



## Review Paper

# Plants: Green Route for Nanoparticle Synthesis

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## Abstract

The synthesis of nanoparticles has become the matter of great interest in recent times due to its various advantageous properties and applications in various fields. Though physical and chemical methods are more popular for nanoparticle synthesis, the biogenic production is a better option due to eco-friendliness. This review reports the potential of plants i.e. "green chemistry" to synthesize nanoparticles not only in the laboratory scale but also in their natural environment. Furthermore, factors affecting biosynthesis along with current and future applications are also discussed.

**Keywords:** Nanoparticles, plants, biosynthesis, applications.

## Introduction

Nanoparticle having one or more dimensions of the order of 100nm or less- have attracted considerable attraction due to their unusual and fascinating properties, with various applications, over their bulk counterparts<sup>1,2</sup>. Currently, a large number of physical, chemical, biological, and hybrid methods are available to synthesize different types of nanoparticles<sup>3-6</sup>. Though physical and chemical methods are more popular for nanoparticle synthesis, the use of toxic compounds limits their applications. The development of safe eco-friendly methods for biogenetic production is now of more interest due to simplicity of the procedures and versatility<sup>7,8</sup>. Due to their amenability to biological functionalization, the biological nanoparticles are finding important applications in the field of medicine<sup>9</sup>. The antimicrobial potential of metal based nanoparticles has led to its incorporation in consumer, health-related and industrial products<sup>10</sup>.

## Plants-the Green route for biosynthesis of nanoparticles

Nature has devised various processes for the synthesis of nano- and micro- length scaled inorganic materials which have contributed to the development of relatively new and largely unexplored area of research based on the biosynthesis of nanomaterials. Synthesis using bio-organisms is compatible with the green chemistry principles. "Green synthesis" of nanoparticles makes use of environmental friendly, non-toxic and safe reagents. Figure 1 shows the general biosynthesis of metal nanoparticles from biological sources<sup>11-13</sup>.

Phytomining is the use of hyper accumulating plants to extract a metal from soil with recovery of the metal from the biomass to return an economic profit<sup>14</sup>. Hyper accumulator species have a physiological mechanism that regulates the soil solution concentration of metals. Exudates of metal chelates from root system, for example, will allow increased flux of soluble metal

complexes through the root membranes<sup>15</sup>. It has been observed that stress tolerant plants have more capacity to reduce metal ions to the metal nanoparticles<sup>16</sup>. Mechanism of biosynthesis of nanoparticles in plants may be associated with phytoremediation concept in plants<sup>17-19</sup>. Biosilicification also results in nanoparticles in some higher plants as shown in figure 2<sup>20</sup>.

**Factors affecting biosynthesis of nanoparticles:** Temperature plays an important role to control the aspect ratio and relative amounts of gold nanotriangles and spherical nanoparticles. Temperature variations in reaction conditions results in fine tuning of the shape, size and optical properties of the anisotropic nanoparticles<sup>21</sup>. More than 90% of leaf extracts of two plants- *Magnolia kobus* and *Diopyros kaki* was converted to gold nanoparticles at a reaction temperature of 95 °C in few minutes, suggesting reaction rates higher or comparable to those of nanoparticle synthesis by chemical methods<sup>22</sup>. The size of gold nanoparticles was shown to increase at higher reaction temperatures as explained by an increase in fusion efficiency of micelles which dissipates supersaturation<sup>23</sup>.

pH of the medium influence the size of nanoparticles at great concern. For example, the size of gold nanoparticles was controlled by altering the pH of the medium in *Avena sativa*<sup>24</sup>. The reaction mechanism for the formation of magnetite nano particles have been found to be influenced by pH when coprecipitation method was followed<sup>25</sup>.

Other than pH and temperature other factors also play role in nanoparticle synthesis. The size and crystallinity of magnetite nanoparticles was found to increase with increasing molar ratios of ferric/ferrous ions during synthesis by hydrothermal synthesis method according to the Schikorr reaction<sup>26</sup>. The band gap energy was found to decrease with increase in dopant concentration in ZnS samples as determined by optical absorption spectroscopic technique<sup>27</sup>. The sizes of gold nanoparticles decreases with increasing NaCl concentrations

(size ranges, 5-16 nm) than those synthesized without addition of NaCl (size ranges 11-32 nm)<sup>28</sup>. Chloride, bromide and iodide affect nanoparticle formation in plants. Chloride promotes growth of nanotriangles while iodide causes distraction in nanotriangle morphology and induces formation of aggregated spherical nanoparticles<sup>29</sup>. Chloride ion results in the formation of diamond-shaped copper nanoparticles<sup>30</sup>. Sun-dried biomass of *Cinnamomum camphora* leaf when incubated with

aqueous silver or gold precursors at ambient temperature produces both silver nanoparticles (55–80 nm) and triangular or spherical gold nanoparticles. The marked difference in shape of gold and silver nanoparticles could be attributed to the comparative potential of protective and reductive biomolecules from leaf extracts. The polyol and water-soluble heterocyclic components were mainly responsible for the reduction of silver ions or chloroaurate ions<sup>31</sup>.

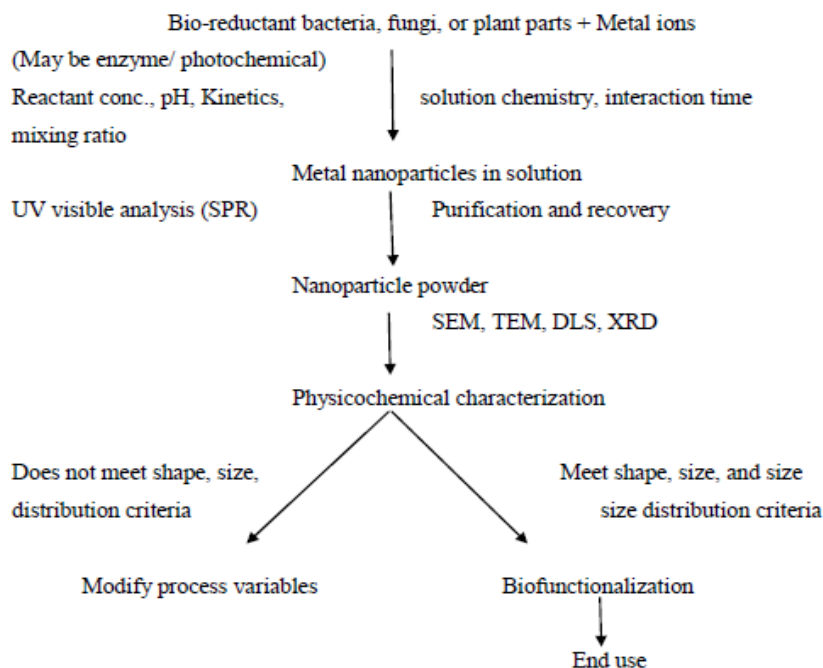


Figure-1  
 Generalized flow chart for nanobiosynthesis

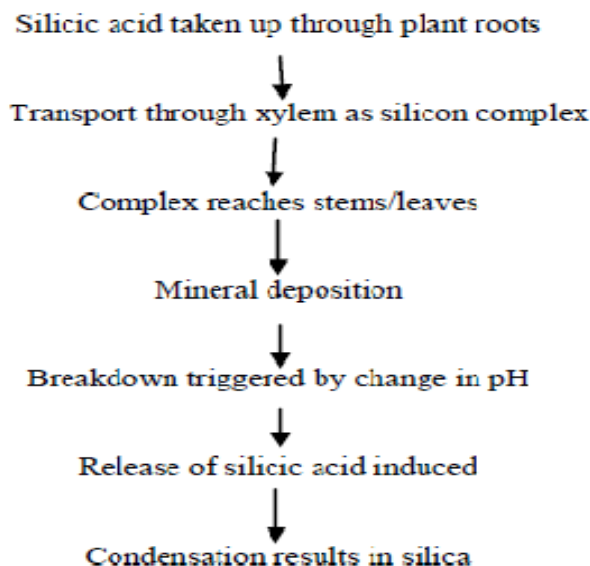


Figure-2  
 Flowchart for biosilicification process

## Nanoparticles synthesized by plants

Nanoparticle biosynthesis is an eco-friendly approach for recovering metals, for example gold, which are catalytically active for reactions and selective oxidation of CO<sup>32,33</sup>. Synthesis of mixed nanoparticles alloys having various technological applications and catalysts of specific composition that can't be produced by traditional methods can also be possible. Nanoparticles can be applied in sensors<sup>34, 35</sup> and medicine i.e. nanomedicine<sup>36</sup>.

## Si-Ge-O nanocomposite by diatoms

Fabrication of Silicon-Germanium nanoparticles have been reported from freshwater diatom *Stauroneis sp.*<sup>37</sup>. Silica from cells of *Nitzschia frustulum*, which possess blue photoluminescence have been cultured in bioreactor whose intensity and wavelength depends on the change in frustules nanostructure<sup>38</sup>.

## Silver nanoparticles

Silver nanoparticles have attracted intensive research interest because of their important applications in antimicrobial, catalysis, and surface-enhanced Raman scattering<sup>39-41</sup>. For centuries, silver has been used as an antimicrobial agent. The recent resurgence in interest for this element particularly focuses on the increasing threat of antibiotic resistance, caused by the abuse of antibiotics<sup>42,43</sup>.

There are several hypotheses to explain the antibacterial activity of silver nanoparticles. The rapid breakdown of silver nanoparticles releases ionic silver that inactivates vital bacterial enzymes by interacting with essential thiol groups. Silver ions can inhibit bacterial DNA replication, damage bacterial cytoplasm membranes, depleting levels of intracellular adenosine triphosphate (ATP) and finally cause cell death<sup>44</sup>. The high specific surface-to-volume ratio of silver nanoparticles increases their contact with microorganisms, promoting the dissolution of silver ions, thereby improving biocidal effectiveness. The ability of silver nanoparticles to release silver ions is a key to their bactericidal activity<sup>45</sup>. Silver nanoparticles can be synthesized in a number of ways, the borohydride reduction of silver salts being the most common. Stabilization is achieved using capping agents that bind to the nanoparticle surface and improve stability and water solubility, which are essential to prevent aggregation; examples include water-soluble polymers, oligosaccharides and polysaccharides, sodium dodecyl sulphate (SDS) and sophorolipid (glycolipid)<sup>46</sup>.

Silver nanoparticles of 20-30 nm from leaves of *Acalypha indica* showed antimicrobial activity against *E. coli* and *Vibrio cholera*<sup>47</sup> while silver nanoparticles of 3-12 nm from peels of *Citrus sinensis* have been reported to show activity against *Bacillus subtilis*<sup>48</sup>. Particles of size 33.67 nm from *Allium cepa* stem show antimicrobial activity against *E. coli* and *S. typhimurium*<sup>49</sup>. Silver nanoparticles of size 8 nm from leaves of

*Nicotiana tabacum* inhibits *Pseudomonas putida*, *P. vulgaris*, *Escherichia coli* DH5 $\alpha$ , *B. subtilis*, *P. aeruginosa* and *Salmonella typhi*<sup>50</sup>.

## Gold nanoparticles

Au particles are particularly and extensively exploited in organisms because of their biocompatibility<sup>51</sup>. Gold nanoparticles (Au) generally are considered to be biologically inert but can be engineered to possess chemical or photo thermal functionality. On near infrared (NIR) irradiation the Au-based nanomaterials, Au nanospheres, Au nanocages, and Au nanorods with characteristic NIR absorption can destroy cancer cells and bacteria via photo thermal heating. Au-based nanoparticles can be combined with photo sensitizers for photodynamic antimicrobial chemotherapy. Au nanorods conjugated with photo sensitizers can kill MRSA by photodynamic antimicrobial chemotherapy and NIR photo thermal radiation<sup>52, 53</sup>.

Aggregated forms of nanoparticles like gold nanotriangles have been reported in lemon grass extracts and tamarind leaf extracts<sup>54</sup> and dead biomass of *Humulus lupulus* also produces gold nanoparticles<sup>55</sup>. Extra cellular synthesis of gold nanoparticles has been observed using *Emblica officinalis* fruit extract as a reducing agent.

## Platinum nanoparticles

Nanoparticles ranging from 2-12 nm was the first to be reported in platinum and was synthesized using >10% *Diopyros kaki* leaf extract as reducing agent from an aqueous H(2)PtCl(6).6H(2)O solution at a reaction temperature of 95<sup>0</sup>C<sup>56</sup>. Platinum nanoparticles of 23 nm size have been prepared using leaf extract of *Ocimum sanctum* as reducing agent from aqueous chloroplatinic acid at a reaction temperature of 100<sup>0</sup>C that finds application in water electrolysis<sup>57</sup>.

## Zinc nanoparticles

To the best of our knowledge, biological approach using milky latex of *Calotropis procera* has been used for the first time as a reducing material as well as surface stabilizing agent for the synthesis of spherical-shaped ZnO-NPs. The structure, phase, and morphology of synthesized product were investigated by the standard characterization techniques. Milky latex of *Calotropis procera* has been used for the synthesis of spherical ZnO NPs. Highly stable and spherical ZnO NPs have also been synthesized using *Aloe vera* extract<sup>59</sup>.

## Conclusion

The "green" route for nanoparticle synthesis is of great interest due to eco-friendliness, economic prospects, feasibility and wide range of applications in nanomedicine, catalysis medicine, nano-optoelectronics, etc. It is a new and emerging area of research in the scientific world, where day-by-day developments is noted in warranting a bright future for this field.

## References

1. Daniel M.C. and Astruc D., Gold nanoparticles: assembly, supramolecular chemistry, quantum-size-related properties, and applications toward biology, catalysis, and nanotechnology, *Chem Rev.*, **104**, 293–346 (2004)
2. Kato H., In vitro assays: tracking nanoparticles inside cells, *Nat Nanotechnol.*, **6**, 139–140 (2011)
3. Liu J., Qiao S.Z., H, Q.H. and Lu G.Q., Magnetic nanocomposites with mesoporous structures: synthesis and applications, *Small.*, **7**, 425–443 (2011)
4. Grass L.R.N., Athanassiou E.K. and Stark W.J., Bottom-up fabrication of metal/metal nanocomposites from nanoparticles of immiscible metals, *Chem Mater.*, **22**, 155–160 (2010)
5. Tiwari D.K., Behari J. and Sen P., Time and dose-dependent antimicrobial potential of Ag nanoparticles synthesized by top-down approach, *Curr Sc.i.*, **95**, 647–655 (2008)
6. Mohanpuria P., Rana N.K. and Yadav S.K., Biosynthesis of nanoparticles: technological concepts and future applications, *J Nanopart Res.*, **10**, 507–517 (2008)
7. Li X., Xu H., Chen Z., Chen G., Biosynthesis of nanoparticles by microorganisms and their applications, *J Nanomater.*, **2011**, 1-16 (2011)
8. Popescu M., Velea A. and Lorinczi A., Biogenic production of nanoparticles, *Dig J Nanomater Bios.*, **5**, 1035 – 1040 (2010)
9. Dushenkov V., Kumar P.B.A.N., Motto H. and Raskin I., Rhizofiltration: the use of plants to remove heavy metals from aqueous streams, *Environ Sci Technol.*, **29**, 1239-1245 (1995)
10. Dibrov P., Dzioba J., Gosink K.K. and Hase C.C., Chemiosmotic mechanism of antimicrobial activity of Ag<sup>+</sup> in *Vibrio cholera*, *Antimicrob, Agents Chemother.*, **46**, 2668-2670 (2002)
11. Li S., Qui L., Shen Y., Xie A., Yu X., Zhang L. and Zhang Q., Green synthesis of silver nanoparticles using *Capsicum annum* L. extract, *Green Chem.*, **9**, 852-858 (2007)
12. Sharma V.K., Yngard R.A. and Lin Y., Silver nanoparticles: Green synthesis and their antimicrobial activities, *Adv. Collo. Interf. Sci.*, **145**, 83–96 (2009)
13. Prathna T.C., Mathew L., Chandrasekaran N., Raichur AM., Mukherjee A., Biomimetic Synthesis of Nanoparticles: Science, Technology and Applicability, Edited A. Mukherjee, InTech Publishers, Croatia 1-20 (2010)
14. Lamb A.E., Anderson C.W.N. and Haverkamp R.G., The extraction of gold from plants and its applications to phytomining, *Chem New Zealand*, **65**, 31-33 (2001)
15. Vedpriya A., Living systems: eco-friendly nanofactories, *Dig J Nanomater Bios.*, **5**, 9 – 21 (2010)
16. Ankamwar B., Damle C., Ahmad A., Sastry M., Biosynthesis of gold and silver nanoparticles using *Emblica officinalis* fruit extract, their phase transfer and transmetallation in an organic solution, *J Nanosci Nanotechnol.*, **5**, 1665–1671 (2005)
17. Anderson C.W.N., Brooks R.R., Stewart R.B. and Simcock R., Induced hyperaccumulation of gold in plants, *Nature.*, **395**, 553-554 (1998)
18. Huang J.W. and Cunningham S.D., Lead phytoextraction: species variation in Lead uptake and translocation, *New Phytol.*, **134**, 75-84 (1996)
19. Haverkamp R.G., Marshall A.T. and Agterveld D.V., Pick your Carats: Nanoparticles of gold–silver–copper alloy produced In Vivo, *J Nanopart Res.*, **9**, 697-700 (2007)
20. Lopez P.J., Gautier C., Livage J. and Coradin T., Mimicking biogenic silica nanostructures formation, *Curr. Nanosci.*, **1**, 73-83 (2005)
21. Armendariz V., Herrera I., Peralta- Videa J.R., Jose Yacaman M., Troiani H., Santiago P. and Gardea-Torresdey J.L., Size controlled gold nanoparticle formation by *Avena sativa* biomass: use of plants in nanobiotechnology, *J Nanopart Res.*, **6**, 377–82 (2004)
22. Song JY., Jang H.K. and Kim B.S., Biological synthesis of gold nanoparticles using *Magnolia kobus* and *Diospyros kaki* leaf extracts, *Process Biochem*, **44**, 133-1138 (2009)
23. Muralidharan G., Subramanian L., Nallamuthu S.K., Santhanam V. and Sanjeev Kumar., Effect of reagent addition rate and temperature on synthesis of gold nanoparticles in microemulsion route, *Ind. Eng. Chem. Res.*, **50**, 8786-8791 (2011)
24. Shanker S. S., Bhargava S. and Sastry M., Synthesis of gold nanospheres and nanotriangles by the Turkevich approach, *J. Nanosc. Nanotechnol.*, **5**, 1721-1727 (2005)
25. Faiyas A.P.A., Vinod EM., Joseph J., Ganesan R. and Pandey R.K. Dependence of pH and surfactant effect in the synthesis of magnetite (Fe<sub>3</sub>O<sub>4</sub>) nanoparticles and its properties, *J Magn Magn Mater.*, **322**, 400-404 (2010)
26. Mizutani N., Iwasaki T., Watano S., Yanagida T., Tanaka H. and Kawai T., Effect of ferrous/ferric ions molar ratio on reaction mechanism for hydrothermal synthesis of magnetite nanoparticles, *Bull. Mater. Sci.*, **31**, 713–717 (2008)
27. Brightson M., Selvarajan P., John Kennady V., Freeda T.H. and Meenakshi Sundar S., Investigations on the effect of manganese ions on the structural and optical properties of ZnS nanoparticles synthesized by solvo-thermal route, *Recent Res Sci and Technol.*, **2**, 29-33 (2010)

28. Mohamad M. F., Kamarudin K.S.N., Fathilah N. N.F.N.M. and Salleh M.M., The Effects of Sodium Chloride in the Formation of Size and Shape of Gold (Au) Nanoparticles by microwave-polyol method for mercury adsorption, *World Acad. Sci. Eng. Technol.*, **74**, 691-695 (2011)
29. Bai H.J., Zhang Z.M. and Gong J., Biological synthesis of semiconductor zinc sulfide nanoparticles by immobilized *Rhodobacter sphaeroides*, *Biotechnol. Lett.*, **28**, 1135 – 1139 (2006)
30. Kitchens C.L., McLeod M.C. and Roberts C.B., Chloride ion effects on synthesis and directed assembly of copper nanoparticles in liquid and compressed alkane microemulsions, *Langmuir*, **21**, 5166-73 (2005)
31. Huang J, LiQ, Sun D, Lu Y, Su Y, Yang X, et al, Biosynthesis of silver and gold nanoparticles by novel sundried *Cinnamomum camphora* leaf, *Nanotechnol.*, **18**, 105104–105114 (2007)
32. Andreeva D., Low temperature water gas shift over gold catalysts, *Gold Bull.*, **35**, 82 -88 (2002)
33. Grisel R.J., Weststrate K.J., Gluhoi A. and Nieuwenhuys B. E., catalysis by gold nanoparticles, *Gold Bull.*, **35**, 39-45 (2002)
34. Yanez-Sedeno P. J. and Pingarron M., Gold nanoparticle-based electrochemical biosensors, *Anal. Bioanal. Chem.*, **382**, 884-886 (2005)
35. Liu J. and Yi L., Colorimetric Biosensors Based on DNzyme-Assembled Gold Nanoparticles, *J. Fluorescence*, **14**, 343-354 (2004)
36. Paciotti G.F., Myer L., Weinreich V., Goia D., Pavel N., McLaughlin R. E. and Tamarkin L., Colloidal gold: a novel nanoparticle vector for tumor directed drug delivery, *Drug Delivery*, **11**, 169-183 (2009)
37. Mubarak Ali D., Divya C., Gunasekaran M. and Thajuddin N., Biosynthesis and characterization of Silicon – Germanium oxide nanocomposite by Diatom, *Dig J Nanomater Bios.*, **6**, 117 – 120 (2011)
38. Jeffryes C., Gutu T., Jiao, J. and Rorrer G.L. Two-stage photobioreactor process for the metabolic insertion of nanostructured germanium into the silica microstructure of the *Diatom* Pinnularia sp, *Mater. Sci. Eng C: Biomimetic Supramol Syst.*, **28**, 107-118 (2008)
39. Li Z., Lee D., Sheng X. X., Cohen R. E. and Rubner M. F., Two-level antibacterial coating with both release-killing and contact-killing capabilities, *Langmuir.*, **22**, 9820-9823 (2006)
40. Chen Y.Y., Wang C.A., Liu H.Y., Qiu J.S. and Bao X.H., Ag/SiO<sub>2</sub>: A novel catalyst with high activity and selectivity for hydrogenation of chloronitrobenzenes, *Chem. Commun.*, **42**, 5298-5300 (2005)
41. Setua P., Chakraborty A., Seth D., Bhatta M.U., Satyam P.V. and Sarkar N., Synthesis, optical properties, and surface enhanced Raman scattering of silver nanoparticles in nonaqueous methanol reverse micelles, *J. Phys. Chem. C.*, **111**, 3901-3907 (2007)
42. Panaek A., Kvitek L., Pucek R., Kolar M., Veerova R., Pizurova N., Sharma V. K., Nevena T. and Zboril R., Silver colloid nanoparticles: Synthesis, characterization and their antibacterial activity, *J. Phys. Chem. B.*, **110**, 16248–16253 (2006)
43. Sandbhy V., MacBride M.M., Peterson B.R.. and Sen A., Silver bromide nanoparticles/polymer composites: dual action tunable antimicrobial materials, *J. Am. Chem. Soc.*, **128**, 9798-9808 (2006)
44. Feng Q.L., Wu J., Chen G.Q., Cui F.Z., Kim T.N. and Kim J. O., A mechanistic study of the antibacterial effect of silver ions on *Escherichia coli* and *Staphylococcus aureus*, *J. Biomed. Mater. Res.*, **52**, 662-668 (2000)
45. Stobie N., Duffy B., McCormack D.E., Colreavy J., Hidalgo M. and McHale P., Prevention of *Staphylococcus epidermidis* biofilm formation using a low –temperature processed silver doped phenyltriethoxysilane solgel coating, *Biomater*, **29**, 963-969 (2008)
46. Sing S., Patel P., Jaiswal S., Prabhune A.A., Ramana C.V. and Prasad B.L.V., A direct method for the preparation of glycolipid-metal nanoparticle conjugates: sophorolipids as reducing and capping agents for the synthesis of water re-dispersible silver nanoparticles and their antibacterial activity, *New J. Chem.*, **33**, 646-652 (2009)
47. Krishnaraj C., Jagan E.G., Rajasekar S., Selvakumar P., Kalaichelvan P.T. and Mohan N., Synthesis of silver nanoparticles using *Acalypha indica* leaf extracts and its antibacterial activity against water borne pathogens, *Coll Surf B: Biointer.*, **76**, 50-56 (2010)
48. Konwarh R., Gogoi B., Philip R., Laskar M.A. and Karak N., preparation of polymer-supported free radical scavenging, cytocompatible and antimicrobial green silver nanoparticles using aqueous extract of *Citrus sinensis* peel, *Colloids Surf. B: Biointerfaces.*, **84**, 338-345 (2011)
49. Saxena A., Tripathi R.M. and Singh R.P., Biological synthesis of silver nanoparticles by using onion (*Allium cepa*) extract and their antibacterial activity, *Dig J Nanomater. Bios.*, **5**, 427-432 (2010)
50. Suranjit K., Pathak D., Patel A, Dalwadi P., Prasad R., Patel P. and Selvaraj K., Biogenic synthesis of silver nanoparticles using *Nicotiana tobaccum* leaf extract and study of their antibacterial effect, *African J Biotech.*, **10**, 8122-8130 (2011)
51. Bhattacharya R. and Mukherjee P., Biological properties of “naked” metal nanoparticles *Advanced Drug Delivery Rev.*, **60**, 1289-1306 (2008)

52. Kuo W.S., Chang C.N., Chang Y.T. and Yeh C.S., Antimicrobial gold nanorods with dual-modality photodynamic inactivation and hyperthermia, *Chem. Commun. Camb.*, **32**, 4853–4855 (2009)
53. Pissuwan D., Cortie C.H., Valenzuela S.M. and Cortie M.B., Functionalised gold nanoparticles for controlling pathogenic bacteria, *Trends in Biotechnol.*, **28**, 207-213 (2009)
54. Ankamwar B., Chaudhary M. and Sastry M., Gold nanotriangles biologically synthesized using tamarind leaf extract and potential application in vapor sensing., *Synth React Inorg Metal-Org Nano-Metal Chem.*, **35**, 19–26 (2005)
55. Lopez M.L., Parsons J.G., Peralta-Videa J.R. and Gardea-Torresdey J.L., A XAS study of the binding and reduction of Au(III) by hops biomass, *Microchem Journal*, **81**, 50-56 (2005)
56. Song J.Y., Kwon E.Y. and Kim B.S., Biological synthesis of platinum nanoparticles using *Diopyros kaki* leaf extract, *Bioprocess Biosyst Eng.*, **33**, 159-64 (2010)
57. Soundarrajan C., Sankari A., Dhandapani P., Maruthamuthu S., Ravichandran S., Sozhan G. and Palaniswamy N., Rapid biological synthesis of platinum nanoparticles using *Ocimum sanctum* for water electrolysis applications, *Bioproc Biosys Eng.*, **35**, 827-833 (2012)
58. Singh R.P., Shukla V.K., Yadav R.S., Sharma P.K., Singh P.K., Pandey A.C., Biological approach of zinc oxide nanoparticles formation and its characterization, *Adv. Mat. Lett.*, **2**, 313-317 (2011)
59. Sangeetha G., Rajeshwari S. and Venckatesh R., Green synthesis of zinc oxide nanoparticles by *Aloe barbadensis miller* leaf extract: Structure and optical properties, *Mater Res Bull.*, **46**, 2560–2566 (2011)