas hydrophila* in acute gastroenteritis among Children, *Indian J.Med. Res.*, 123, 61-66 (2006)
- Raul Zemelman, Carlos Gonzalez, Maria A, Mondaca Juan Silva, Carlos Merino and Mariana Dominguez, Resistance of *Aeromonas hydrophila* to B- Lactum antibiotics, *Journal of Antimicrobial chemotherapy*, **14**, 575-579 (1984)
- Andrea Belem Costa and Jose Eurico Possebon Cyrino, Antibiotic resistance of *Aeromonas hydrophila* isolated from *Piaractus Mesopotamicus* (Holmberg, 1887) and *Oreochromis niloticus* (Linnaeus, 1758), *Sci. Agric. (Piracicaba Braz)*, **63**, 281-284 (2006)
- Rahimi Larki E. and Nane S.S., The prevalence of *Aeromonas hydrophila* induced diarrhea in the Pig, Buffalo and human in Pune area, India, *Iranian Journal of Veterinary Research*, **7**, 53-58 (2006)
- Khalil A.H. and Mansour E.H., Toxicity of crude extracellular products of *Aeromonas hydrophila* in Tilapia, *Tilapia nilotica*. *Letters in Applied Microbiology*, **25**, 269-273 (1997)
- Zaur Rahim, Sanyal S.C., Aziz K.M.S., Huq M.I. and Chowdhury A.A., Isolation of enterotoxigenic, hemolytic, and antibiotic resistant *Aeromonas hydrophila* Strains from infected fish in Bangladesh, *Applied and Environmental Microbiology*, **48**, 865-867 (1984)
- Citarasu T., Alfred Dhas K., Velmurugan S., Thanga Viji V., Kumaran T., Michael Babu M. and Selvaraj T., Isolation of *Aeromonas hydrophila* from infected ornamental fish Hatchery during massive disease outbreak, *International Journal of Current Research*, **2**, 37-41(2011)

22. Wrekling B. and Heden L., Formation of extracellular hemolysin by *Aeromonas hydrophila* in relation to Proteases and Staphylolytic enzyme, *Journal of General Microbiology*, **78**, 57-65 (1973)
23. Bradford A., Kay Cesar E., Guerrero Bradley and Sack R., Media for isolation of *Aeromonas hydrophila*. *Journal of Clinical Microbiology*, **22**, 888-890 (1985)
24. Monica Thelestam and Asa Ljungh, Membrane damaging and cytotoxic effects on human fibroblasts of Alpha and Beta: Hemolysins from *Aeromonas hydrophila*, *Infection and Immunity*, **34**, 949-955 (1981)
25. Sudipta Roy and Debdulal Banerjee, Isolation of antimicrobial compound by endoplytic bacteria from *vinca rosea*, *International Journal of Current Reseach*, ISSN:0975-833X, 047-051 (2010)
26. Erova T.E., Sha J., Horneman A.J., Identification of a new hemolysin from diarrheal isolate SSU of *Aeromonas hydrophila*, *FEMS Microbiology Letters*, **275(2)**, 301–311 (2007)
27. Namdari H. and Bottone E.J., Cytotoxin and enterotoxin production as factors delineating enteropathogaen city of *Aerimonas caviae.*, *J. Clin. Microbiol.*, **28** 1796-1798 (1990)